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The objective of this study is to summarize the effects of surfactants on anaerobic 18 digestion of waste activated sludge. The increasing amount of waste activated sludge 19 has caused serious environmental problems. Anaerobic digestion (AD), as the main 20 treatment for waste activated sludge containing three stages (i.e., hydrolysis, 21 acidogenesis and methanogenesis), has been widely investigated. Surfactant addition 22 has been demonstrated to improve the efficiency of AD. Surfactant, as an amphipathic 23 substance, can enhance the efficiency of hydrolysis by separating large sludge and 24 releasing the encapsulated hydrolase, providing more 25 e for subsequent acidogenesis. Afterwards, the short chain fatty acids (CFA), as the major product, have 26 been produced. Previous investigations revealed that surfactant could affect the 27 transformation of SCFA. They changed the types of acidification products by 28 nd in the ratio of carbon to nitrogen (C/N), promoting changes in microbial activity 29 especially the ratio of acetic and propionic acid, which were applied for either the 30 removal of nutries or the production of polyhydroxyalkanoate (PHA). In addition, the 31 activity of microorganisms can be affected by surfactant, which mainly leads to the 32 activity changes of methanogens. Besides, the solubilization of surfactant will promote 33 the solubility of contaminants in sludge, such as organic contaminants and heavy metals, 34 by increasing the bioavailability or desorbing of the sludge 35

Keywords: Surfactant; Sludge; Anaerobic digestion; Contaminants removal

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39 **1. Introduction**

Nowadays, more and more contaminants existing in water need to be 40 remediated adequately. One of the most widely used means for 41 contaminant removal is activated sludge process [1]. As a kind of 42 biological wastewater treatment, large amounts of waste activated sludge 43 (WAS) are produced in wastewater treatment [2]. The amount of WAS 44 increases with the quantitative dilatation of municipal and industrial 45 wastewater [3]. The increasing quantity of WA become an 46 environmental problem which needs to be solved urseally [4]. For example, 47 the quantity of WAS in China is expected to perease to 34 million tons (at 48 a moisture content of 80%) in 2018, and more than 75% would be handled 49 Therefore, an efficient sludge treatment technique is insecurely [5]. 50 converting the easily WAS theatment [6]. strongly desired for 51 biodegradable organic matters into relatively stable substances, and 52 keeping the residues below the standard values [7-9]. 53

To solve the problems of sludge over quantification, many techniques have been used. In developed countries, concentration and anaerobic digestion are the most common methods for WAS treatment. The corresponding cost for WAS treatment accounts for 60% of the total running cost of wastewater treatment plants (WWTPs) [10], which suggests that the method of concentration and anaerobic digestion cannot be widely accepted. Other methods such as landfill and ocean dumping

technology, due to their negative impacts on the environment-spread of 61 toxic substances in soil and low-cost utilization of land, are being used 62 rarely [10]. Currently, the re-utilization of sludge resources is generally 63 expected [11, 12]. Anaerobic digestion, a method consisting of three stages, 64 i.e., hydrolysis, acidogenesis and methanogenesis, has been widely applied 65 for sludge stabilization. Pollution control and energy recovery can be 66 fulfilled at the same time, which is one of the advantages of anaerobic 67 digestion [13]. In addition, stabilizing organic matters of the sludge and 68 hindering the harmful chemicals into environment an be achieved 69 concurrently [4]. Moreover, getting biogas of short chain fatty acids from 70 this process can also be achieved [8, 14, The main component of the 71 biogas is methane which can be used as the resource, and SCFA can be 72 anoates or used as a preferable carbon applied to generate polyhydroxya 73 source to remove nutrient during wastewater treatment [15, 16]. The 74 characteristics including production of renewable sources, concepts of 75 integrated biorefining and advanced waste treatment, will make it possible 76 for anaerobic digestion to be widely used in the future [17]. Therefore, 77 more and more researchers have studied to improve the efficiency of 78 anaerobic digestion [18]. However, the disadvantages of long reaction 79 period and low efficiency limit its development. 80

81 Surfactant, possessing both hydrophobic groups and hydrophilic 82 groups, is now widely studied and used [19, 20]. The classification of

surfactants is generally divided into chemical surfactants (CSF) and 83 biological surfactants (BSF); or divided into cationic, anionic, nonionic, 84 and zwitterionic surfactants [21, 22]. The chemical surfactants, for instance, 85 sodium dodecyl sulfate (SDS), sodium dodecyl benzene sulfonate (SDBS), 86 linear alkylbenzene sulphonates (LAS), and Triton X-100 etc., are widely 87 used in practical applications [23]. However, CSFs are commonly toxic to 88 the environment and easily accumulated in the environment [24]. BSFs, 89 such as rhamnolipid (RL), saponin (SP), surfactin, (SF) and glycolipid, 90 have advantages such as biodegradability, efficience at harsh temperature 91 or pH, and lower toxicity compared to the chanical counterparts [14, 25]. 92 Surfactant can be adsorbed on the interface reduce surface tension, or form 93 micelles. The insoluble substance gets into the micelles and enhances its 94 solubility [26-28]. 95

In process of anaerabic digestion, surfactant can enhance solubility of 96 organic matter to the cludge in order to improve the efficiency of hydrolysis 97 and remediation 29, 30]. Different surfactants can lead to different 98 amounts of acetic and propionic acid (short chain fatty acids), and can also 99 affect the growth of polyphosphate accumulating organisms (PAO), 100 glycogen accumulating organisms (GAO) and methanogens. In addition, 101 surfactant can influence some characteristics of the sludge. For example, 102 surfactant can change the pH and the structure of the sludge component 103 such as proteins and carbohydrates. Moreover, surfactant can alter sludge 104

floc diameter which is closely related to anaerobic digestion of sludge [31, 105 32]. Furthermore, surfactants also have a good performance in the removal 106 of pollutants during the anaerobic digestion process [33]. Two kinds of 107 organic pollutions are discussed generally in the sludge-organic pollution. 108 hydrophobic organic compounds and heavy metal pollutants [34]. It is 109 concluded that the removal mechanisms about organic contaminants by 110 surfactant can be summed up into three aspects: emulsification of liquid 111 pollutant, micellar solubilisation, and facilitated transport All mechanisms 112 are designed to increase microbial contact with pollutent, and improve the 113 efficiency of microbial treatment [27]. In order to promote the removal of 114 metal ions from the sludge, surfactant may act in two ways, i.e., ion 115 exchange and complexing with metal ons [24]. 116

Recently, some investigation, have found the advances in utilization of surfactants during the dewatering of sludge. But the process of anaerobic digestion has been neelected, which will be summaried in this article. The surfactants can work in two aspects: improving anaerobic digestion effciency and enhancing removal of contaminants, which will be discussed in this paper. In addition, this review expounds the theory direction and future prospect.

124 **2. Influence of surfactants on properties of WAS**

125 **2.1 Effect on physicochemical properties**

126 In the process of anaerobic digestion, pH value is a significant impact

127	factor, and its effect runs through the whole anaerobic digestion process.
128	At pH 6.5-7.2, the process of methane production is optimized; while at
129	pH 4.0-8.5, it is optimized for fermentative process [8]. Different pH values
130	also have different influences on acidification process. When pH is higher
131	than 8, the products obtained in the process of acidification are gradually
132	shifted from acetic and butyric acids to acetic and propionic acids [8]. The
133	addition of surfactants can change the pH value of sludge. It was reported
134	that with an increase of SDS dosage in the sludge from 0 to 50 mg/g SS of
135	SDS, the sludge pH increased from 6.1 to 7.1 [3, 35] The effect of another
136	chemical surfactant, SDBS, on the pH of the sudge is different from SDS.
137	When 20-50 ppm SDBS was added into WAS, the pH dropped from 7.4 to
138	6.0 within 20 days [36, 37]. However, the alkaline condition is better than
139	the acid condition to promote the macrobic digestion process [8].
140	Generally, the main constituents of sludge are proteins, carbohydrates
141	and lipids [14] The chemical surfactants, which are more toxic than
142	biological surfactants, may lead to changes in the structure of the sludge.
143	The surfactant can lead to denaturation of proteins via tertiary structure
144	unfolding in the sub-micellar and chain expansion [38]. The micelle-like
145	cluster composed of surfactants will form in the presence of proteins. Chen
146	et al. (2013) found that with the existence of SDBS, the fluorescence peak
147	intensity of proteins decreased from 5.14×10^6 to 2.88×10^6 , indicating that
148	SDBS was the main contributor to the denaturation of protein. The

monomers of SDS binding to protein by hydrophobic interactions lead to 150 unfold of the tertiary structure at the sub-CMC concentration. However, 151 when SDS concentration is above CMC, the micelles nucleate on the 152 hydrophobic patches of the protein chain cause it to expand [38]. The most 153 important conclusion obtained from the experiment is that when a variety 154 of interactions between proteins and surfactants are compared, specific ion 155 interactions are greater than nonspecific hydrophobic interactions, 156 indicating that the details of the process depend on the type of surfactant 157 [39, 40]. The anionic surfactants, such as SDS and Set whing to longer 158 alkyl chain, having more strongly protein flucescence. The different head 159 groups of different anionic surfactants do not show distinct difference. 160 However, the nonionic surfactants, such as TX-100, surfactin and saponin, 161 show less interaction with prote because of non-mainstream type of 162 reaction [40, 41].

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Morphology is the characterization parameter investigated generally 163 for sludge. Generally, the addition of surfactant is accompanied by the 164 occurrence of saponification [42, 43], which would cause the fast reduction 165 of mean projected area, diameters and alter the flocs shape expressed as 166 circularity index. In consideration of the influence of morphology by SDS 167 addition, two ranges of concentrations can be discriminated: 0.0025–0.025 168 g/L and 0.25–2.5 g/L. It was increased by 8% for mean projected area after 169 24 h without SDS addition. However, this parameter was reduced about 30% 170

in the run containing SDS at concentrations 0.0025-0.025 g/L and >55% at 171 concentrations of 0.25-2.5 g/L. In addition, other factors including 172 diameter, convex perimeter, ferret diameter and perimeter were also 173 enhanced about 8-9% after 24 h in the control run; while decreased in the 174 run with SDS about 10-15% for concentrations at 0.0025-0.025 g/L, and 175 30-40% for concentrations at 0.25-2.5 g/L [42]. At present, this 176 phenomenon has not been systematically elaborated and the mechanism 177 has not been fully expounded, which also provides a reference for future 178 research. 179

However, there is few papers clearly indicating that the BSF has any observed effects on the pH and its construents as well as morphology, which suggests that the direction is worth studying in depth.

183 2.2 Impact on biological properties

The microbial elements of sludge play very important role in the 184 anaerobic digestion process, and thus the microbiological factors should be 185 The microorganisms contained several groups are focused [44]. 186 complicated and delicate in the process of anaerobic digestion. The whole 187 microbe-process requires the design of the optimal method because the 188 optimal operation conditions of different microbes cannot be completely 189 overlapped [8]. The complex microbial community usually consists of a 190 large number of bacteria and archaea populations, which are usually related 191 to hydrolysis, acidification, biogas production and pollutant removal [45]. 192

The core groups of bacteria in anaerobic digesters consist of *Chloroflexi*, 193 Betaproteobacteria, Bacteroidetes, and Synergistetes. However, 194 Methanosarcinales, Methanomicrobiales, and Arc I phylogenetic groups 195 are mainly archaeal community [14, 46]. The effects of surfactant on the 196 characteristics and structure of microbes deserve the attention of 197 researchers [47]. 198

Generally, surfactant can transform cell structure of microorganism 199 via making materials on cell surface depart from the attached site and 200 dissolved in aqueous solution [4]. Specifically, as a subjective surfactant 201 combines the proteins with hydrophilic groups, surfactants could impair 202 the function and integrity of biological meripranes, thereby causing native 203 structure disturbing [48]. Whereas the hydrophobic groups combine with 204 lipids, causing the liquefaction of membranes and impairment of their 205 barrier properties [49]. In the comparison between chemical surfactants 206 and biological surfactants, the biological surfactants show better 207 biocompatibility. With the addition of the same dosage of 40 mg/g TSS of 208 SDS, SDBS, and RL to the sludge, the percentage of Proteobacteria 209 decreased to 46.1% in control, 35.6% with SDBS, 34.7% with SDS, and 210 43.2% with RL after 8 days, respectively, and the same tendency was also 211 found for Bacteroidetes and Chloroflexi [50]. However, for Firmicutes, a 212 critical participant in hydrolysis and acidification, the percentage was the 213 highest (in relative abundance) in SDS about 26.9%, higher than that in RL 214

about 24.4%, and were found to be lower in SDBS about 6.7% and the 215 lowest in Control about 4.2% [31, 51, 52]. However, the exact mechanism 216 of SDS enhancing the abundance of Firmicutes is still unknown. In 217 addition, RL shows better biocompatibility, which allows highly active 218 hydrolysis and is favorable to functional microbe for further interaction in 219 anaerobic digestion [51]. The effects of different biosurfactants are 220 different. Previous experiments showed that the negative effects of SF and 221 RL on diversity of metabolic and species were conforming. However, SP 222 showed much fewer block on diversity of metabolic species than RL 223 and SF, and exhibited better biocompatibility even if possessed inferior 224 surface activity than SF and RL [45, 46]. The reasons may be related 225 to the difference of characteristic between anionic surfactants (SP and RL) 226 and nonionic surfactants (SP). filuences of the addition of surfactants 227 on microbes are shown in Fig. 1. 228

In general microorganisms and enzymes are always complementary 229 to the biological characteristics of the sludge [54]. In this complex enzyme 230 system, the various processes in which the enzymes involved are protease 231 and α -glucosidase during hydrolysis, acetatekinase during acidification, 232 coenzyme during methanation, and dehydrogenase about 233 F_{420} microorganisms [15]. Among them, the composition of the hydrolase is 234 more complex. Four kinds of hydrolases, i.e., a-glucosidase, alanine-235 aminopeptidase, esterase, and dehydrogenase, play important roles in 236

hydrolysis [55]. A-glucosidase can degrade starch, while alanine-237 aminopeptidase is responsible for the degradation of proteins. For esterase 238 activity, measuring as hydrolysis fluorescein diacetate (FDA), does not 239 produce information about specific substances degradation. Whereas, 240 dehydrogenase activity has been found to correlate with substrate removal 241 in sludge [56-59]. These enzymes organically form an important part of the 242 enzyme system, which can promote the disintegration of large particles and 243 produce more surface for attaching of microbes, leading to a high 244 efficiency degradation [11]. 245

The effects of surfactants on enzyme activity have also been 246 investigated. SDS suppresses the ATPase activity of P-glycoprotein at low 247 concentrations [60]. However, it stimulates protease and amylase activities. 248 The increased amount can be astributed to destruction of the sludge matrix 249 and release of the enzymes immobilized on the floc structure [3, 26, 35]. 250 When the dose increases to a certain level, it will have a hindrance. Another 251 chemical anionic surfactant, SDBS, has the analogous effects on enzymes. 252 It can enhance the activity of protease and a-glucosidase, but the exact 253 reasons for this phenomenon are still unclear [2]. When the dose of 254 additional SF, RL, and SP was 50, 50, 100 mg/g DS, respectively, the 255 biosurfactant RL was the most powerful one compared to others which 256 gained the activities of neutral protease and α -glucosidase to 4.07 and 5.73 257 times, respectively. SF and SP had the same effects, but the increase was 258

less than that of RL. Furthermore, the addition of RL made the activity of 259 coenzyme F₄₂₀ decrease by 40%. RL also possessed a violent negative 260 impact on the dehydrogenase and acetate kinase activities. SF also had a 261 wicked effect on the activity of the coenzyme F_{420} , dehydrogenase and 262 acetate kinase, but the effects were weaker than that of RL. For example, 263 the activity of acetate kinase was 73% after SF addition, but it was 26% 264 after RL addition. When it came to methane production, SF addition 265 slightly reduced with the increase of dose in the original period, but the 266 throughput gradually raising and outstripping the control test after 6 d. In 267 contrast, RL invariably showed powerful invibition of methanogenesis 268 which was probably attributed to the decrease of coenzyme F_{420} activity 269 [15]. SP addition had not shown any individual in the test 270 with each dose of SP remaining the same to the control test which indicated 271 that methanogenesis was not influenced by SP addition [15]. In general, 272 researches have summed up that the influence of enhancing or inhibiting 273 by surfactants may depend on the length of alkyl chain, but there is no 274 specific experiment to prove it, which could be an in-depth point [26, 61, 275 62]. 276

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278 **3. Impact on anaerobic digestion**

Anaerobic digestion, containing hydrolysis, acidification, and methanogenesis, is a complex biochemical process. Surfactant, owing to special characteristics, may have some effects on sludge during anaerobic
digestion. Hence, the effect of surfactants on the three stages of anaerobic
digestion will be discussed, respectively.

284 **3.1 Hydrolysis**

The mechanisms of the increment of hydrolysis by surfactant can be 285 summed up in two aspects: sludge components and enzyme activity (Fig. 286 2). The sludge blocks will be dispersed, and the hydrolase will be released 287 from the sludge, which increase the efficiency of hydrolysis. And the latter 288 has been discussed in the second chapter. Expectitular polymeric 289 substances (EPS) is the main part of sludge components [44]. The main 290 fractions of EPS are proteins and carbohydrates [63]. Surfactant can cause 291 the break-up of sludge substance, especially the EPS, which releases more 292 proteins and carbohydrates 4]. The existence of the electrostatic 293 interaction between enzymes and extracellular polymer substances leads to 294 the complexes of extracellular polymer substance-enzyme, which traps 295 enzymes in substrate. Therefore, the activity of enzymes have increased 296 due to the release of enzymes by surfactant addition [65, 66]. In addition, 297 surfactants enhance solubility of material particles by reducing surface 298 tension or forming micelles, which can also improve the hydrolysis 299 efficiency [14]. As one of the most widely used chemical surfactant, the 300 impact of SDS was discussed previously. With the addition of SDS, the 301 thicknesses of protein and carbohydrate all increased. In a fermentation 302

experiment, the thicknesses of protein and carbohydrate reached 0.3418 303 and 0.5159 g/L in the control test, 0.8277 and 0.1576 g/L with 100 mg/g 304 SDS, and 1.3729 and 0.2209 g/L with 300 mg/g SDS dosage, respectively, 305 in the sixth day of fermentation [1]. Ji et al. (2010) found that by adding 20 306 mg/g of SDBS in fermentation system, the maximal proteins and 307 carbohydrates released were 1.7 and 1.9 times of those from the control in 308 the sixth day of fermentation, respectively. As for biosurfactant, alkyl poly 309 glycosides (APG), a kind of widely used surfactant, its influence has been 310 investigated. In previous experiments, the maximal concentrations of 311 proteins and carbohydrates were 3.3020 and 0.6580 g/L in WAS+SDS 312 system with SDS dose of 200 mg/g (DS, respectively, whereas the 313 corresponding concentrations were 1.870 and 1.2060 g/L in WAS+APG 314 TSS, respectively, indicating that SDS system with APG dose of 200 mg 315 enhanced the protein production, and APG enhanced the carbohydrate 316 production [26]. This appearance can be attributed to the hindrance of 317 composing of the enzymes involved in protein hydrolysis [67]. For the 318 other biosurfactants, it has been confirmed that the concentration of 319 proteins increased with the addition of RL, SF and SP during the initial 60 320 mins, which was related to the dose. Comparing with SP, however, SF as 321 well as RL had a preferable impact on the solubilization of EPS [68]. The 322 essence of this phenomenon is partly because of the weaker surface activity 323 of SP than RL and SF; and the other part is because of the degradation of 324

SP, and this degradation is not observed for RL and SF [15, 69, 70].

326 **3.2 Acidification**

Generally, SCFAs, the products of acidification, are the designation 327 of a series of acids, including acetic acid, propionic acid, and butyric acid, 328 etc. [71]. Wherein, acetic acid and propionic acid are the two kinds of acids 329 with the largest amount, and their proportion has a profound effect on the 330 properties of SCFAs. For different purposes of production, there are 331 different requirements for the intermediate products. The ultimate purpose 332 of acidification is promoting more methane production Nerefore, a higher 333 proportion of acetate is required, which is attributed to the direct 334 degradation of acetic acid by methanogen [72] and other SCFAs should 335 be converted into acetic acid before being used to produce methane [73]. 336 However, if the purpose of asid seation is to enhance the efficiency of 337 biological nutrient removal (BNR), one feasible means of supplying PAO 338 with a selective advantage over GAO is through operating the carbon 339 source composition. Investigation has suggested that PAO activity with 340 propionate is greater as compared to acetate, so it is required that obtaining 341 higher proportion of propionic [74]. The increasing efficiency of BNR by 342 propionic could be attributed to the different characteristics between PAOs 343 and GAOs [75]. Both PAO and GAO can consume SCFA to obtain energy, 344 but only PAO can hydrolyze polyphosphate. Therefore it is necessary to 345 inhibit the activity of GAO, and then promote the polyphosphate 346

hydrolysis by enhancing PAO activity [76, 77]. Acetate, as a kind of SCFA, 347 can easily be adsorbed and consumed by GAO and PAO. However, the 348 consumption of propionic acid is distinct. The rate of propionic 349 consumption by GAO is slower than that by PAO. Moreover, PAO has been 350 proven to be more accommodable when carbon source changes as 351 compared to GAO [78, 79]. Therefore, more proportion of propionic can 352 enhance the activity of PAO and baffle the activity of GAO, leading to the 353 enhancement of biological nutrient removal efficiency 354

The most influential factor in the composition short chain fatty 355 acids is pH. With the increase of pH value, the amount of acetate, butyrate, 356 and iso-butyrate all increased, and the same trend has been observed for 357 amount of higher weight molecular, such as valerate, iso-valerate, and 358 caproate [80, 81]. However, the optimal amount interval of propionate is 359 between 6.0-9.0 and its optimal proportion can up to 50% [74]. In addition, 360 the C/N also has definite impact on product of SCFA. Generally, both 361 carbon and nitrog n originate from the product of hydrolysis-protein and 362 carbohydrate. The improvement of C/N of digestive matrix was favorable 363 to the production of propionate [82]. Hence, with a high content of protein, 364 the enhancement of nitrogen elements content has been caused. Therefore, 365 the C/N of digestive matrix becomes excessively small and the production 366 of propionic is limited, and the addition of carbohydrate matter is 367 indispensable [74]. Nevertheless, excessively large C/N also results in 368

some negative phenomenon. The production of PHA requires the nitrogenlimited condition, whereas high C/N prevent the merisis of active biomass
for close connection between nutrition and cell merisis. Nevertheless, Jiang
et al. (2009) have found that enhancing feed degree or optimizing process
factors can achieve high production of PHA even without nitrogen-removal
[83]. The maximum permissible value of C/N is 50, and this value
exceeding 50 may cause the cessation of process [84, 85].

The addition of surfactant may improve acidification efficiency. 376 SCFA yield was enhanced by SDS [86]. Jiang et a (2007) found that, in 377 the sixth day of zymolysis, the concentration of SCFA was 2243.04 mg 378 COD/L with 100 mg/g SDS, whereas it was merely 191.10 mg COD/L in 379 the control. However, with higher concentration of SDS being added, less 380 SCFA was produced during the original stage of zymolysis, which could 381 be ascribed to the negative influence of SDS. For instance, the destruction 382 of microbial protein structure and accumulation in the environment to 383 produce toxic byproducts [87]. The rank of the composition SCFAs was in 384 the order of acetic > propionic > iso-valeric in the control. However, the 385 addition of SDS changed the array to acetic >iso-valeric> propionic. The 386 results showed that the production of SCFA was enhanced remarkably in 387 the presence of SDBS. At 6 days of fermentation time, the maximum SCFA 388 was 2599.1 mg COD/L with 20 mg/g SDBS addition, whereas it was 339.1 389 mg COD/L in the control test without sodium dodecyl benzene sulfonate 390

addition [2]. The same situation was observed when the dose of sodium 391 dodecyl benzene sulfonate was higher than 200 mg/g. After all, the 392 inhibition of microorganism caused by SDS and SDBS cannot be 393 neglectable. However, the rank of all kinds of SCFAs was different from 394 that with SDS addition. During the original 6-day fermentation, there was 395 no doubt that acetic acid was the most universal component, but the 396 propionic acid was the sub major products, their percentages were acetic 397 acid about 27.1%, and propionic acid about 22.8% respectively. The 398 maximum SCFA concentration reached 800 mg CON n the fifth day in 399 the control without biosurfactants. Its production enhanced with the 400 increasing dose of SF or RL (ranging from, 23 to 50 mg/g DS), and the 401 maximum concentration was nearly 3.3 g COD/L. However, SCFA 402 production was distinctly improve when the dose of SP varied from 20 to 403 100 mg/g DS, and the miximum concentration of SCFA was 3.1 g COD/L. 404 There was no stanificant enhancement for higher dosage of SP, RL, and SF 405 [2]. In the aspect of transformation of SCFA components, the emphasis is 406 on the changes in the content of acetic and propionic acid. With SP, RL and 407 SF addition, the percentage of acetic, propionic and n-butyric acid were 408 enhanced in pace with the augmenting dose of the biosurfactants. However, 409 the proportion of acetic acid to propionic acid in reactors with SF or RL 410 addition was higher than that with SP and in the control [16]. In fact, the 411 propionic acid was the main product during the acidification of glucose, 412

whereas the ratio of acetic acid was high when protein was degraded [88], 413 which was probably correlated to the influence of surfactant addition. 414 However, the effect of surfactant degradation on the production of SCFA 415 is also worthy to discuss. There are two mechanisms to enhance SCFA 416 generation, i.e., biological effect and chemical effect. The latter particularly 417 depended on the degradation of surfactant itself. However, in terms of 418 some surfactants that have been discussed, SP possessed a analogical 419 enhancement of SCFA production by its degradation, although its surface 420 ability was lower than SF and RL [15]. Whereas in the ase of SDS, SDBS, 421 RL, and SF, the improved yield of SCFA was primarily caused by 422 biological impact rather than chemical impact 423

424 **3.3 Methanogenesis**

anaerobic digestion, is defined as the Methanogenesis, the last ster 425 process of converting acetate and hydrogen from acidification to methane 426 and carbon divide by methanogenic bacterial [89, 90]. Generally, a 427 significant SCFA consumption was observed in the fermentation of WAS, 428 assumably be attributed to the consumers participation, for instance, 429 methanogens [26]. The order of SCFA consuming is acetate, butyrate, and 430 propionate during the methanogenesis [64], which indicates that a higher 431 proportion of acetic acid is accompanied by a larger amounts of methane 432 production. Two parameters are of great significance in the process of 433 methane production, i.e., pH and activities of methanogenic bacteria. 434

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During the whole period of fermentation, the methane yield enhanced with 435 pH increasing from 4.0 to 6.0, and declined when pH further increased to 436 10.0 [91]. Apparently, the highest methane production was achieved at pH 437 7.0 during fermentation time, which indicated that both the higher and 438 lower pH could decrease the activity of methanogenic bacteria [92]. 439 Previous investigation showed that the production rate of biogas general 440 tended to accord with sigmoid function (S curve), indicating that the 441 methane production can be split into three stages: lag phase-decomposition 442 phase, and flattening phase [3]. Methane production is leastitude at the start 443 and end of curve, suggesting that the mehane generated in reactor 444 corresponds to specific growth rate of methanogenic microbe [93]. To 445 enhance the methanation efficiency and biogas production, various efforts 446 have been made. Preatment efforts contains physical, chemical, and 447 biological treatment. However, different purposes lead to different 448 consequences. Supposing that the purposes were SCFA accumulation and 449 PHA production, the methods reducing methane production had to be 450 adopted. 451

In specific surfactants, SDS was observed to affiliate with the inhibition of methanogens activity during the sludge fermentation [26]. It has been reported that the SDS would inhibit the methanogens acitivities in the period of sludge fermentation. With the dose of SDS raised from 20 to 300 mg/g, the hindrance ratio of methane yield augmented definitely

from 3% to 100% [94]. Another anionic surfactant SDBS also prevented 457 the process of methanogenesis. Total gas yields and methanogenesis from 458 glucose were decreased to half maximal rates at 20 to 50 ppm SDBS during 459 the original period of fermentation [36]. It has shown that the surfactants 460 with aromatic and cyclic, such as SDBS, were found to be the most 461 hazardous compounds for anaerobic acetoclastic methanogenesis [49]. 462 However, SDS is one of the few surfactants with minimal toxicity to 463 methanogens [49]. The inhibition of methane production by saponins is the 464 smallest, which is attributed to the negligible en f methanogenic 465 bacteria activity after saponins addition. Achally, RL possessed serious 466 antibacterial activity to methanogenic bacteria and some related enzymes. 467 As mentioned above, the addition of the activity of coenzyme 468 ecreased by 40%. In addition, it was F₄₂₀, a methane related enzyme 469 confirmed that the RL net only prevent the methanogenesis, but also retard 470 the metabolism of other microbes, which might cause the destruction of 471 biological activity of sludge [94]. 472

473

474 **4. Effects of surfactant addition on contaminant removal**

During waste water treatment, activated sludge process produces a large number of waste activated sludge, which contains a lot of pollutants [95, 96]. They might shift to the different compartments involving atmosphere, soli, and surface water via pinpoint or diffuse inputs [97]. The most discussed contaminants, including hydrophobic organic matter (HOC)
and heavy metals [98, 99]. The removal of these pollutants has always been
a hot issue. The influences of surfactant on representative HOC
contaminants have been summarized in Table 1.

Surfactants have a great potential of solubilization. Generally, there 483 three influence mechanisms for the advancement of HOC are 484 biodegradation by surfactants addition (Fig. 3). The first mechanism is 485 forming micelle shaped by surfactant and encasing the HQC. Therefore 486 microorganisms are able to adsorb the contaminant and the micelles core. 487 In the second mechanism, surfactants endance the mass transfer of 488 contaminants to the aqueous phase for further degrade by microorganisms, 489 which is attributed to the reduced surface tension by surfactant [100]. And 490 drophobicity has been changed by the for the third mechanism, the 491 addition of surfactants, resulting in the direct contact between cells and 492 contaminants [1-13] In addition, there is another mechanism that has 493 been conjectured, in which surfactants promote microorganisms to be 494 adsorbed to sludge surface sites occupied by contaminants [104]. 495

Because of the application of various kinds of metals in industry, there are also various heavy metal ions in the wastewater, which causes the sludge filled with heavy metal ions [105, 106]. Heavy metals cannot be biodegraded. On the contrary, they can only be transformed from one configuration to another, which can change their mobility and toxicity

[107]. Some forms of heavy metals can be transformed by process of redox 501 or by alkylation. There were two main mechanisms for desorption of heavy 502 metals from sludge by surfactant addition (Fig. 4). Firstly, the cationic 503 surfactant can permute the same charged metal ions by rivalry for some but 504 not all negatively charged surface, because of the interaction of repulsion 505 between cationic surfactant and heavy metals. Secondly, the anionic 506 surfactants form nonionic complexes with heavy metal by ionic bonds 507 which are stronger than the bonds of metal with sludge [108, 109]. The 508 metal-surfactant complexes are desorbed from sludge nortance to aqueous 509 due to the decrease of the surface tension [No-112]. In general, the two 510 mechanisms can be concluded in ion exchange and counterion binding. 511

512

513 5. Conclusion and future prospect

This review summarized the utilization of surfactant in the process of 514 anaerobic digestion, including the influences on sludge properties, and 515 conversion process of hydrolysis, acidification and methanogenesis. In 516 addition, due to the excellent solubilization of surfactant, the removal of 517 organic pollution and heavy metals might also be affected by surfactant 518 addition. Surfactants, as an amphiprotic compound, have characteristic of 519 solubilization via reducing interface tension or forming micelles, when the 520 concentration of surfactant is under or above CMC, respectively. In the 521 process of anaerobic digestion, due to the rate-limiting influence of 522

hydrolysis, enhancing its efficiency will lead to the increment of acidification substrate. Therefore, the increased SCFA production can be applied to remove nutrient and produce PHAs. Surfactant can not only affect the proportion of various SCFA, but also influence the activities of certain microorganisms, which have significant roles in anaerobic digestion.

Notably, future investigations can be paid attention to the following 529 aspects: i) Establishing technological process for the production of 530 biosurfactant for industrial production. Although the top icity and risk of 531 biosurfactant are smaller than those of chemical surfactant, the price of 532 biosurfactant is higher than chemical surfactants, which limits its wide 533 application. ii) Establishment of modes to describe the influence surfactant 534 addition on different microorganizers. Due to the complex constitution of 535 microorganism involved in anaerobic digestion, as well as the dual 536 character of susfactant, the quantify effects of surfactant need specific 537 model. iii) Process improvement of producing PHAs from SCFA. The 538 production of PHAs is in the theoretical stage. In order to achieve the 539 consummate craft of waste resources re-utilization, it is worth investigating 540 how to establish a systematic process for PHAs production. 541

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567 **References**

- 568 [1] Jiang, Y. Chen, Q. Zhou, Effect of sodium dodecyl sulfate on waste
- 569 activated sludge hydrolysis and acidification, Chemical Engineering
- 570 Journal 132 (2007) 311-317.
- 571 [2] Jiang., Y. Chen, Q. Zhou, G. Gu, Biological short-chain fatty acids
- 572 (SCFAs) production from waste-activated sludge affected by surfactant,
- 573 Water Res 41 (2007) 3112-3120.
- [3] S. Kavitha, C. Jayashree, S. Adish Kumar, I.T. Yeum L Rajesh Banu,
- 575 The enhancement of anaerobic biodegradability of verte activated sludge
- 576 by surfactant mediated biological pretreatment, Bioresource technology
- 577 168 (2014) 159-166.
- 578 [4] K. Luo, Q. Yang, J. Yu, X.M. L. O.J. Yang, B.X. Xie, F. Yang, W.
- Zheng, G.M. Zeng, Combined Flect of sodium dodecyl sulfate and
 enzyme on waste activated sludge hydrolysis and acidification,
 Bioresource technology 102 (2011) 7103-7110.
- [5] L. Nizzetto, S. Langaas, M. Futter, Pollution: Do microplastics spill on
 to farm soils?, Nature 537 (2016) 488-488.
- [6] P. Xu, G.M. Zeng, D.L. Huang, C.L. Feng, S. Hu, M.H. Zhao, C. Lai,
- ⁵⁸⁵ Z. Wei, C. Huang, G.X. Xie, Use of iron oxide nanomaterials in wastewater
- treatment: A review, Science of the Total Environment 424 (2012) 1-10.
- [7] J. Xiao, L. Zhao, Z. Shen, Enhanced sludge anaerobic fermentation
- 588 using microwave pretreatment combined with biosurfactant alkyl

- 589 polyglycoside, RSC Advances 7 (2017) 43772-43779.
- [8] L. Appels, J. Baeyens, J. Degrève, R. Dewil, Principles and potential of
- the anaerobic digestion of waste-activated sludge, Progress in Energy &
- 592 Combustion Science 34 (2008) 755-781.
- ⁵⁹³ [9] R. Hartenstein, Sludge Decomposition and Stabilization, Science 212
- 594 (1981) 743-749.
- [10] D. Zhang, Y. Chen, Y. Zhao, X. Zhu, New sludge pretreatment method
- to improve methane production in waste activated sludge digestion,
 Environmental Science & Technology 44 (2010) 4802 4808.
- 598 [11] K. Luo, Q. Yang, X.-m. Li, G.-j. Yang, Xiu, D.-b. Wang, W. Zheng,
- 599 G.-m. Zeng, Hydrolysis kinetics in anaeropic digestion of waste activated
- sludge enhanced by α-amylase, Biochenical Engineering Journal 62 (2012) 17-21.
- 602 [12] A. Zhou, C. Yang, Z. Guo, Y. Hou, W. Liu, A. Wang, Volatile fatty 603 acids accumulation and rhamnolipid generation in situ from waste
- activated sludge formentation stimulated by external rhamnolipid addition,
- Biochemical Engineering Journal 77 (2013) 240-245.
- [13] Y. Chen, J.J. Cheng, K.S. Creamer, Inhibition of anaerobic digestion
- process: a review, Bioresource technology 99 (2008) 4044-4064.
- 608 [14] K. Luo, Q. Ye, X. Yi, Q. Yang, X.M. Li, H.B. Chen, X. Liu, G.M.
- 609 Zeng, Hydrolysis and acidification of waste-activated sludge in the
- 610 presence of biosurfactant rhamnolipid: effect of pH, Applied microbiology

- and biotechnology 97 (2013) 5597-5604.
- [15] X. Huang, C. Shen, J. Liu, L. Lu, Improved volatile fatty acid
 production during waste activated sludge anaerobic fermentation by
 different bio-surfactants, Chemical Engineering Journal 264 (2015) 280290.
- [16] W.S. Lee, A.S.M. Chua, H.K. Yeoh, G.C. Ngoh, A review of the
- ⁶¹⁷ production and applications of waste-derived volatile fatty acids, Chemical
- 618 Engineering Journal 235 (2014) 83-99.
- 619 [17] M. Madsen, J.B. Holm-Nielsen, K.H. Esberger, Monitoring of
- anaerobic digestion processes: A review perspective, Renewable and Sustainable Energy Reviews 15 (2011) 3141-3155.
- [18] D.R. Lovley, Happy together: microbial communities that hook up to
 swap electrons, The Isme Journal (2016) 327.
- [19] A. Rodrigues, R. Yogueira, L.F. Melo, A.G. Brito, Effect of low
 concentrations of synthetic surfactants on polycyclic aromatic
 hydrocarbons (PAH) biodegradation, International Biodeterioration &
 Biodegradation 83 (2013) 48-55.
- [20] Z. Liu, Z. Zeng, G. Zeng, J. Li, H. Zhong, X. Yuan, Y. Liu, J. Zhang,
- M. Chen, Y. Liu, Influence of rhamnolipids and Triton X-100 on adsorption
- of phenol by Penicillium simplicissimum, Bioresource technology 110(2012) 468.
- [21] S. Wang, C.N. Mulligan, Rhamnolipid biosurfactant-enhanced soil

- flushing for the removal of arsenic and heavy metals from mine tailings,
 Process Biochemistry 44 (2009) 296-301.
- 635 [22] T.T. Nguyen, N.H. Youssef, M.J. McInerney, D.A. Sabatini,
- 636 Rhamnolipid biosurfactant mixtures for environmental remediation, Water
- 637 Res 42 (2008) 1735-1743.
- [23] J. Jensen, Fate and effects of linear alkylbenzene sulphonates (LAS)
- 639 in the terrestrial environment, Science of the Total Environment 226 (1999)
- 640 **93-111**.
- 641 [24] I.M. Banat, R.S. Makkar, S.S. Cameotra, Retained commercial
- applications of microbial surfactants, pplied Microbiology &
 Biotechnology 53 (2000) 495.
- [25] D.T. Sponza, O. Gok, Effect of rhymolipid on the aerobic removal of
- 645 polyaromatic hydrocarbons (Phys) and COD components from
- 646 petrochemical wastewater, Bioresource technology 101 (2010) 914-924.
- [26] L. Xu, Cherry Li, X. Sun, J. Shen, J. Li, L. Wang, Role of
 surfactants on the hydrolysis and acidogenesis of waste-activated sludge,
- 649 Desalination and Water Treatment 57 (2015) 16336-16345.
- [27] F. Volkering, A.M. Breure, W.H. Rulkens, Microbiological aspects of
- surfactant use for biological soil remediation, Biodegradation 8 (1997)
 401-417.
- [28] J. Wan, G. Zeng, D. Huang, L. Hu, P. Xu, C. Huang, R. Deng, W. Xue,
- 654 C. Lai, C. Zhou, Rhamnolipid stabilized nano-chlorapatite: Synthesis and

- enhancement effect on Pb-and Cd-immobilization in polluted sediment,
 Journal of hazardous materials 343 (2017).
- [29] X. Tan, Y. Liu, G. Zeng, X. Wang, X. Hu, Y. Gu, Z. Yang, Application
- of biochar for the removal of pollutants from aqueous solutions,
- 659 Chemosphere 125 (2015) 70-85.
- [30] Z. Liu, M. Yu, G. Zeng, M. Li, J. Zhang, H. Zhong, Y. Liu, B. Shao,
- 661 Z. Li, Z. Wang, Investigation on the reaction of phenolic pollutions to
- 662 mono-rhamnolipid micelles using MEUF, Environmental Science &
- Pollution Research International 24 (2016) 1-11.
- [31] Y. Chen, K. Liu, Y. Su, X. Zheng, Q. Wang Continuous bioproduction
- of short-chain fatty acids from sludge enhanced by the combined use of
 surfactant and alkaline pH, Bioresource technology 140 (2013) 97-102.
- 667 [32] Liu., G.M. Zeng, H. Zhons, X.Z. Yuan, H.Y. Fu, M.F. Zhou, X.L. Ma,
- H. Li, J.B. Li, Effect of dirhamnolipid on the removal of phenol catalyzed
 by laccase in aqueous solution, World Journal of Microbiology &
 Biotechnology 28(2012) 175.
- [33] X. Ren, G. Zeng, L. Tang, J. Wang, J. Wan, H. Feng, B. Song, C.
- Huang, X. Tang, Effect of exogenous carbonaceous materials on the
- bioavailability of organic pollutants and their ecological risks, Soil Biology
- 674 & Biochemistry 116 (2018) 70-81.
- [34] G. Zeng, W. Jia, D. Huang, H. Liang, H. Chao, C. Min, W. Xue, X.
- 676 Gong, R. Wang, D. Jiang, Precipitation, adsorption and rhizosphere effect:

- the mechanisms for Phosphate-induced Pb immobilization in soils-A
 review, Journal of hazardous materials 339 (2017) 354.
- [35] S. Kavitha, P.B.C. Stella, S. Kaliappan, I.T. Yeom, J.R. Banu,
 Enhancement of anaerobic degradation of sludge biomass through
 surfactant-assisted bacterial hydrolysis, Process Safety and Environmental
 Protection 99 (2016) 207-215.
- [36] E.F. Khalil, T.N. Whitmore, H. Gamal-El-Din, A. El-Bassel, D. Lloyd,
- ⁶⁸⁴ The effects of detergents on anaerobic digestion, Appled Microbiology &
- 685 Biotechnology 29 (1988) 517-522.
- [37] Z.F. Liu, G.M. Zeng, J. Wang, H. Zhong, Y. Ding, X.Z. Yuan, Effects
- 687 of monorhamnolipid and Tween 80 on the degradation of phenol by 688 Candida tropicalis, Process Biochemistry 45 (2010) 805-809.
- [38] A.K. Bhuyan, On the mechanism of SDS-induced protein denaturation,
- 690 Biopolymers 93 (2010) 86-199.
- [39] S. De, A. Girigosvami, S. Das, Fluorescence probing of albumin–
 surfactant interaction, Journal of Colloid and Interface Science 285 (2005)
 562-573.
- 694 [40] M.V. And, D. Angelescu, M.A. And, A. Valstar, Interactions of
- Globular Proteins with Surfactants Studied with Fluorescence ProbeMethods, Langmuir (2008).
- [41] R.-C. Lu, A.-N. Cao, L.-H. Lai, J.-X. Xiao, Effect of anionic surfactant
- 698 molecular structure on bovine serum albumin (BSA) fluorescence,

- Colloids and Surfaces A: Physicochemical and Engineering Aspects 278(2006) 67-73.
- 701 [42] E. Liwarska-Bizukojc, M. Bizukojc, Digital image analysis to
- estimate the influence of sodium dodecyl sulphate on activated sludge flocs,
- 703 Process Biochemistry 40 (2005) 2067-2072.
- [43] Z.F. Liu, G.M. Zeng, H. Zhong, X.Z. Yuan, L.L. Jiang, H.Y. Fu, X.L.
- Ma, J.C. Zhang, Effect of saponins on cell surface properties of Penicillium
- simplicissimum : Performance on adsorption of cadmum(II), Colloids &
- 707 Surfaces B Biointerfaces 86 (2011) 364-369.
- [44] Q. Yuan, R. Sparling, J.A. Oleszkiewicz VFA generation from waste
- activated sludge: effect of temperature and mixing, Chemosphere 82 (2011)
 603-607.
- 711 [45] X. Zheng, Y. Su, X. Li, Xieo, D. Wang, Y. Chen, Pyrosequencing
- reveals the key microorganisms involved in sludge alkaline fermentation
 for efficient short-chain fatty acids production, Environ Sci Technol 47
 (2013) 4262-4268
- [46] T.N. Zhilina, G.A. Zavarzin, Alkaliphilic anaerobic community at pH
- ⁷¹⁶ 10, Current Microbiology 29 (1994) 109-112.
- [47] B. Shao, Z. Liu, H. Zhong, G. Zeng, G. Liu, M. Yu, Y. Liu, X. Yang,
- 718 Z. Li, Z. Fang, Effects of rhamnolipids on microorganism characteristics
- and applications in composting: A review, Microbiological Research 200
- 720 (2017) 33-44.

[48] Alexander, K.A. Walters, Interactions of Nonionic Polyoxyethylene
Alkyl and Aryl Ethers with Membranes and Other Biological Systems,
ACS Symposium Series, 1984, pp. 189-207.

[49] V.A. Shcherbakova, K.S. Laurinavichius, V.K. Akimenko, Toxic
effect of surfactants and probable products of their biodegradation on
methanogenesis in an anaerobic microbial community, Chemosphere 39
(1999) 1861-1870.

[50] L. Bertin, C. Bettini, G. Zanaroli, S. Fraraccio, A. Nogroni, F. Fava,
Acclimation of an anaerobic consortium consists of effective
biomethanization of mechanically : orted organic fraction of municipal
solid waste through a semi > ontinuous environment procedure, Journal of
Chemical Technology & Biotechnology \$7 (2012) 1312-1319.

[51] A. Zhou, W. Liu, C. Varrene, Y. Wang, A. Wang, X. Yue, Evaluation
of surfactants on waste activated sludge fermentation by pyrosequencing
analysis, Biorecource technology 192 (2015) 835-840.

[52] M.I. Van Dyke, P. Couture, M. Brauer, H. Lee, J.T. Trevors,
Pseudomonas aeruginosa UG2 rhamnolipid biosurfactants: structural
characterization and their use in removing hydrophobic compounds from
soil, Canadian Journal of Microbiology 39 (1993) 1071-1078.

[53] X. Huang, T. Mu, C. Shen, L. Lu, J. Liu, Effects of bio-surfactants
combined with alkaline conditions on volatile fatty acid production and
microbial community in the anaerobic fermentation of waste activated

- ⁷⁴³ sludge, International Biodeterioration & Biodegradation 114 (2016) 24-30.
- 744 [54] R. Sun, A. Zhou, J. Jia, Q. Liang, Q. Liu, D. Xing, N. Ren,
- 745 Characterization of methane production and microbial community shifts
- ⁷⁴⁶ during waste activated sludge degradation in microbial electrolysis cells,
- 747 Bioresource technology 175 (2015) 68-74.
- [55] O. Nybroe, P.E. Jørgensen, M. Henze, Enzyme activities in waste
 water and activated sludge, Water Research 26 (1992) 579-584.
- 750 [56] R.J. Chrost, Characterization and Significance of \$\beta-
- 751 Glucosidase\$ Activity in Lake Water, Limnology & Ceanography 34
- 752 (1989) 660-672.
- [57] R.J. Chróst, R. Wcisło, G.Z. Halemejko, Enzymatic decomposition of
 organic matter by bacteria in an eutrophic lake, Archiv Fur Hydrobiologie
- 755 107 (1986).
- [58] A.L. Rosso, F. Azam, Proteolytic activity in coastal oceanic waters:
 Depth distribution and relationship to bacterial populations, Marine
 Ecology Progress 11 (1987) 231-240.
- [59] J. Schnürer, T. Rosswall, Fluorescein diacetate hydrolysis as a
 measure of total microbial activity in soil and litter, Applied &
 Environmental Microbiology 43 (1982) 1256-1261.
- [60] T. Cserháti, E. Forgács, G. Oros, Biological activity and
 environmental impact of anionic surfactants, Environment International 28
 (2002) 337.

[61] M.R. Housaindokht, A.A. Moosavi-Movahedi, J. Moghadasi, M.N.
Jones, Interaction of glucose oxidase with ionic surfactants: a
microcalorimetric study, International Journal of Biological
Macromolecules 15 (1993) 337-341.

[62] Zhang., G. Zeng, L. Tang, J. Chen, Y. Zhu, X. He, Y. He,
Electrochemical Sensor Based on Electrodeposited Graphene-Au
Modified Electrode and NanoAu Carrier Amplified Signal Strategy for
Attomolar Mercury Detection, Analytical Chemistry 87 (2015) 989-996.
[63] A.S. Ucisik, M. Henze, Biological hydrolysis and acidification of
sludge under anaerobic conditions: the effect of sludge type and origin on

- the production and composition of volatile fatty acids, Water Res 42 (2008)
 3729-3738.
- [64] C. Ting, D. Lee, Production of hydrogen and methane from
 wastewater sludge using anaerobic fermentation, International Journal of
 Hydrogen Energy 32 (2007) 677-682.

[65] R. Guan, X Yuan, Z. Wu, H. Wang, L. Jiang, Y. Li, G. Zeng,
Functionality of surfactants in waste-activated sludge treatment: A review,
The Science of the total environment 609 (2017) 1433-1442.

[66] A. Cadoret, A. Conrad, J.C. Block, Availability of low and high
molecular weight substrates to extracellular enzymes in whole and
dispersed activated sludges, Enzyme & Microbial Technology 31 (2002)
179-186.

- [67] J.B. Russell, S.A. Martin, Effects of Various Methane Inhibitors on
 the Fermentation of Amino Acids by Mixed Rumen Microorganisms in
 Vitro, Journal of Animalence 59 (1984) 1329-1338.
- 790 [68] W. Chen, P. Westerhoff, J.A. Leenheer, K. Booksh, Fluorescence
- 791 excitation-emission matrix regional integration to quantify spectra for
- dissolved organic matter, Environmental Science & Technology 37 (2003)
 5701.
- [69] Y. Yamashita, E. Tanoue, Chemical characterization of protein-like
- 795 fluorophores in DOM in relation to aromatic variation acids, Marine
- 796 Chemistry 82 (2003) 255-271.
- [70] G.P. Sheng, H.Q. Yu, Characterization of extracellular polymeric
 substances of aerobic and anaerobic sludge using three-dimensional
 excitation and emission matrix fluorescence spectroscopy, Water Research
 40 (2006) 1233-1239.
- [71] Q. Wang, Kuninobu, H.I. Ogawa, Y. Kato, Degradation of volatile
 fatty acids in highly efficient anaerobic digestion, Biomass and Bioenergy
 16 (1999) 407-416.
- 804 [72] H. Yuan, Y. Chen, H. Zhang, S. Jiang, Q. Zhou, G. Gu, Improved
- 805 bioproduction of short-chain fatty acids (SCFAs) from excess sludge under
- alkaline conditions, Environmental Science & Technology 40 (2006) 2025.
- [73] M. Öztürk, Conversion of acetate, propionate and butyrate to methane
- under thermophilic conditions in batch reactors, Water Research 25 (1991)

809 1509-1513.

[74] L.Y. Feng, Y.G. Chen, Z. Xiong, Enhancement of waste activated
sludge protein conversion and volatile fatty acids accumulation during
waste activated sludge anaerobic fermentation by carbohydrate substrate
addition: the effect of pH, Environmental Science & Technology 43 (2009)
4373-4380.

815 [75] Z.K. Erdal, C.W. Randall, E.M. Gregory, CHAPTER III:

816 BIOCHEMISTRY OF THE ENHANCED BIOLOGICAL PHOSPHORUS

- 817 REMOVAL SYSTEMS, Proceedings of the Water Expression Federation
- 818 2003 (2003) 591-599(599).

[76] M. Steup, C. Schächtele, Analysis of the microbial community
structure and function of a laboratory scale enhanced biological
phosphorus removal reactor Environmental Microbiology 4 (2010) 559569.

[77] Y. Chen, A. Rindall, T. McCue, The efficiency of enhanced
biological phospherus removal from real wastewater affected by different
ratios of acetic to propionic acid, Water Res 38 (2004) 27-36.

826 [78] M. Beer, Y.H. Kong, R.J. Seviour, Are some putative glycogen 827 accumulating organism (GAO) in anaerobic: Aerobic activated sludge 828 systems members of the α -Proteobacteria?, Microbiology 155 (2009) 829 2267-2275.

830 [79] M.T. Wong, F.M. Tan, W.J. Ng, W.T. Liu, Identification and

occurrence of tetrad-forming Alphaproteobacteria in anaerobic-aerobic
activated sludge processes, Microbiology 150 (2004) 3741-3748.

[80] S.R. Harper, F.G. Pohland, Recent developments in hydrogen
management during anaerobic biological wastewater treatment,
Biotechnology & Bioengineering 28 (2010) 585-602.

836 [81] H.Q. Yu, X.J. Zheng, Z.H. Hu, G.W. Gu, High-rate anaerobic

hydrolysis and acidogenesis of sewage sludge in a modified upflow reactor,

- 838 Water Science & Technology A Journal of the International Association on
- Water Pollution Research 48 (2003) 69.
- [82] J. Zhao, Q. Yang, X. Li, D. Wang, K. Lux Y. Zhong, Q. Xu, G. Zeng,

Enhanced production of short-chain fatty acid from food waste stimulated by alkyl polyglycosides and its mechanism, Waste management 46 (2015)

843 133-139.

[83] Q. Jia, H. Wang, X. Wang, Dynamic synthesis of
polyhydroxyall moales by bacterial consortium from simulated excess
sludge fermentation liquid, Bioresource technology 140 (2013) 328-336.

[84] H.M. Poggi-Varaldo, R. Rodríguez-Vázquez, G. FernándezVillagómez, F. Esparza-García, Inhibition of mesophilic solid-substrate
anaerobic digestion by ammonia nitrogen, Applied Microbiology &
Biotechnology 47 (1997) 284-291.

[85] H.M. Poggivaraldo, E. Arcemedina, G. Fernandezvillagomez, S.
Caffarelmendez, Inhibition of mesophilic solid substrate anaerobic

digestion (DASS) by ammonia-rich wastes, Industrial Waste Conference,
1998.

- [86] D.Y. Okada, T.P. Delforno, A.S. Esteves, J. Polizel, J.S. Hirasawa, I.C.
- 856 Duarte, M.B. Varesche, Influence of volatile fatty acid concentration
- stability on anaerobic degradation of linear alkylbenzene sulfonate, Journal
- of environmental management 128 (2013) 169-172.
- 859 [87] H. Feitkenhauer, Anaerobic digestion of desizing wastewater:
- influence of pretreatment and anionic surfactant on degradation and
 intermediate accumulation, Enzyme & Microbial Compology 33 (2003)
 250-258.
- [88] X. Yi, K. Luo, Q. Yang, X.M. Li, W.G. Deng, H.B. Cheng, Z.L. Wang,
- 64 G.M. Zeng, Enhanced hydrolysis and acidification of waste activated sludge by biosurfactant changelipid, Applied biochemistry and biotechnology 171 (2011) 1416-1428.
- [89] Zhang., X. Li, Cha, L. Dai, J. Zhao, Y. Chen, X. Dai, A review:
 factors affecting occess sludge anaerobic digestion for volatile fatty acids
 production, Water science and technology : a journal of the International
 Association on Water Pollution Research 72 (2015) 678-688.
- [90] H.D. Ariesyady, T. Ito, S. Okabe, Functional bacterial and archaeal
 community structures of major trophic groups in a full-scale anaerobic
 sludge digester, Water Res 41 (2007) 1554-1568.
- 874 [91] Y. Chen, S. Jiang, H. Yuan, Q. Zhou, G. Gu, Hydrolysis and

acidification of waste activated sludge at different pHs, Water Res 41 (2007)
683-689.

[92] P. Zhang, Y. Chen, Q. Zhou, Waste activated sludge hydrolysis and
short-chain fatty acids accumulation under mesophilic and thermophilic
conditions: effect of pH, Water Res 43 (2009) 3735-3742.

[93] J.H. Patil, M.A. Raj, P.L. Muralidhara, S.M. Desai, G.K.M. Raju,

Kinetics of anaerobic digestion of water hyacinth using poultry litter as
inoculum, International Journal of Environmental Science & Development
3 (2012) 94-98.

[94] D.S. Yoo, B.S. Lee, E.K. Kim, paracteristics of microbial
biosurfactant as an antifungal agent against plant pathogenic fungus,
Journal of Microbiology & Biotechnology 15 (2005) 1164-1169.

[95] M. Chen, P. Xu, G. Zene, C. Yang, D. Huang, J. Zhang,
Bioremediation of suils contaminated with polycyclic aromatic
hydrocarbons, etroleur, pesticides, chlorophenols and heavy metals by
composting: Applications, microbes and future research needs,
Biotechnology Advances 33 (2015) 745.

[96] X. Ren, G. Zeng, L. Tang, J. Wang, J. Wan, Y. Liu, J. Yu, H. Yi, S. Ye,

- R. Deng, Sorption, transport and biodegradation An insight into
 bioavailability of persistent organic pollutants in soil, Science of the Total
 Environment 610-611 (2017) 1154-1163.
- 896 [97] M. Blanchard, M.J. Teil, D. Ollivon, L. Legenti, M. Chevreuil,

- Polycyclic aromatic hydrocarbons and polychlorobiphenyls in wastewaters
 and sewage sludges from the Paris area (France), Environmental research
 95 (2004) 184-197.
- 900 [98] J.L. Gong, B. Wang, G.M. Zeng, C.P. Yang, C.G. Niu, Q.Y. Niu, W.J.
- 201 Zhou, Y. Liang, Removal of cationic dyes from aqueous solution using
- magnetic multi-wall carbon nanotube nanocomposite as adsorbent, Journal
 of hazardous materials 164 (2009) 1517-1522.
- [99] H. Wu, C. Lai, G. Zeng, J. Liang, J. Chen, J. Xu, Dai X. Li, J. Liu,
 M. Chen, The interactions of composting and blochar and their
 implications for soil amendment and pollution remediation: a review,
 Critical Reviews in Biotechnology 37 (2017) 754.
- 908 [100] Z. Yu, C. Zhang, Z. Zheng, L. Nu X. Li, Z. Yang, C. Ma, G. Zeng,
- 909 Enhancing phosphate adsorption capacity of SDS-based magnetite by
- surface modification of citric acid, Applied Surface Science 403 (2017)
- 911 413-425.

[101] Q. Aemig, Čheron, N. Delgenes, J. Jimenez, S. Houot, J.P. Steyer,
D. Patureau, Distribution of Polycyclic Aromatic Hydrocarbons (PAHs) in
sludge organic matter pools as a driving force of their fate during anaerobic
digestion, Waste management 48 (2016) 389-396.

- 916 [102] F. Long, J.L. Gong, G.M. Zeng, L. Chen, X.Y. Wang, J.H. Deng, Q.Y.
- 917 Niu, H.Y. Zhang, X.R. Zhang, Removal of phosphate from aqueous
- solution by magnetic Fe–Zr binary oxide, Chemical Engineering Journal

919 171 (2011) 448-455.

920 [103] J. Liang, Z. Yang, L. Tang, G. Zeng, M. Yu, X. Li, H. Wu, Y. Qian,

X. Li, Y. Luo, Changes in heavy metal mobility and availability from
contaminated wetland soil remediated with combined biochar-compost,
Chemosphere 181 (2017) 281.

- 924 [104] T.S. Poeton, H.D. Stensel, S.E. Strand, Biodegradation of
 925 polyaromatic hydrocarbons by marine bacteria: effect of solid phase on
 926 degradation kinetics, Water Research 33 (1999) 868-800
- 927 [105] W.W. Tang, G.M. Zeng, J.L. Gong, J. Liang, K. C. Zhang, B.B.
- Huang, Impact of humic/fulvic acid on the removal of heavy metals from
 aqueous solutions using nanomaterials
 review, Science of the Total
 Environment 468-469 (2014) 1014.

931 [106] J.H. Deng, X.R. Zhang, G.Y. Zeng, J.L. Gong, Q.Y. Niu, J. Liang,

Simultaneous removal (f Gd(II) and ionic dyes from aqueous solution
using magnetic graphene oxide nanocomposite as an adsorbent, Chemical
Engineering Journal 226 (2013) 189-200.

935 [107] C. Zhou, C. Lai, D. Huang, G. Zeng, C. Zhang, M. Cheng, L. Hu, J.

Wan, W. Xiong, M. Wen, Highly porous carbon nitride by supramolecular

- 937 preassembly of monomers for photocatalytic removal of sulfamethazine
- ⁹³⁸ under visible light driven, Applied Catalysis B Environmental 220 (2017).
- 939 [108] A.A. Juwarkar, A. Nair, K.V. Dubey, S.K. Singh, S. Devotta,
- 940 Biosurfactant technology for remediation of cadmium and lead

- outaminated soils, Chemosphere 68 (2007) 1996.
- 942 [109] P. Xu, G.M. Zeng, D.L. Huang, C. Lai, M.H. Zhao, Z. Wei, N.J. Li,
- C. Huang, G.X. Xie, Adsorption of Pb(II) by iron oxide nanoparticles
 immobilized Phanerochaete chrysosporium : Equilibrium, kinetic,
 thermodynamic and mechanisms analysis, Chemical Engineering Journal
 203 (2012) 423-431.
- 947 [110] M. Cheng, G. Zeng, D. Huang, L. Cui, P. Xu, C. Zhang, Y. Liu,
- 948 Hydroxyl radicals based advanced oxidation processes (AOPs) for
- 949 remediation of soils contaminated with organic compounds: A review,
- 950 Chemical Engineering Journal 284 (2016) 58 598.
- 951 [111] C. Zhang, C. Lai, G. Zeng, D. Huang, L. Tang, C. Yang, Y. Zhou, L.
- 952 Qin, M. Cheng, Nanoporous Au-based hronocoulometric aptasensor for
- 953 amplified detection of Pb(2), using DNAzyme modified with Au
- nanoparticles, Biosensons & Bioelectronics 81 (2016) 61-67.
- [112] C. Zhang, Z.O. YJ, G.M. Zeng, M. Jiang, Z.Z. Yang, F. Cui, M.Y.
 Zhu, L.Q. Shen, L. Hu, Effects of sediment geochemical properties on
 heavy metal bioavailability, Environment International 73 (2014) 270.
- 958
- 959