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Electricity generation using *p*-nitrophenol as substrate in microbial fuel cell

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ABSTRACT

The cell voltage and degradation rate of *p*-nitrophenol (PNP) were monitored in a two-chambered microbial fuel cell (MFC) system. Degradation metabolites in the anode solution of MFC were analyzed by gas chromatography—mass spectrometry (GC—MS). PNP was used as substrate by the MFC that was inoculated with anaerobic sludge. The results showed that electricity output increased with the PNP concentration increased, the MFC displayed a maximum power density of 1.778 mW m⁻² and a maximum PNP degradation rate of 64.69% when PNP was used as a sole substrate. However, the cell voltage and the PNP degradation rate with sodium acetate (402.3 mV and 95.96%) were higher than those fed with glucose (341.9 mV and 83.51%) when glucose and sodium acetate were used as a substrate, respectively. Furthermore, GC—MS analysis showed that the PNP was biodegraded completely after 142 h in the MFC. These results demonstrate that PNP can be used for electricity generation in MFC for practical applications of wastewater treatment.

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1. Introduction

p-Nitrophenol (PNP) is an important nitro-aromatic compound that is used in manufacturing of dyes, pigments, explosives, fungicides, industrial solvent and organic phosphorus pesticide (Paola et al., 2003; Kuosa and Kallas, 2009). Due to its high chemical stability and toxicity to organisms, PNP is recognized as the European Economic Community list I of toxic pollutant and a priority pollutant by the US Environmental Protection Agency, and its concentration in natural water is restricted to less than 10 mg l⁻¹ for environmental safety (Zhang et al., 2009).

The microbial fuel cell (MFC) is an electrochemical device that uses microorganisms as the catalysts to oxidize organic matter and convert chemical energy to electrical energy (Logan, 2005; leropoulos et al., 2005; Aelterman et al., 2006; Liu et al., 2009; Sun et al., 2010). Most natural substrate remains in the form of glucose, oxalate, butyrate, and other easily degradable substrates (Min and Logan, 2004; Liu et al., 2005; Wen et al., 2009). Some toxic organics can also be used as substrate in the MFC. In this study, the cell voltage and the removal of PNP in the two-chambered MFC system were investigated.

2. Materials and methods

2.1. MFC configuration

A two-chambered microbial fuel cell system was designed and constructed. Each chamber (anode or cathode chambers) was filled with 2 l of medium solution, and the two chambers were connected with a glass tube (total length, 8 cm; inner diameter, 4 cm) with two pieces of organic microporous filter membranes (pore size, 0.22 μ m) in the middle of the tube. The electrodes were made of kryptol (surface area, 100 cm²), and connected via a 1900 Ω external resistor with copper wire.

2.2. MFC operation

The MFC was started using anaerobic sludge (500 ml) in the anode chamber and phosphate buffer solution (4.22 g l^{-1} NaH₂PO₄ + 2.75 g l^{-1} Na₂HPO₄, pH, 7) in the cathode chamber at room temperature (25 ± 5 °C). The cathode chamber was continuously aerated by an aquarium membrane pump.

Substrates used in the anode chamber of MFC included PNP, PNP and glucose (G), PNP and sodium acetate (NaAC). The MFC was operated with 500 mg l^{-1} G as the fuel at the beginning. After electrical output was reached a steady state, the MFC was operated sequentially using PNP, PNP and G, PNP and NaAC as the substrates. Besides the substrates, the anodic solution also contained (per liter of deionized water): 0.31 g NH₄Cl, 0.13 g KCl, 4.97 g NaH₂PO₄, 2.75 g

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Na₂HPO₄, trace metal solution 12.5 ml, a vitamin stock solution 12.5 ml and trace metal solution 12.5 ml as described in a previous study (Lovely and Phillips, 1988).

In all operations, once the voltage outputs were below 50 mV, the substrate was replaced with a new solution for the next cycle. The voltage was measured hourly, COD and PNP concentrations of the anode chamber were analyzed daily.

2.3. Analysis and calculate

COD was analyzed by the potassium dichromate method (Huang, 1996). The concentration of PNP in the anode chamber was analyzed from the absorption intensities of species obtained spectrophotometrically (Shimadzu Kyoto, Japan) at a wavelength of 318 nm and a pH of 3–4 (Guo and Deng, 1998). GC–MS (Thermo Finnigam-TRACE GC, Polaris Q, USA) was used for identification of possible metabolites after biodegradation of PNP, the specific operating conditions refer to Chauhan (2010).

Voltage (*V*) was continuously measured by a digital multimeter (UNI-T 803) with a data acquisition system. Power density (*P*) was calculated according to $P = U^2/RA$; where *U* is voltage (V), *R* is resistance (Ω), *A* the surface area of electrode (m²). Coulombic efficiency (CE) was calculated according to CE = $(\sum_{i=1}^{n} U_i t_i / RFb\Delta SV)M \times 100\%$ (Logan et al., 2006); where *F* is Faraday's constant (96,485 C mol⁻¹ electrons), *b* is the number of moles of electrons produced per mole of substrate (4 mol mol⁻¹), ΔS is the substrate concentration, *M* is the molecular weight of the substrate (32 g mol⁻¹).

3. Results

3.1. The experimental results of MFC using different concentrations of PNP as substrate

The cell voltage, PNP degradation rate and COD removal rate in the MFC were studied when using different concentrations of PNP as a substrate. The cycle of electricity generation for cathode fed with different initial PNP concentrations is shown in Fig. 1. In this figure, the maximum cell voltage increased from 100.6 to 183.8 mV, and the maximum power density increased from 0.447 to 1.778 mW m⁻² when the initial PNP concentration increased from 50 to 300 mg l⁻¹. Results show that the cell voltage value increased gradually first and reached to the maximum value due to sludge



Fig. 1. Time-voltage curves of the MFC using different concentrations of PNP as substrate.

biological activity, and then decreased gradually till it was stable. The maximum voltage increased with the PNP concentration increase, but after the steady state, the average voltage decreased with the increase of PNP concentration.

Fig. 2 shows the PNP degradation rates and COD removal rates at different initial PNP concentrations. Over the same period, PNP degradation rate decreased from 64.69 to 33.68%, COD removal rate decreased from 62.87 to 30.67% and CE decreased from 1.128 to 0.264% when the initial PNP concentration increased from 50 to 300 mg l^{-1} . The results indicated that substrate degradation decreased with the increase of the initial PNP concentration.

3.2. The experiment results of MFC using different mixed organic matters as co-substrate

PNP can inhibit the growth of microorganisms for it is toxic, and using an easily degradable organic matter as a co-substrate is beneficial for the microbial growth. The MFC was operated using different mixed substrates with an external 1900 Ω resistor. The cycle of electricity generation for the cell fed with different initial substrate(s) (PNP 100 mg l^{-1} , PNP 100 mg l^{-1} + G 100 mg l^{-1} , PNP 100 mg l^{-1} + G 500 mg l^{-1} , PNP 100 mg l^{-1} + G 800 mg l^{-1} , PNP 100 mg l^{-1} + NaAC 500 mg l^{-1} , respectively) is shown in Fig. 3. Results show that the electricity generation was stable in the experiments. The addition of G or NaAC affected the cell voltage, which was significantly higher than that without G or NaAC, an increase from 139.2 to 341.9 mV when the G concentration increased from 0 to 800 mg l⁻¹. Moreover, when 500 mg l⁻¹ NaAC was used as a co-substrate, the average voltage value was 402.3 mV and the maximum power density was 8.518 mW m^{-2} , which were significantly higher than that when 500 mg l^{-1} G was added alone.

Fig. 4 shows the results of the PNP degradation rates and COD removal rates with different co-substrate combinations. By maintaining an initial PNP concentration of 100 mg l⁻¹ and G as a co-substrate, the maximum PNP degradation rate was 83.51% when G was 500 mg l⁻¹. The COD removal rates were 58.64, 74.89, 74.82 and 80.44% when 0, 100, 500, 800 mg l⁻¹ G were added, respectively. But when 500 mg l⁻¹ NaAC was used as a co-substrate, the



Fig. 2. PNP degradation rates (a) and COD removal rates (b) in the MFC using different concentrations of PNP as substrate.



Fig. 3. Time-voltage curves of the MFC using mixed organic matters as co-substrate.

PNP degradation rate and COD removal rate were greater than those by G added, up to 95.96 and 90.33%, respectively. A maximum CE of 1.708% was obtained when PNP 100 mg l^{-1} + NaAC 500 mg l^{-1} as the co-substrate in the MFC, which is considerably lower than the CE of domestic sewage. There are many reasons behind these results, such as toxicity of PNP, the type of proton exchange membrane (PEM), etc.

3.3. Metabolites of anaerobic PNP biodegradation in the MFC

The decoloration of yellow color of PNP indicated that there were metabolites produced in the process of PNP biodegradation. The metabolites were analyzed by GC–MS at the end of electric cycle (142 h) in the MFC. In the GC–MS chromatogram (Fig. 5), the major peaks showing that 2-pyridinecarboxaldehyde could be produced during the metabolism of PNP in fuel cell since it was originally absent in the anaerobic sludge. Its concentration was very



Fig. 4. PNP degradation rates (a) and COD removal rates (b) in the MFC using mixed organic matter as co-substrate.



Fig. 5. GC-MS analysis of sample of the supernatant collected at the end of electric cycle in the MFC.

low, but there were not other metabolites to be detected according to the peaks, therefore PNP was thoroughly biodegraded after 142 h in the MFC.

4. Discussion

Electricity generated in MFC system increased with the increase of PNP concentration. When compared with results reported earlier, it is much smaller than that using acetate or butyrate as the substrate. In addition, the CE of MFC was very low. There are many reasons responsible for this: 1) PNP inhibits the growth of microorganisms for it is toxic, resulting in low electrical energy; 2) The power density generated is known to be limited by high internal resistance resulting from larger distances between the electrodes (Liu et al., 2005; Cheng et al., 2006). The feasibility of MFC in bioelectricity generation from PNP was demonstrated, but there are many further experiments awaiting us to prove the most effective anaerobic microorganisms producing electricity in the MFC, improvement of MFC configuration, etc.

When using some easily degradable substrates as a supplementary substrate together with PNP in the MFC, the electricity generation, PNP degradation rate were all higher than those using PNP alone as substrate. When using NaAC as a co-substrate, the PNP degradation rate and the average voltage were all higher than those by using glucose alone. Because NaAC can be biodegraded more easily to supply electrons to increase the metabolic rate of anaerobic bacteria with sufficient anaerobic terminal electron acceptors, higher removal rate of PNP was achieved by the MFC. However, once a sufficient amount of biomass formed, the use of an excessive substrate was unnecessary, because when the concentration of cosubstrate is higher than some threshold value, the metabolic reaction rate of microorganism on electrode surface reached peak value, so were the electronic quantity of the electrode surface and PNP degradation rate.

PNP can be metabolized by microorganisms and degradation has been shown by GC–MS analyses. PNP was degraded through 2-Pyridinecarboxaldehyde as the key intermediate. We proposed that 2-Pyridinecarboxaldehyde are produced from PNP followed by a transformation into phenol or some long-chain alkanes with a related release of nitrite, and finally, phenol or long-chain alkanes further is transformed to acetic acid and NH₄–N. Research results to current literature (Kuşçu and Kallas, 2005; Gemini et al., 2005; Wan et al., 2007; Sreenivasulu et al., 2012) about the anaerobic degradation pathways and intermediate products of phenol compounds indicated that further studies are needed because of many disparities and gaps of all kinds of research.

5. Conclusions

The feasibility of MFC in bioelectricity generation from *p*-nitrophenol was demonstrated, simultaneity degrading PNP in high efficiency. A maximum voltage of 402.3 mV and a maximum PNP degradation rate of 95.96% were obtained using PNP as substrate in the MFC in the study. Easily degradable organic matter, such as glucose and sodium acetate can obviously improve the electricity generation and PNP degradation rate, and their optimal added concentration was 500 mg l⁻¹, moreover, sodium acetate is more conductive to the degradation of PNP than glucose in the MFC. Furthermore, the remains of the PNP analysis with GC–MS showed that the PNP was thoroughly biodegraded after 142 h in the MFC.

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References

- Aelterman, P., Rabaey, K., Pham, T.H., Boom, N., Verstraete, W., 2006. Continuous electricity generation at high voltages and currents using stacked microbial fuel cells. Environmental Science & Technology 40, 3388–3394.
- Chauhan, A., Pandey, G., Sharma, N.K., Pual, D., Pandey, J., Jain, R.K., 2010. p-Nitrophenol degradation via 4-Nitrocatechol in *Burkholderia* sp. SJ98 and cloning of some of the lower pathway genes. Environmental Science & Technology 44, 3435–3441.
- Cheng, S., Liu, H., Logan, B.E., 2006. Increased power generation in a continuous flow MFC with advective flow through the porous anode and reduced electrode spacing. Environmental Science & Technology 40, 2426–2432.
- Gemini, V.L., Gallego, A., Oliveira, V.M., Gomez, C.E., Manfio, G.P., Korol, S.E., 2005. Biodegradation and detoxification of *p*-nitrophenol by *Rhodococcus wratisla-viensis*. International Biodeterioration & Biodegradation 55, 103–108.

- Guo, K.M., Deng, Y.J., 1998. Study on the suitable conditions of *p*-nitrophenol with ultraviolet spectrophotometer. Environmental Pollution & Control 20, 47–49.
- Huang, Y., 1996. Determination of COD_{cr} using sealed vessel Microwave Difestion method. Shanghai Environmental Sciences 15, 32–34.
 Ieropoulos, I.A., Greenman, J., Melhuish, C., Hart, J., 2005. Comparative study of
- three types of microbial fuel cell. Enzyme and Microbial Technology 37, 238–245.
- Kuosa, M., Kallas, J., 2009. Influence of t-butanol on multicomponent reaction kinetics and mass transfer in p-nitrophenol ozonation at low pH. Chemical Engineering and Processing 48, 1212–1221.
- Kuşçu Ö, S., Sponza, D.T., 2005. Performance of anaerobic baffled reactor (ABR) treating synthetic wastewater containing p-nitrophenol. Enzyme and Microbial Technology 36, 888–895.
- Liu, H., Cheng, S., Logan, B.E., 2005. Production of electricity from acetate or butyrate using a single-chamber microbial fuel cell. Environmental Science & Technology 39, 658–662.
- Liu, Z.D., Liu, J., Zhang, S.P., Su, Z.G., 2009. Study of operational performance and electrical response on mediator-less microbial fuel cells fed with carbon-and protein-rich substrates. Biochemical Engineering Journal 45, 185–191.
- Logan, B.E., 2005. Simultaneous wastewater treatment and biological electricity generation. Water Science and Technology 52, 31–37.
- Logan, B.E., Hamelers, B., Rozendal, R., Schröder, U., Keller, J., Freguia, S., Aelterman, P., Verstraete, W., Rabaey, K., 2006. Microbial fuel cells: methodology and technology. Environmental Science & Technology 40, 5181–5192.
- Lovely, D.R., Phillips, E.J.P., 1988. Novel mode of microbial energy metabolism: organism carbon oxidation coupled to dissimilatory reduction of iron and manganese. Applied and Environmental Microbiology 54, 1472–1480.
- Min, B., Logan, B.E., 2004. Continuous electricity generation from domestic wastewater and organic substrates in a flat plate microbial fuel cell. Environmental Science & Technology 38, 5809–5814.
- Paola, A.D., Augugliaro, V., Palmisano, L., Pantaleo, G., Savinov, E., 2003. Heterogeneous photocatalytic degradation of nitrophenols. Journal of Photochemistry and Photobiology A: Chemistry 155, 207–214.
- Sreenivasulu, C., Megharaj, M., Venkateswarlu, K., Naidu, R., 2012. Degradation of p-nitrophenol by immobilized cells of *Bacillus* spp. isolated from soil. International Biodeterioration & Biodegradation 68, 24–27.
- Sun, M., Mu, Z.X., Sheng, G.P., Shen, N., Tong, Z.H., Wang, H.L., Yu, H.Q., 2010. Hydrogen production from propionate in a biocatalyzed system with in-situ utilization of the electricity generated from a microbial fuel cell. International Biodeterioration & Biodegradation 64, 378–382.
- Wan, N.S., Gu, J.D., Yan, Y., 2007. Degradation of *p*-nitrophenol by Achromobacter xylosoxidans Ns isolated from wetland sediment. International Biodeterioration & Biodegradation 59, 90–96.
- Wen, Q., Wu, Y., Cao, D.X., Zhao, L.X., Sun, Q., 2009. Electricity generation and modeling of microbial fuel cell from continuous beer brewery wastewater. Bioresource Technology 100, 4171–4175.
- Zhang, J.S., Sun, Z.T., Li, Y., Peng, X., Li, W., Yan, Y., 2009. Biodegradation of *p*-nitrophenol by *Rhodococcus* sp. CN6 with high cell surface hydrophobicity. Journal of Hazardous Materials 163, 723–728.