

**Core-shell structured nanoparticles for photodynamic therapy-based cancer treatment and related imaging**

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## 17 Abstract

18 Photodynamic therapy (PDT) is an emerging noninvasive therapy modality for treating cancer  
19 diseases. However, conventional PDT suffers from poor stability of organic photosensitizers, limited  
20 tissue penetration depth of excitation light and hypoxic tumor microenvironment, which hinders its  
21 modern clinical applications. The combination of PDT and nanotechnology is becoming a promising  
22 technology to tackle these troubles. Core-shell structured nanoparticles are of great interest as they  
23 can integrate the functionalities of individual components into one structure and exhibit improved  
24 physical and chemical properties that are different from the single component. Therefore, many  
25 efforts have been paid to develop core-shell structured nanoparticles for PDT of cancer. This review  
26 provides a panorama of the latest achievement in the developments of core-shell structured  
27 nanoparticles for PDT-based cancer treatment and related imaging. Concretely, this review starts with  
28 the categories of core-shell structured nanoparticles, followed by the functions of these nanoparticles  
29 in PDT of cancer, including photosensitizer delivery vehicle, energy transducer, photosensitizer and  
30 hypoxic tumor microenvironment modulator. Then the applications of core-shell structured particles  
31 for photodynamic synergistic therapy of cancer are highlighted as well as their imaging applications  
32 as contrast agents. Finally, perspectives on the major challenges and opportunities are presented for  
33 better developments in the future research.

34 **Keywords:** Core-shell; Photodynamic therapy; Cancer treatment; Synergistic therapy; Imaging

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36	<b>Contents</b>	
37	<b>1. Introduction</b>	4
38	<b>2. Categories of core-shell structured nanoparticles</b>	6
39	2.1. Inorganic core-shell structured nanoparticles	9
40	2.2. Organic core-shell structured nanoparticles	11
41	2.3. Hybrid core-shell structured nanoparticles	12
42	<b>3. Core-shell structured nanoparticles for PDT of cancer</b>	13
43	3.1. Photosensitizer delivery vehicles	14
44	3.2. Energy transducers	16
45	3.3. Photosensitizers	19
46	3.4. Hypoxic tumor microenvironment modulators	22
47	<b>4. Core-shell structured nanoparticles for photodynamic synergistic therapy of cancer</b>	27
48	4.1. PDT combined with chemotherapy	29
49	4.2. PDT combined with photothermal therapy	33
50	4.3. PDT combined with immunotherapy	38
51	4.4. PDT combined with other therapies	41
52	<b>5. Core-shell structured nanoparticles for imaging in PDT-based cancer treatment</b>	45
53	5.1. Optical imaging	46
54	5.2. Photothermal imaging	48
55	5.3. Photoacoustic imaging	49
56	5.4. Magnetic resonance imaging	49
57	5.5. Computed tomography imaging	51
58	<b>6. Conclusions and perspectives</b>	52
59	<b>Acknowledgements</b>	53
60	<b>Reference</b>	53

## 1. Introduction

Cancer, a disease that poses serious threats to the health of human beings, is becoming one of the leading causes of death globally. In 2020, nearly 19.3 million new cancer cases were diagnosed, and about 10.0 million cancer cases deaths occurred [1]. Due to the high risk and mortality of cancer, researchers all over the world have been working to develop effective therapies to treat cancer [2-4]. Conventional cancer therapies mainly include surgery, chemotherapy and radiotherapy, and they have some inescapable shortcomings. For example, surgery usually requires the cooperation of chemotherapy or radiotherapy to completely remove the cancer cells, and some tumors may recur after surgery [5, 6]. Chemotherapy inhibits the cell division, leading to some side effects, such as alopecia and myelosuppression [7, 8]. Besides, radiotherapy is restricted by the radiation site and cumulative radiation dose [9, 10]. Accordingly, although improvement of the traditional cancer therapy modalities is important, it is also necessary to develop alternate therapy modalities that are safer, more effective and more affordable.

Photodynamic therapy (PDT) is an ideal cancer treatment method that can kill cancer cells through reactive oxygen species (ROS) generated by a photosensitizer under light irradiation [11, 12]. Compared with traditional cancer treatment methods, PDT has the advantages of high safety, good repeatability, low long-term morbidity and high life quality of patients [13-15]. Normally, PDT consists of three essential components: photosensitizer, excitation light and molecular oxygen ( $O_2$ ) [16]. These components are not toxic alone, but together they will trigger a photochemical reaction to produce cytotoxic ROS. Under the irradiation of light with a specific wavelength, photosensitizer can be excited and then react with substrates and  $O_2$  to generate free radicals, such as superoxide radical ( $\cdot O_2^-$ ) and hydroxyl radical ( $\cdot OH$ ) (type I reaction). Alternatively, the excited photosensitizer can directly transfer its energy to  $O_2$  to form highly reactive singlet oxygen ( $^1O_2$ ), resulting in the

87 significant cellular toxicity (type II reaction) [17]. Notably, three interrelated mechanisms are  
88 involved in the tumor destruction by PDT: direct tumor cell kill, vascular damage and immune  
89 response [11]. Since the significant breakthrough made in 1975 by Dougherty and co-workers [18],  
90 PDT has been proved to be effective in treating various cancers, such as skin cancer, head and neck  
91 cancers, and superficial bladder cancer.

92 Despite the extensive research and rapid growth, photodynamic cancer therapy still has some  
93 limitations in the modern clinical applications [19-21]. Typically, the traditional small organic  
94 molecule photosensitizers present poor stability and low targeting ability, which will reduce the  
95 efficiency of PDT and may evoke serious side effects [22, 23]. The excitation wavelength of most  
96 photosensitizers is in the visible light region, which will result in the limited tissue penetration depth  
97 and thus hinder the wide application of PDT [24]. Moreover, the hypoxic tumor microenvironment  
98 induced by O<sub>2</sub> consumption will also affect the sustained effect of PDT [25, 26]. In recent years, the  
99 introduction of nanoparticles into PDT has become a promising strategy to resolve these issues [27-  
100 29]. Among the various types of nanoparticles, the core-shell structured nanoparticles have stimulated  
101 great research interest as they can integrate the functionalities of individual components into one  
102 structure and exhibit improved physical and chemical properties that are different from the single  
103 component [30-32]. Meanwhile, the active interfaces between different components in core-shell  
104 structured nanoparticles may produce synergistic effects and novel properties [33, 34]. For example,  
105 some biomolecules shells could not only stabilize the photosensitizers in biological fluids and extend  
106 their blood circulation time, but also provide the ability to actively target tumor sites [35-37]. The  
107 lanthanide-doped upconversion nanoparticles (UCNPs) cores could absorb near-infrared (NIR) light  
108 and convert it to ultraviolet-visible (UV-VIS) light, thereby exciting the photosensitizers loaded in  
109 the shells [38-40]. Compared with the UCNPs/metal-organic frameworks (MOFs) nanocomposite

with Janus structure, the distance between UCNPs and MOFs in core-shell structured UCNPs@MOFs nanoparticle is shorter, which could enhance the energy transfer efficiency from the UCNPs to the MOFs under NIR light irradiation, thus promoting the  $^1\text{O}_2$  generation and improving the PDT efficacy [41-43]. Most importantly, the incorporation of functional materials or agents could enable core-shell structured nanoparticles to be multifunctional nanoplatforms for synergistic therapy and imaging [44-46]. These unique core-shell structured nanoparticles have been widely applied in photodynamic cancer therapy, but lacking a systematic understanding and overview.

This review aims to summarize the recent progress of core-shell structured nanoparticles in PDT-based cancer treatment and related imaging. First, the categories of core-shell structured nanoparticles are introduced according to the material compositions of the core and shell. Second, the functions of core-shell structured nanoparticles in PDT of cancer are comprehensively summarized. Then the achievements of core-shell structured nanoparticles in photodynamic synergistic therapy of cancer are discussed in detail. Additionally, the applications of core-shell structured nanoparticles in imaging during the PDT-based cancer treatment are mentioned. Ultimately, a brief conclusion and some perspectives on the future developments of this area are presented.

## 2. Categories of core-shell structured nanoparticles

In the past decades, various strategies have been developed to prepare core-shell structured nanoparticles because of the great application potentials of core-shell structures in many fields, such as biomedicine [47, 48], energy utilization [49, 50], catalysis [51, 52], etc. Meanwhile, there are some excellent reviews that have summarized the synthesis of core-shell structured nanoparticles in detail [53-55]. Therefore, here we do not intend to provide a repeated summary on the synthesis of core-shell structured nanoparticles, but rather to briefly introduce the categories of core-shell structured nanoparticles in PDT of cancer (Table 1). Broadly, a core-shell structured nanoparticle is composed

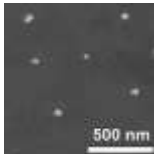
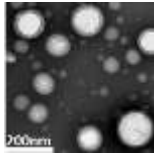

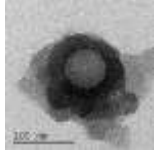
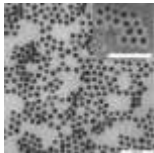

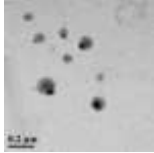

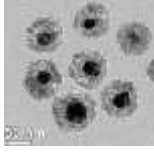
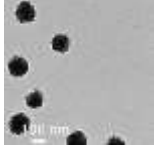
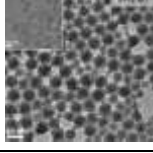
133 of an inner core and an outer shell. According to material compositions of the core and shell, core-  
 134 shell structured nanoparticles can be classified into three categories: inorganic, organic and hybrid.

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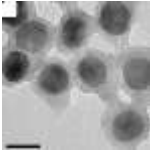
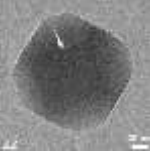
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137 Table 1. Categories of core-shell structured nanoparticles in PDT of cancer.

Nanoparticle	Morphology	Synthesis methods		Functions	Ref.
		Core	Shell		
Class I: Inorganic core-shell structured nanoparticles					
UCNPs@mSiO <sub>2</sub>		Thermal decomposition method	Sol-gel method	Photosensitizer (RB) delivery vehicle Energy transducer	[37]
UCNPs@mSiO <sub>2</sub>		Coprecipitation method	Sol-gel method	Photosensitizer (ZnPc) delivery vehicle Energy transducer	[56]
AuNR@SiO <sub>2</sub>		Seed-mediated growth method	Sol-gel method	Photosensitizer (HB) delivery vehicle	[57]
Fe <sub>3</sub> O <sub>4</sub> @SiO <sub>2</sub> @mSiO <sub>2</sub>		Solvothermal method	Sol-gel method	Photosensitizer (AlC <sub>4</sub> Pc) delivery vehicle	[58]
UPCNs@TiO <sub>2</sub>		Thermal decomposition method	Solvothermal method	Energy transducer Photosensitizer (TiO <sub>2</sub> )	[59]
SiO <sub>2</sub> @MnO <sub>2</sub>		Sol-gel method	Reduction method	Photosensitizer (MB) delivery vehicle Hypoxic tumor microenvironment modulator	[60]
Cu <sub>2-x</sub> S@MnS		One-pot hot-injection method		Photosensitizer (Cu <sub>2-x</sub> S) Hypoxic tumor microenvironment modulator	[61]
UCNPs@CaF <sub>2</sub>		Thermal decomposition method	Epitaxial growth method	Photosensitizer (PpIX) delivery vehicle Energy transducer	[62]
Class II: Organic core-shell structured nanoparticles					

BDPVDA@mPEG-PPDA		Self-assembly method		Photosensitizer (BODIPY)	[63]
Oil@lipid		Sonication method		Photosensitizer (porphyrin-lipid)	[64]
PFTBA@HSA		Ultrasonic emulsification method		Photosensitizer (IR780) delivery vehicle	[65]
RC@PDA		Solvent exchange and nucleation	Self-oxidation and polymerization	Photosensitizer (Ce6) delivery vehicle	[66]
Class III: Hybrid core-shell structured nanoparticles					
Au@PDMS-PEG		Reduction method	Hydrosilylation method	Photosensitizer (HB) delivery vehicle	[35]
Au@PDA		Seed-mediated growth method	Polymerization method	Photosensitizer (DSBDP) delivery vehicle	[67]
Ag@PANI		Reduction method	Polymerization method	Photosensitizer (ICG) delivery vehicle	[68]
Fe <sub>3</sub> O <sub>4</sub> @COFs		Solvothermal method	Templated method	Photosensitizer (COFs)	[69]
UCNPs@g-C <sub>3</sub> N <sub>4</sub>		Thermal decomposition method	Polymerization method	Energy transducer Photosensitizer (g-C <sub>3</sub> N <sub>4</sub> )	[70]
PDA@UCNPs		Polymerization method	Coprecipitation method	Photosensitizer (Ce6) delivery vehicle Energy transducer	[71]
SPN@MnO <sub>2</sub>		Precipitation method	Reduction method	Photosensitizer (PCPDTBT) Hypoxic tumor microenvironment modulator	[72]



AuNR@MOFs		Seed-mediated growth method	Solvothermal method	Photosensitizer (MOFs)	[73]
UCNPs@MOFs		Thermal decomposition method	Precipitation method	Energy transducer Photosensitizer (MOFs)	[42]
ZIF-67@ZIF-8		Precipitation method	Precipitation method	Photosensitizer (PpIX) delivery vehicle Hypoxic tumor microenvironment modulator	[74]

138

## 139 2.1. Inorganic core-shell structured nanoparticles

140 **Inorganic core-shell structured nanoparticles are the most important kind of the three types.** The  
141 cores of the inorganic core-shell structured nanoparticles used for PDT of cancer are usually made of  
142 UCNPs [75, 76], metals [77, 78], metal oxides [79, 80] and sulfides [81, 82], while their shells are  
143 mainly composed of SiO<sub>2</sub> [75, 79, 82], metal oxides [76, 81] and sulfides [61, 83], and CaF<sub>2</sub> [62, 84].  
144 Notably, the SiO<sub>2</sub> coating can endow inorganic cores with low bulk conductivity and high suspension  
145 stability [53, 85]. **Moreover, because of the controllable pore structure, feasible functionalization and**  
146 **excellent biocompatibility, the coated SiO<sub>2</sub> shell can serve as a carrier to deliver photosensitizers [86,**  
147 **87].** Therefore, as a typical shell material, the SiO<sub>2</sub> has attracted much interest in recent years. For  
148 example, to avoid the shift of surface plasmon resonance (SPR) peak of gold nanorods (AuNR) from  
149 NIR region to visible light region, Qin et al. deposited a mesoporous SiO<sub>2</sub> (mSiO<sub>2</sub>) shell on the surface  
150 of AuNR for preventing their aggregation under NIR laser irradiation [57]. After incorporating a  
151 hypocrellin B (HB) photosensitizer into the mSiO<sub>2</sub> shell, the AuNR@mSiO<sub>2</sub>-HB nanoparticles  
152 presented great potential in synergistic PDT/photothermal therapy (PTT). In the study of Xu et al., a  
153 NaGdF<sub>4</sub>:Yb,Er@NaGdF<sub>4</sub>:Nd,Yb core was coated with a mSiO<sub>2</sub> shell containing dual-photosensitizer  
154 for PDT [88]. The chlorin e6 (Ce6) and merocyanine 540 (MC540) photosensitizers were loaded into

155 the mSiO<sub>2</sub> shell through covalent bond and electrostatic interaction, respectively. As a consequence,  
156 the mSiO<sub>2</sub> shell not only enabled the nanoparticles to have a high photosensitizer loading, but also  
157 avoided the direct contact between photosensitizers and cells in organism, thereby protecting them  
158 from the *in vivo* microenvironment.

159 Apart from SiO<sub>2</sub>, metal oxides and sulfides including TiO<sub>2</sub> [89, 90], ZnO [91], MnO<sub>2</sub> [92], CeO<sub>2</sub>  
160 [81], ZrO<sub>2</sub> [93], MnS [61] and FeS [83], and CaF<sub>2</sub> [94] have also been employed as the shell materials  
161 of inorganic core-shell structured nanoparticles for PDT of cancer. For example, TiO<sub>2</sub> is a desirable  
162 photosensitizer as it can be maintained for a long time in human body and is nontoxic and stable  
163 without light irradiation [95]. In the study of Hou et al., a TiO<sub>2</sub> shell was coated on the surface of  
164 NaYF<sub>4</sub>:Yb,Tm@NaGdF<sub>4</sub>:Yb core for PDT [59]. The direct contact between TiO<sub>2</sub> shell and UCNPs  
165 core could ensure the maximum energy transfer from UCNPs to TiO<sub>2</sub>, thereby accelerating the  
166 production and release of ROS. MnO<sub>2</sub> has a high O<sub>2</sub> generation ability in acidic and H<sub>2</sub>O<sub>2</sub>-rich tumor  
167 microenvironment, making it a good candidate for alleviating tumor hypoxia and enhancing PDT  
168 efficacy [96]. Li et al. developed a core-shell structured nanoparticle consisted of a hollow  
169 mesoporous CuS core loaded with the Ce6 photosensitizer and a MnO<sub>2</sub> shell for PDT/PTT [97]. The  
170 MnO<sub>2</sub> shell not only acted as a modulator to effectively alleviate tumor hypoxia, but also served as a  
171 gatekeeper to prevent the premature release of loaded Ce6. ZrO<sub>2</sub> can be utilized in imaging-guided  
172 therapy owing to its excellent biocompatibility and effective imaging ability [98]. Feng et al.  
173 fabricated UCNPs@ZrO<sub>2</sub> nanoparticles to load Ce6 photosensitizer, doxorubicin (DOX) and  
174 tetradecanol for multimodal imaging-guided PDT/PTT/chemotherapy [93]. The hollow and  
175 mesoporous ZrO<sub>2</sub> shell endowed the UCNPs@ZrO<sub>2</sub> nanoparticles with superior drug delivery  
176 capacity and satisfactory computed tomography (CT) imaging performance. Moreover, the CaF<sub>2</sub> shell  
177 can strengthen the upconversion luminescent intensity of UCNPs core and prevent the leakage of rare

178 earth ions in UCNPs core [99]. A core-shell structured NaYF<sub>4</sub>:Yb,Er@CaF<sub>2</sub> nanoparticle was  
179 fabricated by Punjabi et al. for *in vivo* deep tumor PDT treatment [62].

## 180 2.2. Organic core-shell structured nanoparticles

181 Both cores and shells of organic core-shell structured nanoparticles are made of polymers or  
182 other organic materials. Owing to the good biodegradability and high drug encapsulation efficiency,  
183 they are widely applied to the controlled release of photosensitizers in PDT of cancer [100].  
184 Meanwhile, encapsulating photosensitizers in these nanoparticles can significantly increase the  
185 dispersibility and stability of photosensitizers, thereby improving their pharmacokinetic  
186 characteristics [28]. In recent years, poly(ethylene glycol) (PEG) as a non-toxic, nonimmunogenic,  
187 nonantigenic and water soluble polymer, has been frequently employed to construct organic core-  
188 shell structured nanoparticles for PDT of cancer [63, 101]. For example, Kim et al. conjugated a  
189 pheophorbide a (PhA) photosensitizer with methoxy PEG (mPEG) through disulfide bond to fabricate  
190 the core-shell structured mPEG-(ss-PhA) nanoparticles for PDT [102]. The disulfide bond was  
191 broken in the intracellular reductive environment, thereby promoting the rapid release of PhA  
192 photosensitizer. An et al. constructed an organic core-shell structured nanoparticle by conjugating  
193 Ce6 photosensitizer with luminol chemiluminescence substrate and PEG for H<sub>2</sub>O<sub>2</sub>-triggered in situ  
194 PDT [103]. At a pathologically relevant H<sub>2</sub>O<sub>2</sub> concentration, the Ce6 photosensitizer was activated  
195 through chemiluminescence resonance energy transfer to generate <sup>1</sup>O<sub>2</sub> for in situ PDT of tumors and  
196 repressing lung metastasis. Besides, lipids have also been used to fabricate organic core-shell  
197 structured nanoparticles for encapsulating photosensitizers in PDT of cancer. Cheng et al.  
198 encapsulated IR780 photosensitizer and perfluorocarbon by lipids to create organic core-shell  
199 structured nanoparticles for PDT [104]. In the study of Chang et al., porphyrin-lipid shell was utilized  
200 to stabilize the water/oil interface to develop organic core-shell structured nanoparticles for

201 PDT/chemotherapy [64].

### 202 2.3. Hybrid core-shell structured nanoparticles

203 There are two typical forms of hybrid core-shell structured nanoparticles: inorganic core-organic  
204 shell and organic core-inorganic shell. Normally, the inorganic-organic core-shell structured  
205 nanoparticles applied in PDT are made of metals [67], metal oxides [105] and UCNPs [106, 107]  
206 cores and polymers [67, 106, 107] and organic carbonaceous materials [105, 108] shells. One of the  
207 advantages of coating the organic shell on the inorganic core is that it can improve the stability and  
208 biocompatibility of the inorganic core. Meanwhile, the organic shell has abundant functional groups,  
209 which enables further photosensitizers loading and surface modification [109, 111]. For example, Tan  
210 et al. coated a polyaniline (PANI) shell on a Ag core and then loaded an indocyanine green (ICG)  
211 photosensitizer to prepare the core-shell structured ICG-Ag@PANI nanoparticles for PDT/PTT [68].  
212 The cell viability could still be maintained at about 70% in the dark when the concentration of  
213 Ag@PANI nanoparticles was as high as  $400 \mu\text{g mL}^{-1}$ , which indicated the good biocompatibility of  
214 Ag@PANI nanoparticles. Liu et al. fabricated a novel core-shell structured gold  
215 nanoprism@mesoporous organosilica nanoparticle to load zinc(II) phthalocyanine (ZnPc)  
216 photosensitizer for PDT/PTT [112]. Owing to the  $\pi$ - $\pi$  stacking and hydrophobic interactions induced  
217 by the mesoporous organosilica shell, the loading of the ZnPc photosensitizer could be as high as 11.8  
218 wt%. In the study of Feng et al.,  $\text{Fe}_3\text{O}_4$  was employed as the core to in situ grow the covalent-organic  
219 frameworks (COFs) shell for PDT/PTT [69]. Due to the excellent biocompatibility of COFs, the cell  
220 viability of  $\text{Fe}_3\text{O}_4$ @COFs nanoparticles in the dark remained about 80% at a high concentration of  
221  $800 \mu\text{g mL}^{-1}$ . Ultimately, the  $\text{Fe}_3\text{O}_4$ @COFs nanoparticles presented satisfactory capacity to kill cancer  
222 cells and inhibit tumor growth through the synergistic effect of PDT/PTT. The structure of organic-  
223 inorganic core-shell structured nanoparticles is just the reverse of the above type. Coating the

224 inorganic shell on the organic core is beneficial to enhance the whole strength and wear resistance of  
225 the nanoparticles [53]. In the study of Zhu et al., a  $\text{MnO}_2$  shell was coated on a semiconducting hybrid  
226 nanoparticles (SPN) core through an in situ growth strategy [72]. Compared with the uncoated SPN,  
227 the  $\text{SPN@MnO}_2$  nanoparticles showed better PDT efficacy as it could produce more  $^1\text{O}_2$  in the  
228 hypoxic and acidic tumor microenvironment.

229 Moreover, metal-organic frameworks (MOFs), as an emerging hybrid functional materials  
230 assembled from inorganic metal nodes and organic linkers, have been extensively utilized to fabricate  
231 core-shell structured nanoparticles for PDT of cancer owing to their large surface area, tunable pore  
232 structure, intrinsic biodegradability and excellent biocompatibility [113-115]. Notably, the porous  
233 structure of MOFs can not only prevent the aggregation of photosensitizers to reduce their self-  
234 quenching, but also promote the diffusion of ROS. In the study of Ren et al., a pH-responsive  
235 nanoparticle was prepared for PDT/chemotherapy by encapsulating a DOX drug and a protoporphyrin  
236 IX (PpIX) photosensitizer in a zeolitic imidazolate framework-67 (ZIF-67) core and a zeolitic  
237 imidazolate framework-8 (ZIF-8) shell, respectively [74]. The ZIF-8 shell degraded in weak acidic  
238 tumor microenvironment, triggering the prior release of PpIX. Then the ZIF-67 core rapidly catalyzed  
239  $\text{H}_2\text{O}_2$  to produce  $\text{O}_2$ , which was utilized by PpIX to generate ROS under laser irradiation for enhanced  
240 PDT. Meanwhile, the decomposition of ZIF-67 core induced the release of DOX for chemotherapy.  
241 Moreover, in the study of Liu et al., an  $\text{O}_2$  self-evolving nanoparticle was fabricated through coating  
242 a Material of Institute Lavoisier- $\text{NH}_2$  (MIL) shell on a  $\text{CeO}_x$  core for PDT [116]. Benefiting from the  
243 encapsulation and protection of the MIL shell, the  $\text{CeO}_x\text{@MIL}$  nanoparticles presented more stable  
244 activity for generating ROS in complex tumor microenvironment.

### 245 3. Core-shell structured nanoparticles for PDT of cancer

246 Recently, core-shell structured nanoparticles have been extensively applied in PDT of cancer.

247 They play four main functions in this treatment: photosensitizer delivery vehicles, energy transducers,  
248 photosensitizers and hypoxic tumor microenvironment modulators. Notably, most core-shell  
249 structured nanoparticles can simultaneously perform multiple functions.

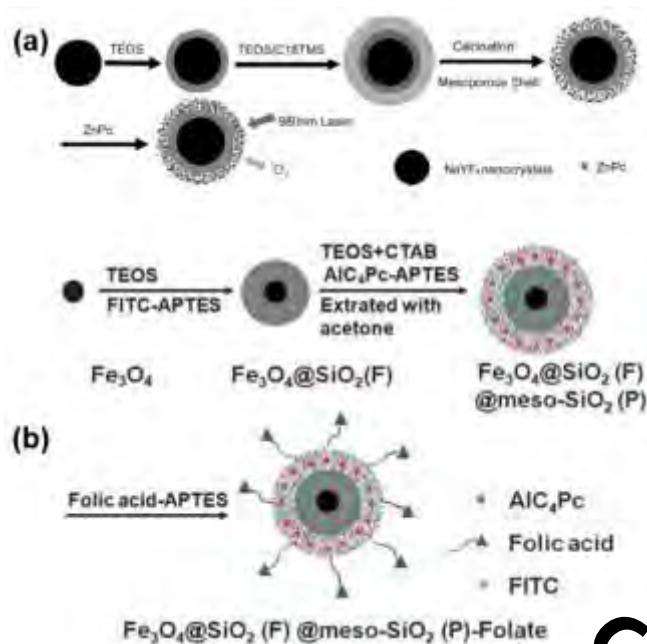
### 250 3.1. Photosensitizer delivery vehicles

251 In general, most photosensitizers are organic small molecules, which are easy to self-aggregate  
252 in aqueous phase, leading to a decrease in PDT efficacy [22, 117]. Accordingly, appropriate delivery  
253 vehicles are needed to enhance their stability and targeting ability in PDT of cancer. The development  
254 of nanotechnology enables nanoparticles with a core-shell structure to meet these demands,  
255 improving the selectivity of photosensitizers to cancer cells [118-120]. In this process, the  
256 photosensitizers are first encapsulated into the core-shell structured nanoparticles through physical  
257 adsorption and chemical bonding. After the core-shell structured nanoparticles reach the targeted  
258 cancer cells and are irradiated by the light with a specific wavelength, the embedded photosensitizers  
259 will be excited and produce a large amount of toxic ROS to kill the cancer cells [121, 122].

260 In a core-shell structured nanoparticle, both the core and shell can be used to load  
261 photosensitizers for PDT of cancer. Particularly, mesoporous nanostructures with large pore volume  
262 and high surface area are extremely beneficial to the loading of photosensitizers [123, 124]. For  
263 example, Qian et al. fabricated  $\text{NaYF}_4:\text{Yb,Er}@ \text{SiO}_2@m\text{SiO}_2$  nanoparticles for PDT of MB49 bladder  
264 cancer cells (Fig. 1a) [125]. Incorporating the ZnPc photosensitizer into the  $m\text{SiO}_2$  shell prevented it  
265 from being degraded in the complex biological environment and accelerated the release of ROS. Zeng  
266 et al. constructed  $\text{MnO}_2@\text{polydopamine (PDA)-folic acid (FA)}$  nanoparticles in which the Ce6  
267 photosensitizer was loaded into the hollow mesoporous  $\text{MnO}_2$  core for PDT of breast cancer [126].  
268 The PDA shell avoided the premature release of Ce6 in blood circulation, while after reaching the  
269 acidic tumor site, the Ce6 was released because of the destruction of PDA shell. Meanwhile,

270 photosensitizers can self-assemble with other organic molecules to form core-shell structured self-  
271 delivery nanoparticles for PDT of cancer [127, 128]. In the study of Liu et al., about 13.89% of the  
272 Ce6 photosensitizer was loaded into a core-shell structured nanoparticle, which was composed of a  
273 core formed by self-assembly of Ce6 and rapamycin as well as a MOFs shell loaded with catalase  
274 [129].

275 In addition, the targeting ability of core-shell structured nanoparticles is critical to deliver  
276 photosensitizers to tumor sites. There are two routes utilized for the controlled delivery: active and  
277 passive delivery. In the case of active targeting, the customized tumor-targeting ligands are introduced  
278 on the core-shell structured nanoparticles for recognizing target cell receptors to deliver  
279 photosensitizers [130, 131]. For example, folic acid (FA) exhibits a high affinity with folate receptor  
280 protein, which usually overexpresses on the surface of various cancer cells [132]. Wang et al. achieved  
281 superior cancer cell targeting ability in PDT by decorating the  $\text{Fe}_3\text{O}_4@\text{SiO}_2@\text{mSiO}_2$  nanoparticles  
282 with FA (Fig. 1b) [58]. On the other hand, for passive targeting, the photosensitizers loaded core-shell  
283 structured nanoparticles will selectively accumulate in targeted cancer cells because of  
284 physicochemical or pharmacological factors [133]. Normally, in the case of passive targeting, most  
285 core-shell structured nanoparticles deliver photosensitizers based on the enhanced permeability and  
286 retention (EPR) effect [134, 135]. In the study of Meng et al., owing to the high angiogenesis of triple-  
287 negative breast cancer, the CDTNs selectively accumulated in it via EPR effect, improving the PDT  
288 efficacy [136]. Wang et al. reported that the accumulation of FA modified core-shell structured  
289 poly(lactic-co-glycolic acid) (PLGA) nanoparticles in tumor was attributed to the synergistic effect  
290 of active targeting and EPR effect [137]. Overall, with the help of the delivery of core-shell structured  
291 nanoparticles, the photosensitizers can successfully reach the targeted cancer cells, thereby reducing  
292 damage to the surrounding healthy cells and enhancing PDT efficacy.



**Fig .1.** (a) Schematic illustration of the synthesis of NaYF<sub>4</sub>:Yb,Er@SiO<sub>2</sub>@mSiO<sub>2</sub> nanoparticles. Reproduced with permission. [125] Copyright 2009, Wiley-VCH. (b) Schematic illustration of the synthesis of

Fe<sub>3</sub>O<sub>4</sub>@SiO<sub>2</sub>@mSiO<sub>2</sub>-FA nanoparticles. Reproduced with permission. [58] Copyright 2011, Royal Society of Chemistry.

### 3.2. Energy transducers

Core-shell structured nanoparticles can not only serve as delivery vehicles for photosensitizers, but also act as energy transducers to excite photosensitizers. Since most traditional photosensitizers are excited by UV-VIS light that possesses limited tissue penetration depth, the clinical application of PDT is greatly hindered [117, 138]. Combining these photosensitizers with core-shell structured nanoparticles with energy conversion properties is an effective way to solve this problem. In this process, the core-shell structured nanoparticles can convert the light with strong tissue penetration (e.g., NIR light and X-ray) to UV-VIS light for exciting the photosensitizers [24, 139].

Among them, UCNPs are the most concerned, which can convert NIR light to UV-VIS light via an anti-Stokes emission process to excite photosensitizers for PDT of cancer [140-142]. Park et al. incorporated a Ce6 photosensitizer into the NaYF<sub>4</sub>:Yb,Er@NaGdF<sub>4</sub> nanoparticles for PDT of



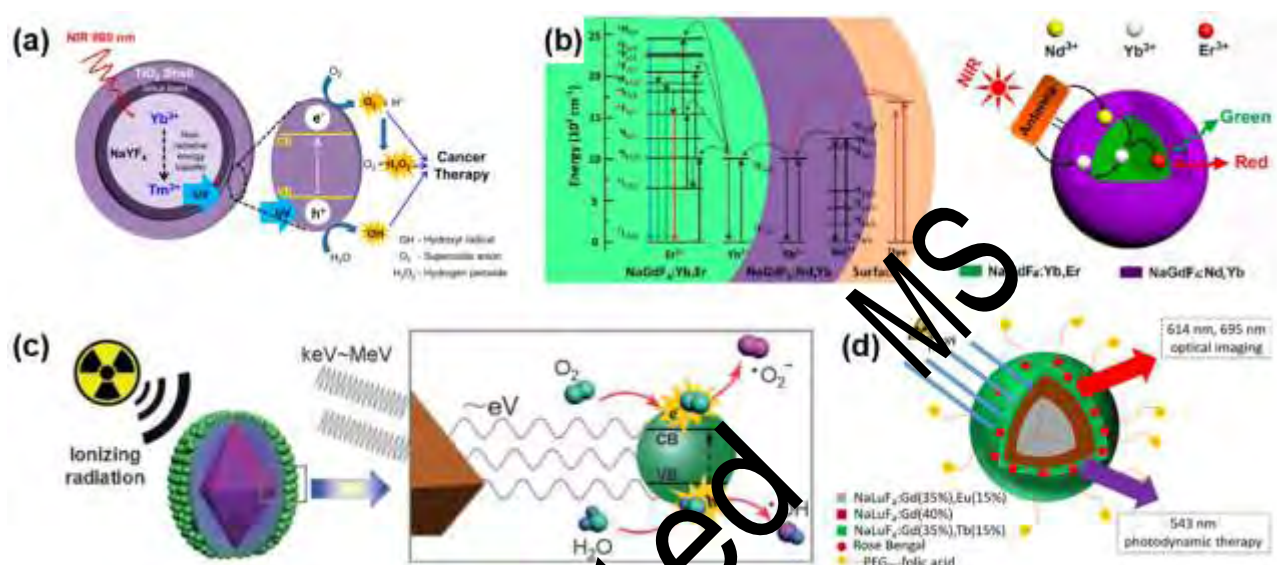
309 U87MG tumor under NIR light irradiation [143]. When irradiated by a 980 nm NIR laser, the  
 310 NaYF<sub>4</sub>:Yb,Er@NaGdF<sub>4</sub> nanoparticles emitted red light, which was exploited to excite the Ce6 to  
 311 produce cytotoxic <sup>1</sup>O<sub>2</sub>, resulting in the necrosis of U87MG tumor. In the study of Lucky et al., the  
 312 NaYF<sub>4</sub>:Yb,Tm@TiO<sub>2</sub>-PEG nanoparticles exhibited admirable activity for PDT of human oral  
 313 squamous cell carcinoma (OSCC) cells both *in vitro* and *in vivo* under 980 nm NIR laser irradiation  
 314 [95]. In this system, electrons in the valence band (VB) of TiO<sub>2</sub> shell was excited to the conduction  
 315 band (CB) because the NaYF<sub>4</sub>:Yb,Tm core could convert NIR light to ultraviolet light (Fig. 2a).  
 316 Consequently, the generation of charge carriers promoted the formation of ROS for killing the OSCC  
 317 cells. Nevertheless, Yb<sup>3+</sup>-sensitized UCNPs usually need to be excited by the 980 nm NIR light,  
 318 which overlaps with the absorption of water molecules, leading to low tissue penetration depth and  
 319 overheating of tissues [144, 145]. Nd<sup>3+</sup> doping can effectively solve this issue because it can tune the  
 320 excitation wavelength of Yb<sup>3+</sup>-sensitized UCNPs from 980 nm to around 800 nm where the tissue  
 321 transparency is maximal and the heating effect is minimal [146-148]. For example, Xu et al. fabricated  
 322 the dye-sensitized NaGdF<sub>4</sub>:Yb,Er@NaGdF<sub>4</sub>:Nd,Yb nanoparticles for dual-photosensitizer PDT of  
 323 cancer upon 808 nm NIR laser excitation [88]. As displayed in Fig. 2b, it converted 808 nm photons  
 324 to green and red light, thereby exciting the MC540 and Ce6 photosensitizers respectively to generate  
 325 ROS for cancer therapy. As expected, the *in vitro* and *in vivo* tests demonstrated that the dye-  
 326 sensitized NaGdF<sub>4</sub>:Yb,Er@NaGdF<sub>4</sub>:Nd,Yb nanoparticles possessed high efficacy for PDT of HeLa  
 327 cancer cells and U14 tumors, respectively.

328 In addition to UCNPs, scintillator nanoparticles (SCNPs) are another promising energy  
 329 transducers in PDT of cancer [149, 150]. They present a high X-ray shielding capability and can  
 330 convert X-ray to UV-VIS fluorescence [151, 152]. For example, Zhang et al. prepared  
 331 LiYF<sub>4</sub>:Ce@SiO<sub>2</sub>@ZnO nanoparticles for PDT of HeLa cancer cells under X-ray radiation [153]. As

332 depicted in Fig. 2c, the  $\text{LiYF}_4\text{:Ce}$  core was excited by X-ray radiation and emitted ultraviolet  
333 fluorescence, which was utilized to induce the formation of photogenerated charge carriers in the ZnO  
334 shell. Subsequently, the photogenerated electrons and holes reacted with  $\text{O}_2$  and  $\text{H}_2\text{O}$  to produce  $\cdot\text{O}_2^-$   
335 and  $\cdot\text{OH}$  respectively, thereby enhancing the antitumor therapeutic efficacy of PDT. Meanwhile,  
336 through the fluorescence resonance energy transfer (FRET) between SCNPs and photosensitizers, the  
337 SCNPs can efficiently realize deep PDT [154]. In the study of Hsu et al.,  
338  $\text{NaLuF}_4\text{:Gd,Eu@NaLuF}_4\text{:Gd@NaLuF}_4\text{:Gd,Tb}$  nanoparticles were designed for deep tissue PDT  
339 under X-ray radiation (Fig. 2d) [155]. Upon X-ray excitation, it emitted 543 nm green light (from  
340  $\text{Tb}^{3+}$ ), which overlapped with the main absorption peak of the loaded rose bengal (RB) photosensitizer  
341 (549 nm), allowing efficient FRET from the  $\text{NaLuF}_4\text{:Gd,Eu@NaLuF}_4\text{:Gd@NaLuF}_4\text{:Gd,Tb}$  donor to  
342 the RB acceptor. By virtue of the integral FRET system, a large amount of  $^1\text{O}_2$  was produced to kill  
343 the MDA-MB-231 and MCF-7 cancer cells.

344 Notably, as variants of traditional core-shell structured nanoparticles, yolk-shell-like  
345 (core@void@shell) and hollow-like (void@shell) nanoparticles are considered to be beneficial for  
346 energy transfer in PDT due to the presence of internal cavity structures that facilitates light scattering  
347 [156-158]. For example, Wang et al. constructed  $\text{UCNPs@Zn}_x\text{Cd}_{1-x}\text{S}$  yolk-shell-like nanoparticles  
348 for PDT of HeLa cancer cells under 980 nm NIR laser irradiation [159]. The steady and dynamic  
349 fluorescence spectra demonstrated that the  $\text{UCNPs@Zn}_x\text{Cd}_{1-x}\text{S}$  yolk-shell-like nanoparticle was an  
350 efficient energy transducer for NIR light because it significantly enhanced the energy transfer  
351 efficiency. In the study of Chang et al.,  $\text{Au@CuS}$  yolk-shell-like nanoparticles were developed for  
352 PDT/PTT/chemotherapy [160]. The yolk-shell structure could enhance local electromagnetic field to  
353 induce a resonance energy transfer from Au core to CuS shell, improving both photodynamic and  
354 photothermal performance. As a result, under 980 nm NIR laser irradiation, it displayed excellent

355 antitumor efficacy for *in vitro* 4T1 cancer cells and *in vivo* 4T1 tumor-bearing mice. Moreover, to  
 356 obtain high energy transfer efficiency, Kamkaew et al. utilized the hollow mSiO<sub>2</sub> nanoparticles as a  
 357 carrier to simultaneously encapsulate <sup>89</sup>Zr isotope and Ce6 photosensitizer [161]. In this system, <sup>89</sup>Zr  
 358 isotope could serve as a Cerenkov radiation source to excite Ce6 photosensitizer to produce ROS for  
 359 PDT of cancer.



360  
 361 **Fig. 2.** (a) Mechanism of the NaYF<sub>4</sub>:Yb,Tm@TiO<sub>2</sub>-PEG nanoparticles in PDT of cancer under 980 nm NIR laser  
 362 irradiation. Reproduced with permission. [151] Copyright 2015, American Chemical Society. (b) Proposed energy  
 363 level diagram and energy-transfer mechanism in the dye-sensitized NaGdF<sub>4</sub>:Yb,Er@NaGdF<sub>4</sub>:Nd,Yb nanoparticles  
 364 under 808 nm NIR laser irradiation. Reproduced with permission. [88] Copyright 2017, American Chemical  
 365 Society. (c) Mechanism of the LiYF<sub>4</sub>:Ce@SiO<sub>2</sub>@ZnO nanoparticles in PDT of cancer under X-ray radiation.  
 366 Reproduced with permission. [153] Copyright 2015, Wiley-VCH. (d) Energy-transfer mechanism in the  
 367 NaLuF<sub>4</sub>:Gd,Eu@NaLuF<sub>4</sub>:Gd@NaLuF<sub>4</sub>:Gd,Tb nanoparticles under X-ray radiation. Reproduced with permission.  
 368 [155] Copyright 2018, American Chemical Society.

### 369 3.3. Photosensitizers

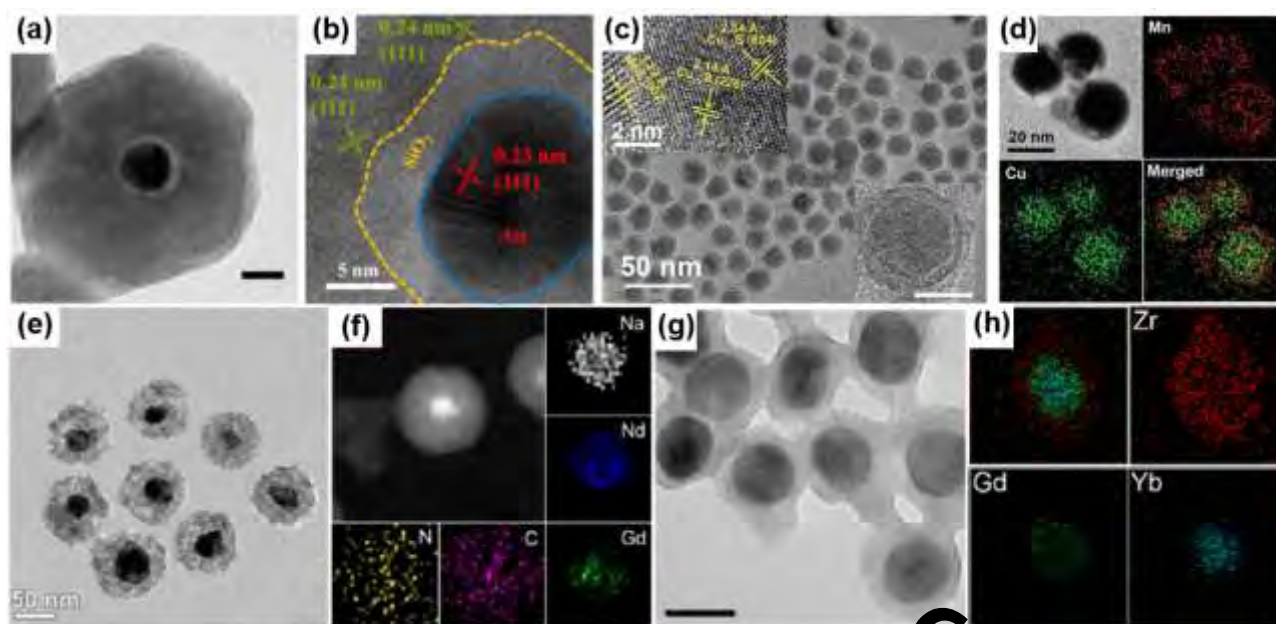
370 Owing to the unique light absorption characteristics, some core-shell structured nanoparticles  
 371 have the capability to produce ROS upon light excitation, which allows them to serve as

372 photosensitizers in PDT by themselves. Metal oxide and sulfide semiconductors have attracted much  
 373 attention as photosensitizers in PDT of cancer because of their efficient photoactivity [162-164].  
 374 Under light irradiation, they are induced to generate electron-hole pairs, which can react with O<sub>2</sub> and  
 375 H<sub>2</sub>O to produce various ROS for killing the cancer cells. For example, Liu et al. constructed  
 376 Au@SiO<sub>2</sub>@Cu<sub>2</sub>O nanoparticles (Fig. 3a and b) and loaded them into perfluorohexane droplets with  
 377 liposome coating for PDT of cancer under 670 nm laser irradiation [165]. The process of plasmon-  
 378 induced resonance energy transfer from Au core to Cu<sub>2</sub>O shell facilitated the generation of charges in  
 379 Cu<sub>2</sub>O shell, resulting in a significant increase in the quantum yield of <sup>1</sup>O<sub>2</sub>. Hence this nanocomposite  
 380 exhibited outstanding anticancer efficacy for *in vitro* MCF-7 cancer cells and *in vivo* MCF-7 tumor  
 381 xenotransplanted BALB/c nude mice. Wang et al. integrated oxygen vacancy-enriched core-shell  
 382 structured crystalline@amorphous black TiO<sub>2</sub> into a chitosan matrix for PDT of cancer [166]. Under  
 383 808 nm NIR laser irradiation, the thermogel showed considerable activity in killing B16F10 cells *in*  
 384 *vitro* and inhibiting B16F10 tumors growth *in vivo*. Moreover, Huang et al. prepared Cu<sub>2-x</sub>S@MnS  
 385 nanoparticles (Fig. 3c and d) for PDT of HeLa cancer cells [61]. In this structure, the Cu<sub>2-x</sub>S core  
 386 acted as a photosensitizer to generate ROS for PDT upon 808 nm NIR laser excitation.

387 Apart from metal oxide and sulfide semiconductors, graphitic carbon nitride (g-C<sub>3</sub>N<sub>4</sub>) also has  
 388 been employed as a photosensitizer for PDT of cancer, which is ascribed to its low cytotoxicity,  
 389 excellent biocompatibility, good photostability and low cost [167-171]. As exhibited in Fig. 3e and f,  
 390 Feng et al. created UCNPs@g-C<sub>3</sub>N<sub>4</sub>-PEG nanoparticles for PDT of HeLa cells *in vitro* and U14  
 391 tumors *in vivo* under 808 nm NIR laser irradiation [70]. The UCNPs core converted absorbed NIR  
 392 light to UV-VIS light, which could excite electrons in the VB of g-C<sub>3</sub>N<sub>4</sub> shell to the CB and thus  
 393 induce the formation of photogenerated electron-hole pairs. These photogenerated electrons and holes  
 394 then reacted with O<sub>2</sub> and H<sub>2</sub>O respectively to produce ·O<sub>2</sub><sup>-</sup> and ·OH, resulting in the death of HeLa

395 cells and inhibition of U14 tumors growth. In the study of Zhang et al., the nitrogen-doped graphene  
396 quantum dot (N-GQD)@hollow mSiO<sub>2</sub>@g-C<sub>3</sub>N<sub>4</sub>-amphipathic polymer (R-NCNP) nanoparticles  
397 presented superior anticancer effects both *in vitro* and *in vivo* [108]. In this nanocomposite, the g-  
398 C<sub>3</sub>N<sub>4</sub> photosensitizer was excited by a 630 nm laser to produce ROS for PDT of cancer.

399 More recently, porphyrinic MOFs have shown great potential as photosensitizers for PDT of  
400 cancer [172-174]. Since porphyrinic MOFs are directly self-assembled by the coordination  
401 interactions between porphyrin photosensitizers and metal ions/clusters, the porphyrin-derived  
402 molecules are uniformly dispersed in the whole porphyrinic MOFs framework, which maximizes the  
403 light harvesting ability. Meanwhile, the abundant pore structures of MOFs accelerate the diffusion of  
404 ROS, thereby enhancing the PDT efficacy. For example, Zeng et al. reported that AuNR@porphyrinic  
405 MOFs nanoparticles exhibited excellent PDT efficacy under NIR laser excitation for killing the cancer  
406 cells *in vitro* and inhibiting the tumor growth and metastasis *in vivo* [73]. In the study of Shao et al.,  
407 UCNPs@porphyrinic MOFs (UCSs) nanoparticles (Fig. 3g and h) were constructed for PDT of  
408 hypoxic tumors [42]. Benefiting from the efficient energy transfer from UCNPs core to porphyrinic  
409 MOFs shell, the UCNPs@porphyrinic MOFs nanoparticles presented rapid generation of <sup>1</sup>O<sub>2</sub> under  
410 980 nm NIR laser irradiation, resulting in the enhanced PDT efficacy.



**Fig. 3.** (a) TEM image and (b) interfacial HRTEM image of the Au@SiO<sub>2</sub>@Cu<sub>2</sub>O nanoparticles. Reproduced with permission. [165] Copyright 2018, American Chemical Society. (c) TEM image and (d) EDS elemental mapping images of the Cu<sub>2-x</sub>S@MnS nanoparticles. Reproduced with permission. [61] Copyright 2019, Wiley-VCH. (e) TEM image of the UCNPs@g-C<sub>3</sub>N<sub>4</sub> nanoparticles, and (f) STEM image and EDS elemental mapping images of the UCNPs@g-C<sub>3</sub>N<sub>4</sub>-PEG nanoparticles. Reproduced with permission. [70] Copyright 2016, American Chemical Society. (g) TEM image and (h) EDS elemental mapping images of the UCNPs@porphyrinic MOFs nanoparticles. Reproduced with permission. [42] Copyright 2020, American Chemical Society.

### 3.4. Hypoxic tumor microenvironment modulators

Hypoxia is a prominent feature of tumor microenvironment, which originates from the uncontrolled cancer cells growth and abnormal angiogenesis [175, 176]. Moreover, the process of PDT also consumes O<sub>2</sub> to generate ROS, thereby exacerbating the tumor hypoxia [177, 178]. Hypoxic tumor microenvironment not only accelerates the cancer metastasis but also impairs the therapeutic efficacy of PDT [179, 180]. Recently, various core-shell structured nanoparticles have been developed to modulate hypoxic tumor microenvironment for attenuating the tumor hypoxia in PDT [181-184]. Normally, there are two main mechanisms in the process of utilizing core-shell structured



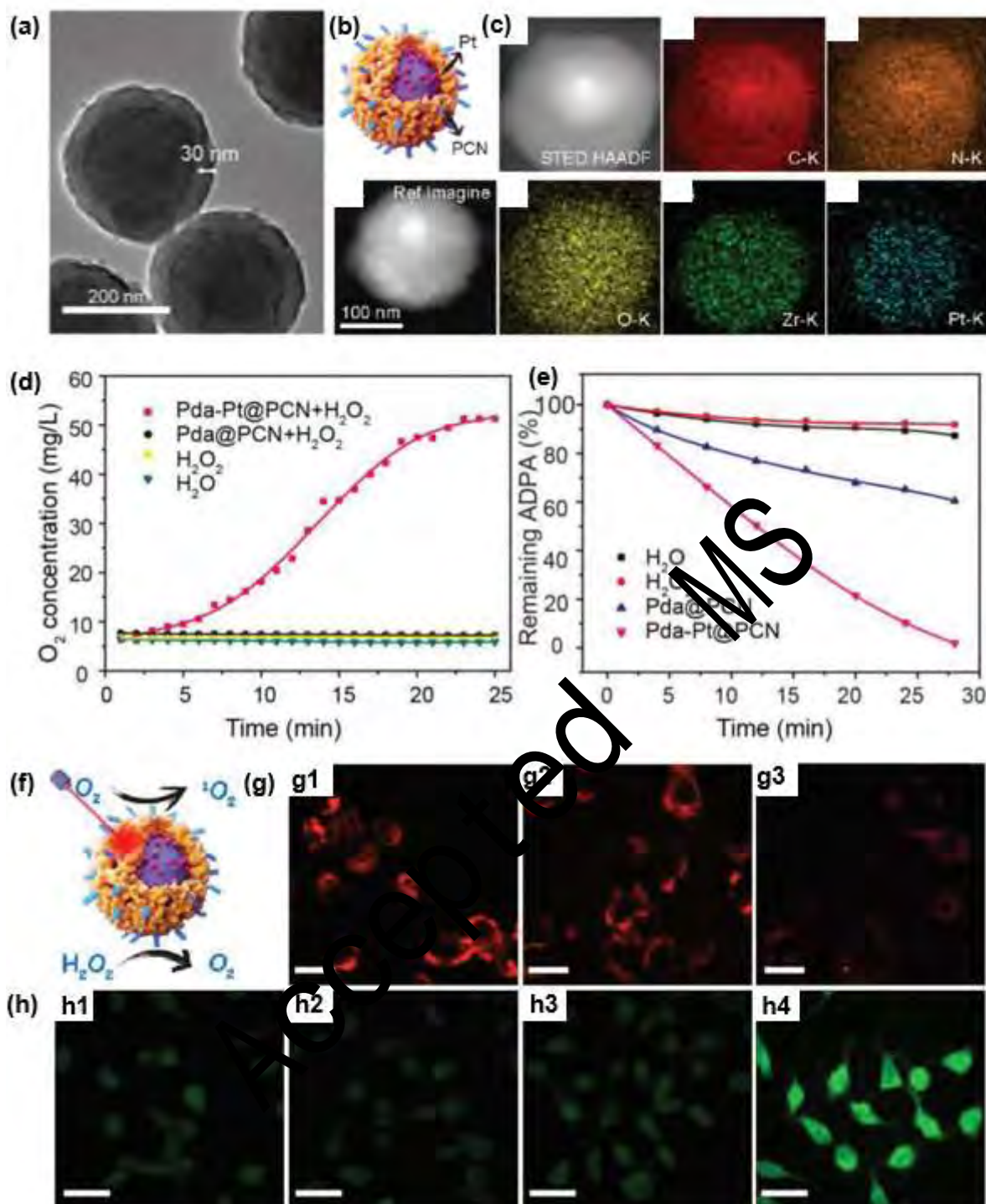
nanoparticles to overcome this obstacle.

One mechanism is the direct transport of O<sub>2</sub> to hypoxic tumor areas through the core-shell structured nanoparticles to effectively oxygenate the tumor. Perfluorocarbon is an efficient O<sub>2</sub> carrier due to its high affinity toward O<sub>2</sub> molecules [65, 185]. In the study of Cheng et al., the IR780 photosensitizer was loaded into an oxygen self-enriched nanoparticle, which was composed of perfluorocarbon droplet core and lipid shell [104]. In this nanoparticle, the IR780 photosensitizer was evenly dispersed inside the lipid shell. When irradiated by a 808 nm NIR laser, the IR780 transferred energy to the oxygen enriched-perfluorocarbon droplet core for cytotoxic <sup>1</sup>O<sub>2</sub> production, leading to an enhanced tumor inhibition. However, perfluorocarbon-based O<sub>2</sub> carriers present a limited ability to transport O<sub>2</sub> to the tumor site. On account of the large pore volume and high surface area, MOFs have been regarded as promising candidates for O<sub>2</sub> storage and transport [186, 187]. Xie et al. constructed a multifunctional nanoplatform by covalently conjugating DOX and NH<sub>2</sub>-poly(ethylene glycol) modified folic acid on the surface of core-shell structured UCNPs@mSiO<sub>2</sub>-RB@ZIF-90 nanoparticles for highly efficient cancer therapy under 808 nm NIR laser irradiation [188]. The outermost ZIF-90 shell was an O<sub>2</sub> reservoir, which decomposed under acidic conditions, enabling rapid release of O<sub>2</sub> at hypoxic tumor microenvironment. After the addition of UCNPs@mSiO<sub>2</sub>-RB@ZIF-90 nanoparticles, the O<sub>2</sub> concentration in deoxygenated phosphate-buffered saline (PBS) solution at low pH value was increased.

Another mechanism is using core-shell structured nanoparticles with catalase-like properties to catalyze endogenous H<sub>2</sub>O<sub>2</sub> for in situ O<sub>2</sub> production. Mn-based materials (e.g., MnO<sub>2</sub> [189], MnS [61] and Mn-Cdots [190], etc.) are the most commonly used catalase-like nanoenzymes to alleviate tumor hypoxia by virtue of their superior activity. For example, Zhu et al. developed a core-shell structured nanoparticle composed of a MnO<sub>2</sub> shell and a SPN core for enhanced PDT of 4T1 cancer cells both

450 *in vitro* and *in vivo* [72]. Under hypoxic and acidic tumor microenvironment, the  $\text{MnO}_2$  shell  
451 decomposed  $\text{H}_2\text{O}_2$  to  $\text{O}_2$ . Subsequently, the  $\text{O}_2$  was activated by the SPN core under 808 nm NIR laser  
452 irradiation to form  $^1\text{O}_2$  for cancer therapy. Compared with the uncoated SPN, the  $\text{MnO}_2$  coated SPN  
453 (SPN-M1) produced 2.68-fold more  $^1\text{O}_2$  at hypoxic and acidic conditions under NIR laser irradiation.  
454 Moreover, Huang et al. reported that the MnS shell in  $\text{Cu}_{2-x}\text{S}@\text{MnS}$  nanoparticles acted as a  $\text{H}_2\text{O}_2$   
455 responder to mediate  $\text{O}_2$  production for efficiently relieving tumor hypoxia [61]. Nevertheless, Mn-  
456 based materials are only suitable for acidic tumor microenvironment since their catalytic activity is  
457 greatly affected by the pH. As an emerging catalase-like nanoenzyme, noble metals have drawn much  
458 attention due to their admirable stability and pH-independent activity [191–193]. For example, Wang  
459 et al. designed Pt-based core-shell structured nanoparticles to promote the decomposition of  
460 endogenous  $\text{H}_2\text{O}_2$  for enhanced PDT efficacy (Fig. 4a–c) [194]. In this nanoparticle, the Pt interlayer  
461 first decomposed the endogenous  $\text{H}_2\text{O}_2$  to  $\text{O}_2$ , which was then converted to cytotoxic  $^1\text{O}_2$  by the  
462 zirconium-porphyrin (PCN) shell when exposed to light irradiation (Fig. 4f). As shown in Fig. 4d and  
463 e, the  $\text{O}_2$  generation and  $^1\text{O}_2$  production efficiencies over the polydopamine (Pda)-Pt@PCN  
464 nanoparticles were significantly enhanced. Meanwhile, cellular level tests further verified the  $\text{O}_2$ -  
465 generating capability and the improved  $^1\text{O}_2$ -producing capability of the Pda-Pt@PCN nanoparticles  
466 (Fig. 4g and h). In another study, a porous  $\text{Au}@\text{Rh}$  core-shell structured nanoparticle was developed  
467 to alleviate tumor hypoxia for improved PDT [78]. As expected, it showed excellent catalase-like  
468 activity to effectively decompose  $\text{H}_2\text{O}_2$  to  $\text{O}_2$  in tumors.

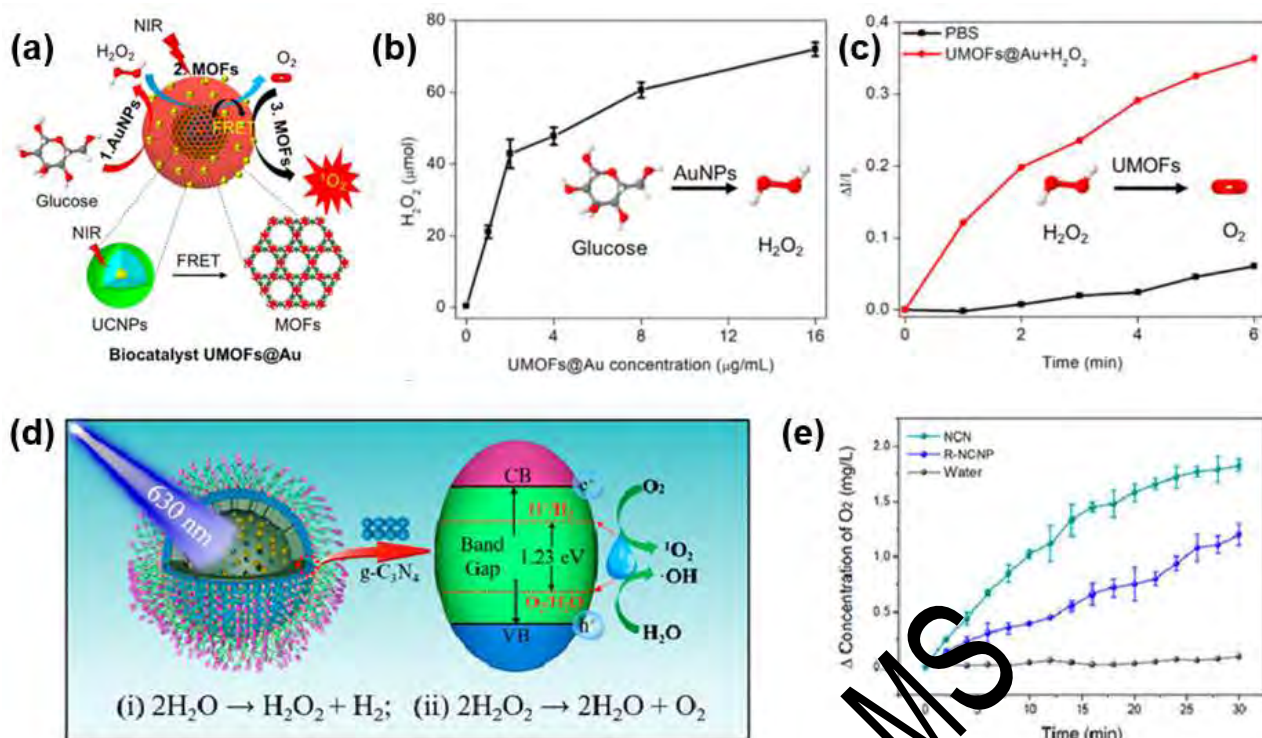




**Fig. 4.** (a) TEM image, (b) schematic illustration, (c) STEM-HAADF image and the corresponding element mapping images of the Pda-Pt@PCN nanoparticles. (d) O<sub>2</sub> generation and (e) <sup>1</sup>O<sub>2</sub> production efficiencies over different samples. (f) Schematic illustration of O<sub>2</sub> generation and <sup>1</sup>O<sub>2</sub> production over the Pda-Pt@PCN nanoparticles upon light excitation. (g) CLSM images of intracellular hypoxia levels in CT26 cells under 5% O<sub>2</sub> treated with (g1) blank, (g2) Pda@PCN-FA and (g3) Pda-Pt@PCN-FA. The scale bar is 36 μm. (h) CLSM images

475 of ROS production in CT26 cells treated with (h1) 2',7'-dichlorofluorescein diacetate in dark, (h2) 2',7'-  
476 dichlorofluorescein diacetate under 660 nm LED irradiation, (h3) Pda-Pt@PCN-FA in dark and (h4) Pda-  
477 Pt@PCN-FA under 660 nm LED irradiation. The irradiation time was 2 min. The scale bar is 36  $\mu$ m. Reproduced  
478 with permission. [194] Copyright 2018, Wiley-VCH.

479 Unfortunately, the amount of O<sub>2</sub> generated by catalase-like core-shell structured nanoparticles is  
480 highly dependent on the decomposition of endogenous H<sub>2</sub>O<sub>2</sub>, while the low level of endogenous H<sub>2</sub>O<sub>2</sub>  
481 in tumor cells is not enough to produce a considerable amount of O<sub>2</sub> to alleviate tumor hypoxia [195].  
482 To solve this problem, He et al. designed a UCNPs@MOFs(UMOFs)@Au cascade biocatalyst to  
483 continuously produce O<sub>2</sub> for PDT (Fig. 5a) [196]. Firstly, the ultrasmall Au nanoparticles first  
484 converted glucose to H<sub>2</sub>O<sub>2</sub> in tumor microenvironment, leading to the increase of H<sub>2</sub>O<sub>2</sub> concentration  
485 (Fig. 5b). Subsequently, the H<sub>2</sub>O<sub>2</sub> was decomposed by the iron porphyrin MOFs shell through the  
486 catalase-like reaction to generate O<sub>2</sub> (Fig. 5c). Finally, the UCNPs core converted NIR light to visible  
487 light, thereby exciting the iron porphyrin MOFs shell to produce cytotoxic <sup>1</sup>O<sub>2</sub> for cancer therapy. In  
488 addition, given the abundance and availability of endogenous H<sub>2</sub>O in tumor microenvironment,  
489 Zhang et al. designed an intelligent nanoregulator (R-NCNP) to execute laser-excited water splitting  
490 for enhanced PDT (Fig. 5d) [198]. As shown in Fig. 5e, the g-C<sub>3</sub>N<sub>4</sub> in R-NCNP split H<sub>2</sub>O to O<sub>2</sub> under  
491 630 nm laser irradiation, which was then converted to <sup>1</sup>O<sub>2</sub> by the photosensitizers in R-NCNP, thereby  
492 efficiently attenuating tumor hypoxia and enhancing PDT efficacy.



**Fig. 5.** (a) Schematic illustration of the UMOFs@Au cascade biocatalyst for PDT. (b) Concentration-dependent H<sub>2</sub>O<sub>2</sub> generation and (c) time-dependent O<sub>2</sub> generation from the UMOFs@Au cascade biocatalyst. Reproduced with permission. [196] Copyright 2020, American Chemical Society. (d) Schematic illustration of the proposed R-NCNP nanoregulator for the light-driven water-splitting process and related reaction equations. (e) O<sub>2</sub> production curve of the R-NCNP nanoparticles in water solution. Reproduced with permission. [108] Copyright 2020, American Chemical Society.

#### 4. Core-shell structured nanoparticles for photodynamic synergistic therapy of cancer

The combination of PDT with other therapies is beneficial to improve the anticancer efficacy [197, 198]. On the one hand, photodynamic synergistic therapy can greatly avoid the side effects on healthy tissues through reducing the therapeutic dosage. On the other hand, it can tackle some of the challenging issues in monotherapy, such as metastasis of tumors and development of resistance. Core-shell structured nanoparticles are an ideal nanoplatform for photodynamic synergistic therapy on account of the convenience of incorporating functional materials or agents [55, 199, 200]. The

applications of core-shell structured nanoparticles in photodynamic synergistic therapy of cancer are listed in Table 2. Herein, core-shell structured nanoparticles for photodynamic synergistic therapy of cancer are introduced according to the combination of PDT with different therapies (e.g., chemotherapy, PTT and immunotherapy).

511

Table 2. Core-shell structured nanoparticles for photodynamic synergistic therapy of cancer and related imaging.

Nanoparticle	Excitation light	Modality	Objects		Imaging	Ref.
			<i>In vitro</i>	<i>In vivo</i>		
NCP@pyrolipid	LED (670 nm)	PDT/Chemotherapy	HNSCC135, SCC61, JSQ3 and SQ20B cells <sup>a)</sup>	SQ20B tumor-bearing mice	Optical	[201]
PTX-S-OA@PPa-PEG	Laser (660 nm)	PDT/Chemotherapy	KB <sup>b)</sup> , 4T1 <sup>c)</sup> and A549 <sup>d)</sup> cells	KB tumor-bearing mice	Optical	[128]
Au@dsDNA/G4	Laser (690 nm)	PDT/Chemotherapy	HeLa cells	HeLa tumor-bearing mice	Optical	[120]
SAD@ZIF-90	Laser (808 nm)	PDT/Chemotherapy	HeLa cells	HeLa tumor-bearing mice	Optical	[114]
UCNPs@mSiO <sub>2</sub> @ZIF-90	Laser (808 nm)	PDT/Chemotherapy	4T1 cells and HeLa cells	H22 <sup>e)</sup> tumor-bearing mice	Optical/MR	[188]
AuNR@SiO <sub>2</sub>	Laser (780 nm)	PDT/PTT	CT26 <sup>f)</sup> cells	-	Optical	[77]
AuNR@MOFs	Laser (640 nm and 808 nm)	PDT/PTT	4T1 cells	4T1 tumor-bearing mice	Optical/Photothermal	[202]
b-P25@PDA-Ce6 (Mn)	Laser (671 nm and 808 nm)	PDT/PTT	4T1 cells	4T1 tumor-bearing mice	Optical/Photothermal/MR	[203]
TiO <sub>2</sub> @RP	Laser (808 nm)	PDT/PTT	OS-RC-2 <sup>g)</sup> and 786-O cells <sup>g)</sup>	786-O tumor-bearing mice	Optical/Photothermal	[204]
HMCuS@MnO <sub>2</sub>	Laser (660 nm and 808 nm)	PDT/PTT	4T1 cells	4T1 tumor-bearing mice	Optical/Photoacoustic/MR	[97]
ZnP@pyrolipid	Laser (670 nm)	PDT/Immunotherapy	4T1 cells	4T1 tumor-bearing mice	Optical	[205]
TPPM@BioPEGDMA	Laser (660 nm)	PDT/Immunotherapy	CT26 cells	CT26 tumor-bearing mice	Optical	[206]
LiYF <sub>4</sub> :Ce@SiO <sub>2</sub> @ZnO	X-ray	PDT/Radiation therapy	HeLa cells	HeLa tumor-bearing mice	-	[153]
PEG/LDNPs@CMSNs	Laser (980 nm)	PDT/CDT	HeLa cells	HeLa tumor-bearing mice	Optical/MR/CT	[207]
mSiO <sub>2</sub> @MnO <sub>2</sub> @PEG	Laser (808 nm)	PDT/CDT	4T1 cells	4T1 tumor-bearing mice	Optical	[92]
UCNPs@mSiO <sub>2</sub> -CuS	Laser (980 nm)	PDT/PTT/ Chemotherapy	HeLa cells	H22 tumor-bearing mice	Optical/Photothermal/MR/ CT	[208]
PDA@UCNPs	Laser (980 nm)	PDT/PTT/ Immunotherapy	4T1 cells	4T1 tumor-bearing mice	Optical/Photothermal/MR	[71]
BiNS-Fe@Fe	Laser (808 nm)	PDT/PTT/CDT	HepG-2 <sup>h)</sup> cells	HepG-2 tumor-bearing mice	Optical/Photothermal/ Photoacoustic/MR/CT	[83]
UCNPs@MOFs	Laser (980 nm)	PDT/Chemotherapy/ Immunotherapy	CT26 cells	CT26 tumor-bearing mice	Optical	[42]
CDTN	Laser (671 nm)	PDT/Chemotherapy/ Gene therapy	4T1 cells	4T1 tumor-bearing mice	Optical	[136]

<sup>a)</sup>human head and neck cancer cells; <sup>b)</sup>human epidermoid cancer cells; <sup>c)</sup>mouse breast cancer cells; <sup>d)</sup>human non-small cell lung cancer cells; <sup>e)</sup>mouse liver cancer cells; <sup>f)</sup>mouse colon cancer cells; <sup>g)</sup>human clear cell renal cell carcinoma cells; <sup>h)</sup>human hepatoma cells.

515

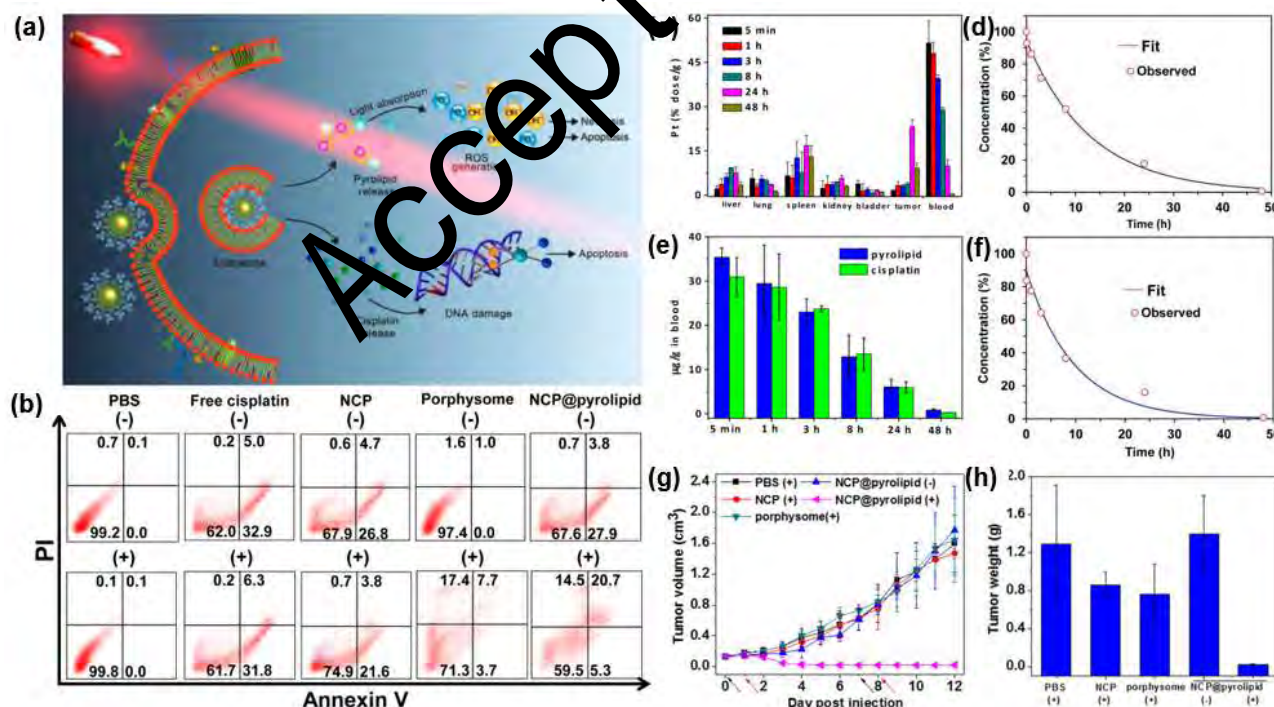
#### 516 4.1. PDT combined with chemotherapy

517 Chemotherapy is one of the most widely used cancer treatment strategies in the past few decades,  
518 which ingests chemotherapeutic drugs orally or intravenously to suppress tumor growth [209, 210].  
519 Although chemotherapy has the unique merits of eliminating cancer cells in the early stage and  
520 improving survival rate in the late stage, there are still some therapeutic limitations, such as premature  
521 drug release, severe drug resistance and side effects on healthy tissues [211, 212]. The integration of  
522 chemotherapy and PDT into a single nanoparticulate system is a promising way to solve these issues  
523 [213-215]. Specifically, nanoparticulate system can promote the delivery of small molecule drugs to  
524 tumor sites via the EPR effect, resulting in the selective distribution of drugs and low toxicity to  
525 healthy tissues. What is more, chemotherapy can increase the sensitivity of tumor cells to  
526 photoinduced ROS, while ROS in turn can restrain the activity of proteins related to drug efflux,  
527 thereby reducing the possibility of drug efflux and restoring multidrug tolerance. Recently, the  
528 development of core-shell structured nanoparticles for cancer therapy by combining PDT with  
529 chemotherapy has attracted great interest to realize superior anticancer effect [74, 181].

530 Normally, chemotherapeutic drugs can self-assemble with traditional small molecule  
531 photosensitizers to form core-shell structured nanoparticles for combined therapy of  
532 PDT/chemotherapy [216]. For example, He et al. reported a self-assembled nanoscale coordination  
533 polymer (NCP)@pyropheophorbide-lipid (pyrolipid) nanoparticle with cisplatin drug in the core and  
534 pyrolipid photosensitizer in the shell for PDT/chemotherapy [201]. As depicted in Fig. 6a, the  
535 NCP@pyrolipid kept structural integrity extracellularly, but released pyrolipid and cisplatin  
536 intracellularly, leading to the apoptosis and necrosis of cancer cells. Flow cytometry results  
537 demonstrated that the NCP@pyrolipid nanoparticles aroused the highest level of apoptosis (26.0%)  
538 and necrosis (14.5%) for SQ20B human head and neck cancer cells under 670 nm LED light



539 irradiation (Fig. 6b). Pharmacokinetic and biodistribution investigations of the NCP@pyrolipid  
 540 nanoparticles in CT26 tumor-bearing mice indicated that the pyrolipid and cisplatin presented low  
 541 uptake in normal organs, high tumor accumulation and extended blood circulation times (Fig. 6c-f).  
 542 By virtue of the synergistic effect of PDT and chemotherapy, the NCP@pyrolipid nanoparticles  
 543 exhibited superior antitumor effect (both in tumor volume and weight) for human head and neck  
 544 cancer SQ20B xenograft mice compared to monotherapy (Fig. 6g and h). In the study of Chen et al.,  
 545 an antitumor drug paclitaxel (PTX) was utilized to induce the self-assembly of Ce6 photosensitizer-  
 546 modified human serum albumin (HSA) and acyclic Arg-Gly-Asp (cRGD<sub>5</sub>K) peptide-modified HSA  
 547 [217]. The self-assembled nanoparticle was composed of a Ce6/PTX-HSA core and a RGD/PTX-  
 548 HSA shell. Both *in vitro* and *in vivo* studies proved that the Ce6/PTX-HSA@RGD/PTX-HAS  
 549 nanoparticles could not only target  $\alpha v \beta 3$ -integrin, but also realize PDT/chemotherapy combination,  
 550 which significantly enhanced the therapeutic efficacy for cancer.



552 **Fig. 6.** (a) Proposed cytotoxicity mechanism of the NCP@pyrolipid nanoparticles. (b) Flow cytometry showing  
 553 the apoptosis and necrosis induced by the NCP@pyrolipid nanoparticles upon irradiation. (c) Tissue distributions

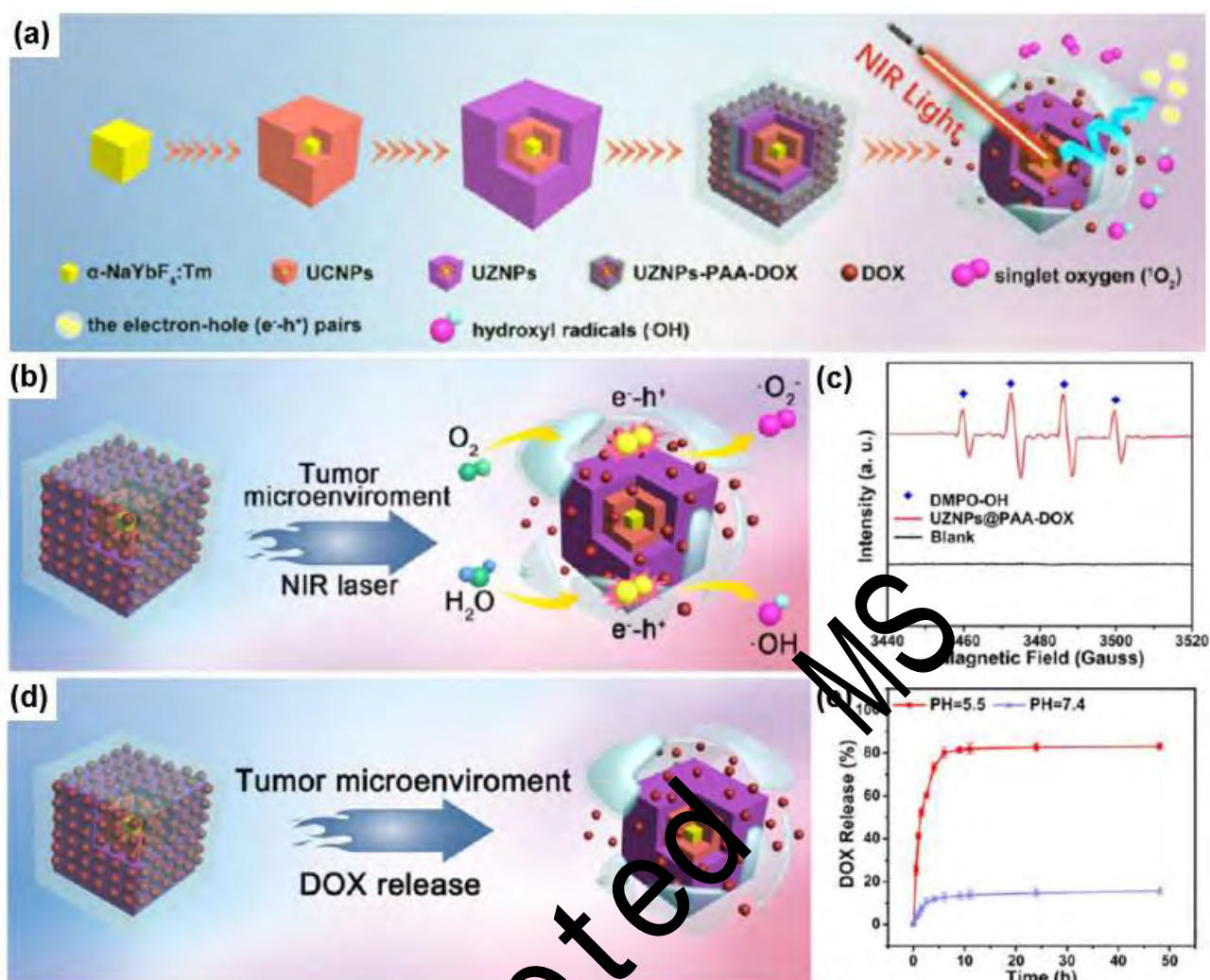
of Pt at different time points after intravenous injection of the NCP@pyrolipid nanoparticles. (d) Observed and fitted time-dependent Pt concentrations in blood following the NCP@pyrolipid administration by one-compartment model. (e) Time-dependent pyrolipid and cisplatin concentrations in blood after intravenous injection of the NCP@pyrolipid nanoparticles. (f) Observed and fitted time-dependent pyrolipid concentrations in blood following the NCP@pyrolipid administration by one-compartment model. (g) Tumor growth inhibition curves. (h) Weights of excised tumors on Day 12. Reproduced with permission. [201] Copyright 2015, American Chemical Society.

Chemotherapeutic drugs can be loaded into core-shell structured nanoparticles together with traditional small molecule photosensitizers for combined therapy of PDT/chemotherapy. For example, Wang et al. assembled the ZnPc photosensitizer and DOX on the surface of UCNPs@mSiO<sub>2</sub>-CuS nanoparticles for PDT and chemotherapy functions respectively [208]. Benefiting from the synergistic effect of PDT and chemotherapy, the UCNPs@mSiO<sub>2</sub>-CuS-ZnPc-DOX nanoparticles showed superior antitumor efficiencies both *in vitro* and *in vivo*. With the rapid development of photosensitizers, chemotherapeutic drugs are also loaded into core-shell structured nanoparticles that can directly act as photosensitizers. In the study of Yang et al., DOX was loaded into the UCNPs@MIL-100(Fe) nanoparticles for PDT/chemotherapy [218]. The MIL-100(Fe) shell not only served as a photosensitizer to produce ROS under irradiation, but also loaded a large amount of DOX due to its porous structure and high specific area. In order to better regulate their interactions, chemotherapeutic drugs and photosensitizers are placed in different layers independently. Peng et al. prepared the dual-template imprinting polymer nanoparticles for targeted PDT/chemotherapy by encapsulating gadolinium-doped silicon quantum dots and Ce6 photosensitizer in fluorescent SiO<sub>2</sub>(FSiO<sub>2</sub>) core and loading DOX and epitope into 3-methacryloxypropyltrimethoxysilane (MPS) shell [219]. Under 655 nm laser irradiation, the implanted Ce6 photosensitizer generated <sup>1</sup>O<sub>2</sub> to kill

577 cancer cells, combining with the embedded DOX to achieve a synergistic treatment.

578 Notably, the controllable release of drugs in core-shell structured nanoparticles is critical to  
579 obtain a desirable therapeutic efficacy [220, 221]. Since tumor microenvironment presents slight  
580 acidity, some pH-responsive core-shell structured nanoparticles have been designed to precisely  
581 control the release of drugs in tumors [74, 188]. For example, Cai et al. constructed pH-responsive  $\alpha$ -  
582 NaYbF<sub>4</sub>:Tm@CaF<sub>2</sub>:Nd@ZnO (UZNPs)-polyacrylic acid (PAA)-DOX nanoparticles for  
583 PDT/chemotherapy (Fig. 7a) [84]. Upon 808 nm NIR laser excitation, the nanoparticles were induced  
584 to generate electron-hole pairs, which subsequently reacted with O<sub>2</sub> and H<sub>2</sub>O to produce  $\cdot\text{O}_2^-$  and  $\cdot\text{OH}$   
585 respectively for cancer therapy (Fig. 7b). As shown in Fig. 7c, the electron paramagnetic resonance  
586 tests also confirmed the ROS generation. Moreover, the PAA coating could load abundant DOX and  
587 decompose at mild acidic tumor microenvironment to release DOX (Fig. 7d). At acid buffer  
588 dispersion with pH of 5.5, the nanoparticles released about 82% of DOX in the first 8 h, verifying the  
589 superior pH-activable ability (Fig. 7e). In addition to pH-responsive core-shell structured  
590 nanoparticles, some ROS-responsive core-shell structured nanoparticles have also been developed to  
591 regulate the release of drugs due to the large number of ROS generated during PDT. In the study of  
592 Lee et al., a chitosan shell was coated on a ROS-generating PhA-linked poly(hydroxyethyl  
593 methacrylate) (poly-HEMA) core, and then linked to an anticancer drug 5'-deoxy-5-fluorocytidine  
594 (DFCR) through phenylboronic acid to form a ROS cleavable boronic ester for PDT/chemotherapy  
595 [222]. Sun et al. fabricated a ROS-responsive nanoparticle composed of a single thioether-bridged  
596 paclitaxel (PTX)-oleic acid (OA) prodrug (PTX-S-OA) core and a pyropheophorbide a (PPa)-  
597 polyethylene glycol 2000 (PEG<sub>2k</sub>) shell for PDT/chemotherapy [128]. Under laser irradiation, the  
598 ROS generated by PPa-PEG<sub>2k</sub> shell not only were used for PDT, but also promoted the release of  
599 PTX from PTX-S-OA in combination with endogenous ROS.





**Fig. 7.** (a) Schematic illustration for the preparation of UZNPs-PAA-DOX nanoparticles. (b) Mechanism of 808 nm NIR laser irradiation-triggered PDT over the UZNPs-PAA-DOX nanoparticles. (c) Electron paramagnetic resonance spectra of the UZNPs-PAA-DOX nanoparticles. (d) Schematic illustration for the pH-responsive drug release of UZNPs-PAA-DOX nanoparticles. (e) DOX release profile of the UZNPs-PAA-DOX nanoparticles in PBS with different pH values. Reproduced with permission. [84] Copyright 2020, American Chemical Society.

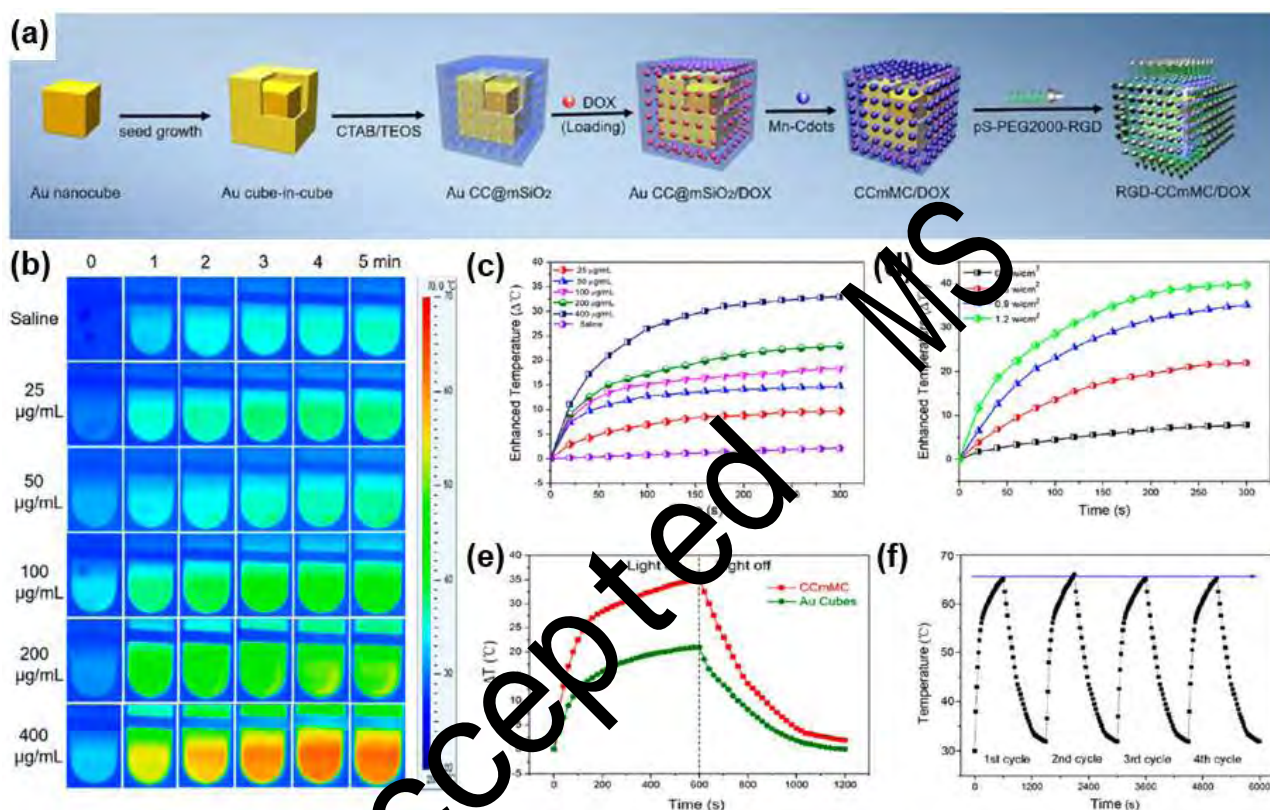
#### 4.2. PDT combined with photothermal therapy

Photothermal therapy (PTT) is a promising noninvasive cancer treatment method that exploits the NIR light and photothermal agents to convert light energy to heat energy, which can effectively destroy tumor tissues and cells [223-225]. Because of the similar photoactivation conditions to PDT and the ability to overcome imperfections of PDT, PTT has been extensively employed to combine

611 with PDT to maximize the curative effect for cancer [204, 226, 227]. In this synergistic therapeutic  
612 modality, the appropriate photothermal effect can increase the permeability of cell membranes,  
613 thereby promoting the efficient absorption and penetration of tumor cells to nanoparticles. Meanwhile,  
614 it can also accelerate the blood flow velocity in tumor and hence transport more O<sub>2</sub> to attenuate the  
615 tumor hypoxia. Recently, a series of NIR light absorbing nanomaterials have been applied to construct  
616 core-shell structured nanoparticles for combined therapy of PDT/PTT, such as gold nanostructures  
617 (e.g., nanorods [202, 228] and nanocages [229, 230]), PDA [66, 71, 231], Nd<sup>3+</sup>-doped UCNPs [232,  
618 233], black TiO<sub>2</sub> [166, 203] and copper sulfide [208, 234, 235].

619 Among them, gold nanostructures are considered to have great potential in PDT/PTT owing to  
620 their SPR induced excellent photothermal effect [236-238]. For example, Qin et al. coated the AuNR  
621 with a HB photosensitizer-incorporated mSiO<sub>2</sub> shell and a folate-modified lipid (LF) bilayer for  
622 PDT/PTT [57]. The AuNR@mSiO<sub>2</sub>-HB@LF nanoparticles possessed a strong SPR peak at 801 nm,  
623 which was expected to achieve PTT under NIR light irradiation. After being irradiated by a 808 nm  
624 laser (1.5 W cm<sup>-2</sup>) for 5 min, the temperature of AuNR@mSiO<sub>2</sub>-HB@LF suspension (0.1 mg mL<sup>-1</sup>)  
625 increased by about 50 °C, which was enough to kill the tumor cells. Moreover, the yield of  
626 photoinduced ROS was enhanced by hyperthermia. Therefore, the AuNR@mSiO<sub>2</sub>-HB@LF  
627 nanoparticles could significantly eliminate the MCF-7 tumor in BALB/c nude mice because of the  
628 synergistic effect of PDT and PTT. To improve the photothermal conversion efficiency and  
629 photothermal stability of gold nanostructures, Zhang et al. synthesized gold cube-in-cubes for  
630 developing the CCmMC PDT/PTT agent [190]. The CCmMC nanovehicles were constructed by  
631 loading Mn-Cdots on gold cube-in-cubes@mSiO<sub>2</sub> core-shell structured nanoparticles (Fig. 8a). As  
632 displayed in Fig. 8b-d, the temperature of CCmMC suspension increased as the increase of CCmMC  
633 concentration and irradiation power density, implying the high NIR light-induced photothermal effect

634 of CCmMC. Fig. 8e and f further demonstrated that the CCmMC possessed a superior photothermal  
 635 conversion efficiency of 65.6% and excellent photothermal stability upon 808 nm laser excitation.  
 636 After coupling with the Mn-Cdots-induced favorable PDT effect, the CCmMC exhibited desirable  
 637 therapy efficacy in treating 4T1 tumor xenografts on nude mice under the dual laser (635 and 808 nm)  
 638 irradiation.



**Fig. 8.** (a) Schematic illustration for the preparation of RGD-CCmMC/DOX nanoparticles. (b) Thermographic images of the CCmMC aqueous solutions irradiated with 808 nm laser at a power intensity of 1 W cm<sup>-2</sup>. (c) Photothermal conversion characterizations of the CCmMC aqueous solution of various concentrations under 1 W cm<sup>-2</sup> 808 nm laser irradiation. (d) Temperature elevation profiles of the CCmMC solutions under various laser power densities. (e) Temperature changes in CCmMC and Au Cube aqueous solutions in response to NIR laser on and off. (f) Temperature curves of CCmMC under continuous NIR laser irradiation for 4 cycles. Reproduced with

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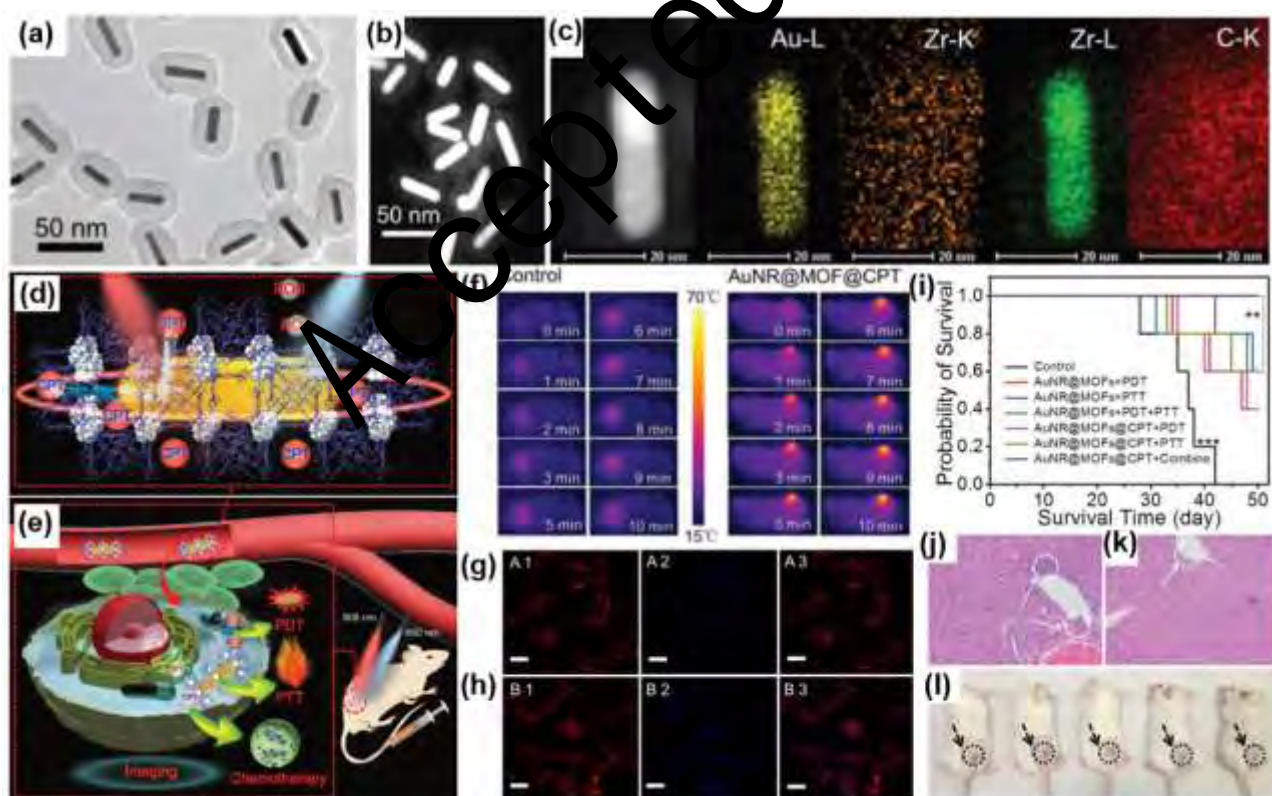
PDA has also been widely employed as a PTT agent in combined therapy of PDT/PTT due to its

648 pronounced absorption in NIR region, satisfactory photothermal conversion capacity and outstanding  
 649 biocompatibility [239, 240]. In the study of Cen et al., a PDA shell was coated on the methylene blue  
 650 (MB)-loaded UCNP@SiO<sub>2</sub> nanoparticles for PDT/PTT [241]. The temperature of UCNP@SiO<sub>2</sub>-  
 651 MB@PDA suspension (0.2 mg mL<sup>-1</sup>) rose to 52.2 °C after 10 min of 980 nm laser (1.5 W cm<sup>-2</sup>)  
 652 irradiation, indicating its great photothermal conversion ability. Through the FRET from UCNP to  
 653 MB photosensitizer and PDA, the UCNP@SiO<sub>2</sub>-MB@PDA nanoparticles presented excellent  
 654 PDT/PTT synergistic effect for killing the cancer cells under 980 nm laser irradiation. Yang et al.  
 655 utilized PDA core as the template to prepare lactose acid (LA)-grafted PDA@cobalt phytate (CoPA)  
 656 nanoparticles for PDT/PTT [242]. Benefiting from the PDA-endowed PTT effect, CoPA-induced  
 657 PDT effect and LA-endued targeting capability, the PDA@CoPA-LA nanoparticles exhibited superior  
 658 antitumor performances both *in vitro* and *in vivo*. Moreover, the abundant amino and catechol groups  
 659 on the surface of PDA make it easy to be modified by various functional biomolecules [243]. Zeng et  
 660 al. improved the performance of targeted breast cancer treatment in PDT/PTT by introducing FA  
 661 molecules on the surface of MnO<sub>2</sub>-CeO<sub>2</sub>@PDA nanoparticles [126].

662 In addition to the combinational therapeutic modality of PDT/chemotherapy and PDT/PTT  
 663 summarized above, tri-modal PDT/PTT/chemotherapy has been developed to further lower laser  
 664 power and reduce drug dosage in cancer treatment. For example, Zeng et al. employed AuNR as the  
 665 seed crystal to prepare AuNR@MOFs@camptothecin (CPT) nanoparticles for  
 666 PDT/PTT/chemotherapy (Fig. 9a-e) [73]. As shown in the *in vivo* photothermal images (Fig. 9f), the  
 667 temperature of AuNR@MOFs@CPT nanoparticles-injected tumor increased quickly from 28.5 to  
 668 48.4 °C after 2 min of 808 nm laser irradiation, and then reached a steady temperature of 54.8 ± 1.2 °C ,  
 669 which was sufficient to cause the death of cancer cells. Meanwhile, the photothermal effect of AuNR  
 670 could also accelerate the intracellular release of CPT (Fig. 9g and h). By virtue of the synergistic



671 effect of photoinduced ROS, photothermal effect and released CPT, the combined therapy  
 672 significantly raised the survival rate of 4T1 tumor-bearing mice (Fig. 9i). Furthermore, the  
 673 AuNR@MOFs@CPT nanoparticles restrained the hepatic metastases because of its accumulation in  
 674 liver and tumor position (Fig. 9j and k). And after 50 d of treatment of mice with  
 675 AuNR@MOFs@CPT nanoparticles, the tumors almost completely disappeared (Fig. 9l).  
 676 Additionally, Chen et al. designed ROS-responsive PPID nanoparticles which was composed by self-  
 677 assembly of a IR780 photosensitizer and DOX co-loaded poly( $\beta$ -amino ester) core and a propylene  
 678 glycol alginate sodium sulfate shell for PDT/PTT/chemotherapy [244]. The PPID nanoparticles could  
 679 greatly improve the PDT and PTT performances of IR780 *in vitro* and further promote the  
 680 internalization of IR780 and DOX in Hep1-6 cells. Compared with free IR780 and free DOX, the  
 681 PPID nanoparticles showed synergistic cytotoxicity in Hep1-6 cells under 808 nm laser irradiation.



**Fig. 9.** (a) TEM image, (b) STEM-HAADF image and (c) EDX elemental mapping images of the AuNR@MOFs nanoparticles. (d) The structure of AuNR@MOFs@CPT nanoparticles. (e) The combined

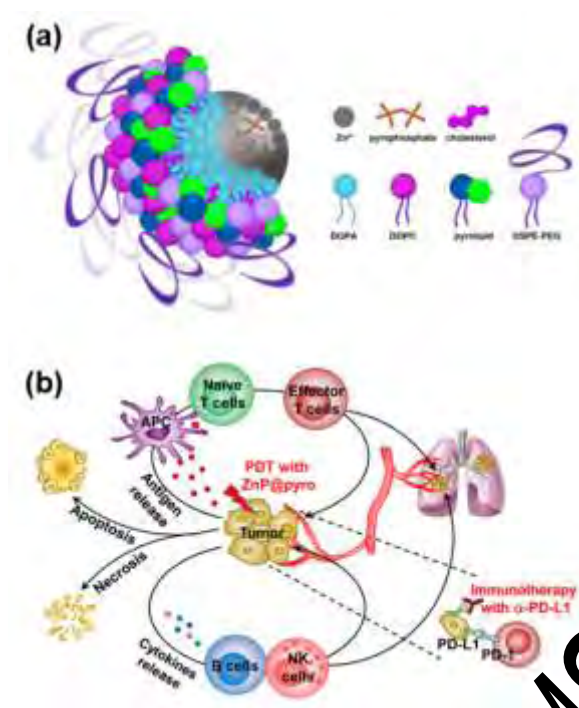
PDT/PTT/chemotherapy of tumor. (f) The *in vivo* thermal images of the mice after intravenous injection of PBS and AuNR@MOFs@CPT with 808 nm laser irradiation. (g) The intracellular drug release behavior of AuNR@MOFs@CPT in dark and (h) under 808 nm laser irradiation. (i) Survival curves of tumor-bearing mice after different treatments ( $n = 5$ ,  $**p < 0.01$  and  $***p < 0.01$  were calculated by a Student's *t* test). (j) H&E staining of liver after intravenous injection of PBS and (k) AuNR@MOFs@CPT on day 18. (l) The photograph of tumor-bearing mice treated with AuNR@MOFs@CPT for combined therapy after 50 d. Reproduced with permission. [73] Copyright 2017, Wiley-VCH.

#### 4.3. PDT combined with immunotherapy

PDT can not only kill tumor cells directly, but also induce immunogenic cell death (ICD) of tumor cells, thereby promoting the maturation of dendritic cells and activation of effector cells, and ultimately leading to a systemic antitumor immune response [245-247]. Recent studies have reported that some core-shell structured nanoparticles can evoke an antitumor immune response during the process of PDT to enhance the therapeutic efficacy for cancer [206, 230]. For example, in the study of Liang et al., in addition to efficiently destroying 4T1 breast cancer cells, the abundant ROS generated by gold nanocage@MnO<sub>2</sub> nanoparticles under laser irradiation also triggered the ICD-mediated antitumor immune response [229]. Specifically, the dying cancer cells released damage associated molecular patterns (e.g., calreticulin, adenosine triphosphate and high mobility group protein B1) for the dendritic cells maturation. And then the specific effector cells (e.g., CD4<sup>+</sup> T cells, CD8<sup>+</sup> T cells and NK cells) were activated to prevent the tumor growth and metastasis. Unfortunately, the immune response induced by PDT is usually mild and not enough to completely suppress the tumor metastasis. Immunosuppressive tumor microenvironment may significantly depress the PDT-induced immunotherapy efficacy through the immune checkpoint pathway [198, 248].

As an effective cancer treatment method with low side effects, immunotherapy kills tumor cells

708 by activating the body's own immune system [249]. Among them, checkpoint blockade  
709 immunotherapy has attracted much attention, which exploits inhibitor molecules to target the  
710 regulatory pathways in T cells for modulating immunosuppressive tumor microenvironment and  
711 enhancing antitumor immune response [250, 251]. Especially, the programmed death 1/programmed  
712 death ligand 1 (PD-1/PD-L1) blockade has already been approved by the U.S. Food and Drug  
713 Administration (FDA) to treat diverse tumors [252, 253]. Although checkpoint blockade  
714 immunotherapy has achieved clinical success, it is only effective in tumors pre-infiltrated by T cells.  
715 Accordingly, PDT that can induce ICD of tumor cells may improve its efficacy. In this case, the  
716 combined therapy of PDT/immunotherapy based on core-shell structured nanoparticles has the  
717 potential to promote the primary tumors destruction and distant metastatic tumors control [254]. In  
718 the study of Duan et al., the Zn-pyrophosphate (ZnP@pyrolipid) nanoparticles (Fig. 10a) were  
719 fabricated to combine PDT with checkpoint blockade immunotherapy for the treatment of metastatic  
720 breast cancer [205]. As depicted in Fig. 10b, the combination of ZnP@pyrolipid-mediated PDT with  
721 PD-L1 antibody ( $\alpha$ -PD-L1)-mediated immunotherapy not only destroyed the primary tumors but also  
722 remarkably inhibited the metastasis to lung in a 4T1 mTNBC murine model. Compared to the PBS  
723 control group, the ZnP@pyrolipid-mediated PDT reduced the 4T1 tumor by 68% in volume and 75%  
724 in weight. But after the introduction of  $\alpha$ -PD-L1, the 4T1 tumor was completely eradicated. Moreover,  
725 the results of gross examination of lung tumor nodules demonstrated that the combined therapy was  
726 much more effective than ZnP@pyrolipid-mediated PDT or  $\alpha$ -PD-L1-mediated immunotherapy  
727 alone in restraining lung metastasis.

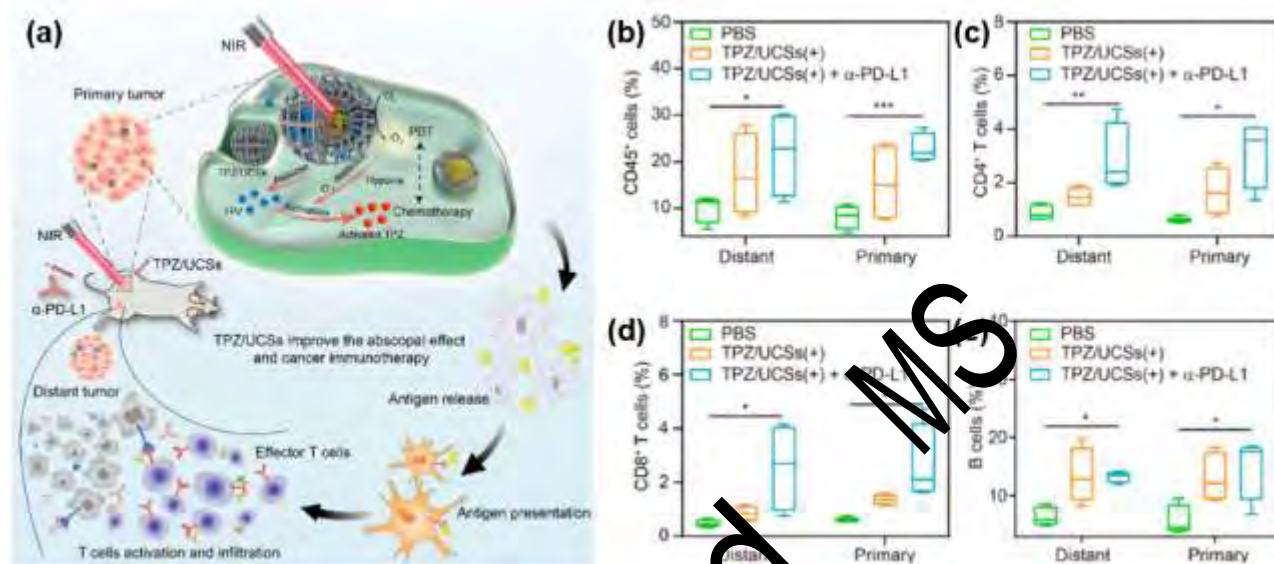


**Fig. 10.** (a) Scheme showing the Zn-pyrophosphate core and the asymmetric lipid bilayer shell of ZnP@pyrolipid nanoparticles. (b) Immunogenic ZnP@pyrolipid PDT sensitizes tumors to PD-L1 blockade immunotherapy for the treatment of metastatic tumors. Reproduced with permission [405]. Copyright 2016, American Chemical Society.

To further enhance the therapeutic efficacy for cancer, tri-modal therapeutic approach has been developed on the core-shell structured nanoparticles, such as PDT/chemotherapy/immunotherapy [42, 255] and PDT/PTT/immunotherapy [71]. As shown in Fig. 11a, Shao et al. constructed tirapazamine (TPZ)-encapsulated UCSs nanoparticles to combine PDT/chemotherapy with checkpoint blockade immunotherapy for the treatment of hypoxic tumors [42]. The combined therapy effectively inhibited the growth of primary tumors and distant tumors in CT26 tumor-bearing mice both in tumor volume and weight. Meanwhile, in the combined therapy group, the percentages of infiltrating CD45<sup>+</sup> cells, CD4<sup>+</sup> T cells, CD8<sup>+</sup> T cells and B cells were increased in both primary tumors and distant tumors (Fig. 11b-e), indicating that the combination of TPZ/UCSs-mediated PDT/chemotherapy with α-PD-L1-mediated immunotherapy improved the immunotherapeutic efficacy through the infiltration of effector T cells. Besides, in the study of Yan et al., PDA@UCNPs-PEG/Ce6 nanoparticles were



743 assembled to combine PDT/PTT with  $\alpha$ -PD-L1-mediated immunotherapy for inhibiting the tumor  
 744 metastasis and relapse [71]. In the combined therapy group, most of the 4T1 tumor-bearing mice  
 745 could survive 100 days, and the survival rate was almost as high as 77.8%, which was much higher  
 746 than that of the control groups.



**Fig. 11.** (a) Schematic illustration of the structure of TPZ/UCSs nanoparticles and their application to tumor

treatment through a combination of NIR light-triggered PDT and hypoxia-activated chemotherapy with

immunotherapy. Percentage of tumor-infiltrating (b) CD45<sup>+</sup> cells, (c) CD4<sup>+</sup> T cells, (d) CD8<sup>+</sup> T cells and (e) B

cells in total tumor cells. Data are means  $\pm$  SD,  $n = 4$ . \* $p < 0.05$ , \*\* $p < 0.01$  and \*\*\* $p < 0.001$ . Reproduced with

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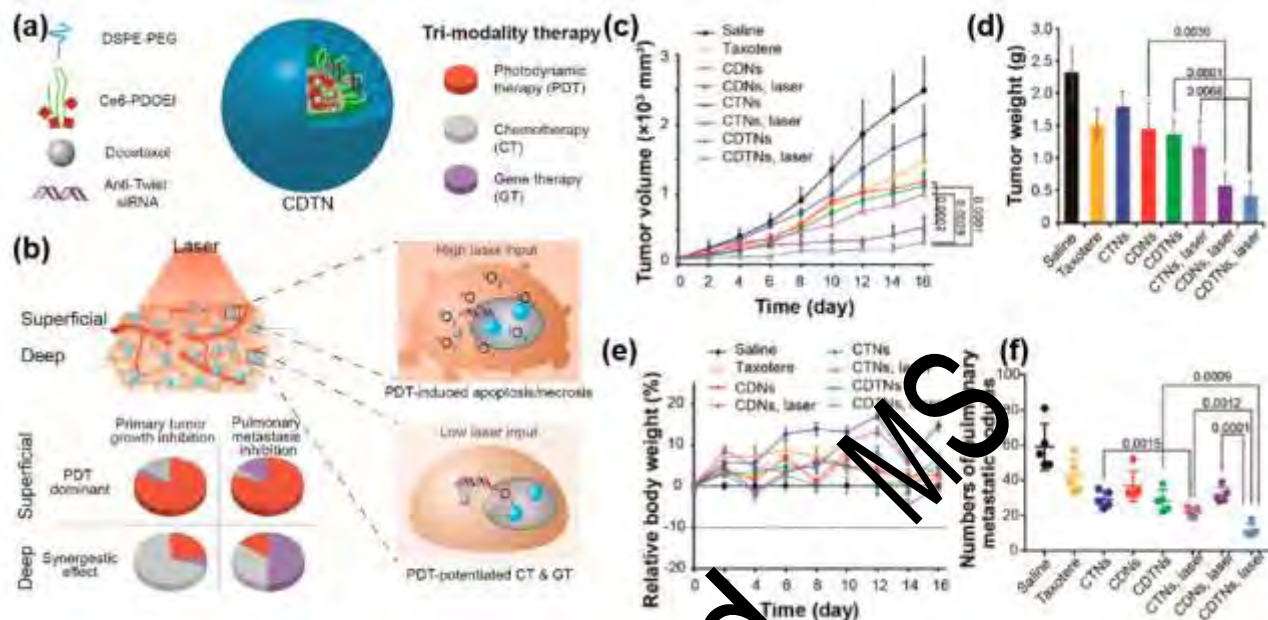
#### 4.4. PDT combined with other therapies

In addition to chemotherapy, PTT and immunotherapy, PDT can also be combined with other therapies, such as radiotherapy [256, 257], gene therapy [258, 259] and chemodynamic therapy [260, 261], to enhance the therapeutic efficacy. With the development of nanotechnology, some core-shell structured nanoparticles have been designed to combine PDT with these therapies. Radiotherapy is a conventional cancer treatment method, which utilizes ionizing radiation to control or kill tumor cells and is not limited by the tissue penetration depth [262]. Benefiting from the inherent antitumor

760 efficacy and strong penetration ability of ionizing radiation, the combined therapy of  
761 PDT/radiotherapy that employs a single excitation source will present a great clinical significance  
762 [263]. In the study of Zhang et al., LiYF<sub>4</sub>:Ce@SiO<sub>2</sub>@ZnO nanoparticles were fabricated for  
763 synchronous PDT and radiotherapy under X-ray radiation [153]. The growth of tumor treated by  
764 PDT/radiotherapy was almost completely suppressed after 15 days, while the growth of tumor treated  
765 by radiotherapy alone was only slightly inhibited, implying the excellent synergistic effect of PDT  
766 and radiotherapy.

767 Gene therapy is a promising cancer treatment method, which delivers therapeutic nucleic acids  
768 into the tumor cells to correct or compensate cancers caused by genetic defects and anomalies [264].  
769 Among various therapeutic nucleic acids, small interfering RNAs (siRNAs) that can intracellularly  
770 silence disease-causing genes have attracted tremendous interest since they can remarkably improve  
771 the specificity and efficacy of gene therapy [265, 266]. Nevertheless, the cellular impermeability and  
772 easy degradability of siRNAs hinder their transfer into tumor cells. A recent study suggested that the  
773 core-shell structured photodynamic nanoparticle is an excellent carrier that can simultaneously  
774 deliver chemotherapeutic drugs and siRNAs into tumor cells for photodynamic synergistic therapy  
775 [136]. As shown in Fig. 12a, core-shell structured nanoparticles named CDTNs were designed via  
776 self-assembly of 1,2-distearoyl-*sn*-glycero-3-phosphoethanolamine-*N*-[maleimide-(polyethylene  
777 glycol)<sub>5000</sub>] (DSPE-PEG), poly- $\beta$ -aminoester derivative Ce6-grafted poly[(1,4-butanediol)-diacrylate-  
778  $\beta$ -oligoethylenimine<sub>600</sub>] (Ce6-PDOEI), docetaxel and anti-Twist siRNA for tri-modal  
779 PDT/chemotherapy/gene therapy of metastatic triple-negative breast cancer (mTNBC). In the  
780 superficial part of the tumor, CDTNs eliminated the primary tumor and inhibited its pulmonary  
781 metastasis mainly through PDT. While in the deep part of the tumor, CDTNs eliminated the primary  
782 tumor mainly through PDT-potentiated chemotherapy and inhibited its pulmonary metastasis through

783 PDT-potentiated gene therapy and chemotherapy (Fig. 12b). Therefore, compared with the  
 784 monotherapy and dual-modal therapy, the CDTNs exhibited superior efficacy in inhibiting the growth  
 785 of the primary tumor and its pulmonary metastasis (Fig. 12c-f).



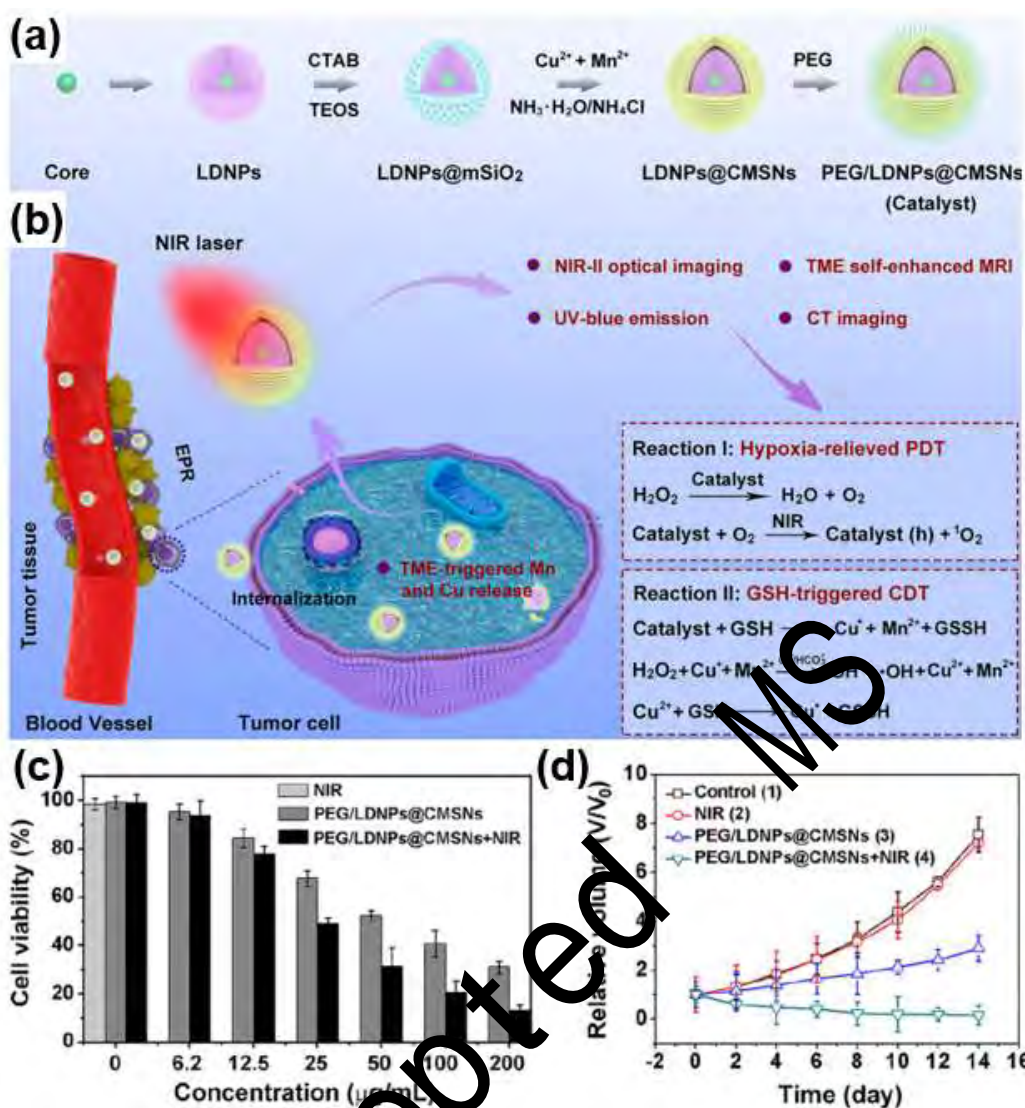
786  
 787 **Fig. 12.** (a) Schematic illustration of design and function of CDTNs. (b) Depth-dependent mechanism of CDTNs  
 788 against mTNBC. (c) Growth profiles and (d) weights of primary TNBC tumors in mice receiving different  
 789 treatments. (e) Changes in body weights of 4T1-bearing mice. (f) Comparison of pulmonary metastatic nodules of  
 790 4T1 tumor-bearing mice treated with various treatments. Reproduced with permission. [136] Copyright 2018,  
 791 American Chemical Society.

792 Chemodynamic therapy (CDT) is a novel cancer treatment method, which exploits transition  
 793 metals and weak acidic TME to catalyze overexpressed endogenous  $\text{H}_2\text{O}_2$  to produce  $\cdot\text{OH}$  for killing  
 794 cancer cells [267, 268]. Since  $\text{O}_2$  and  $\cdot\text{OH}$  can be generated during CDT through Fenton/Fenton-like  
 795 reactions, the tumor hypoxia will be alleviated and the therapeutic efficacy of PDT will be enhanced  
 796 [269, 270]. Accordingly, core-shell structured nanoparticles have been employed to combine PDT  
 797 with CDT for synergistic therapy more recently. As depicted in Fig. 13a, Xu et al. constructed PEG-  
 798 modified lanthanide-doped nanoparticles@copper/manganese silicate nanospheres

799 (PEG/LDNPs@CMSNs) for PDT/CDT synergistic therapy [207]. The CMSNs alleviated tumor  
800 hypoxia by decomposing  $\text{H}_2\text{O}_2$  to generate  $\text{O}_2$ , and served as photosensitizers to utilize the  $\text{O}_2$  to  
801 generate  $^1\text{O}_2$  upon NIR laser excitation for PDT. Meanwhile, the tumor glutathione (GSH)-triggered  
802 release of Fenton-like  $\text{Mn}^{2+}$  and  $\text{Cu}^+$  ions led to CDT by inducing the generation of  $\cdot\text{OH}$  (Fig. 13b).  
803 Benefiting from the synergistic effect of PDT and CDT, the PEG/LDNPs@CMSNs displayed  
804 superior antitumor effects both *in vitro* and *in vivo* under 980 nm NIR laser irradiation (Fig. 13c and  
805 d). Moreover, in the study of Qi et al.,  $\text{NaGdF}_4\text{:Er,Yb@NaGdF}_4\text{:Nd@Cu(II)}$  boron-imidazolate  
806 frameworks (CSNPs@Cu-BIF) nanoparticles were assembled for PDT/PDT/CDT synergistic therapy  
807 [233]. Upon 808 nm NIR laser excitation, the nanoparticles exhibited enhanced antitumor efficacy  
808 for *in vitro* MCF-7 cancer cells and *in vivo* MCF-7 tumor-bearing nude mice.

Accepted





**Fig. 13.** (a) Schematic illustration of the synthesis of PEG/LDNPs@CMSNs and (b) the theranostic mechanism of PEG/LDNPs@CMSNs for TME and NIR laser co-enabled PDT/CDT and trimodal bioimaging. (c) Viabilities of HeLa cells in the control group and treated with NIR, PEG/LDNPs@CMSNs and PEG/LDNPs@CMSNs plus NIR. (d) Variations in the relative tumor volume achieved from the mice under different treatments. Reproduced with permission. [207] Copyright 2020, American Chemical Society.

## 5. Core-shell structured nanoparticles for imaging in PDT-based cancer treatment

Molecular imaging techniques play a vital role in diagnosis and treatment of cancer. In recent years, a variety of molecular imaging techniques, such as optical imaging, photothermal imaging, photoacoustic imaging, magnetic resonance imaging and computed tomography imaging have been

819 employed for the imaging in PDT-based cancer treatment [271-273]. Notably, core-shell structured  
820 nanoparticles are widely used in the PDT-based cancer treatment, and can be used as effective contrast  
821 agents for the imaging in PDT-based cancer treatment. Compared with single-component  
822 nanoparticles, core-shell structured nanoparticles present unique imaging behavior as they possess  
823 combinatorial characteristics of both core and shell materials, which is conducive to multimodal  
824 imaging [121, 200, 274].

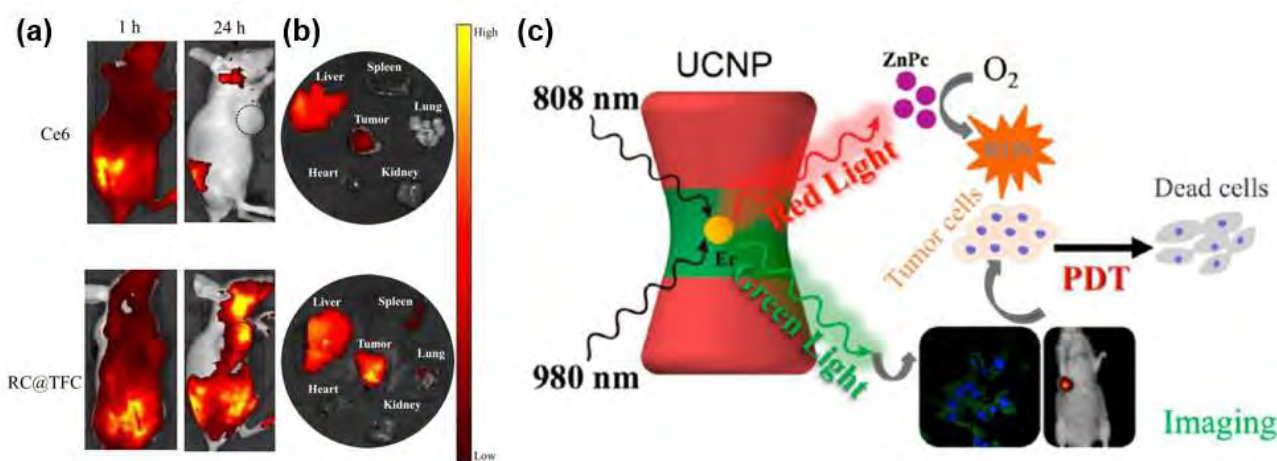
### 825 5.1. Optical imaging

826 Optical imaging is a noninvasive and safe imaging strategy, which utilizes the inherent  
827 luminescent property of the nanomaterials [275, 276]. Some photosensitizers in the excited state not  
828 only produce cytotoxic ROS, but also emit luminescence when they back to the ground state [277].  
829 Consequently, in addition to being employed for PDT, these photosensitizers can also be utilized for  
830 the optical imaging in PDT-based cancer treatment. In recent years, various small molecule  
831 photosensitizers, such as PhA [278, 279], Ce6 [97, 103] and ICG [68, 78], have been encapsulated  
832 into core-shell structured nanoparticles for fluorescence imaging-guided PDT. In the study of Liu et  
833 al., the biodistribution of RC@TFC nanoparticles in mice bearing subcutaneous MDA-MB-231  
834 tumors could be easily tracked through an ex vivo imaging system because of the intrinsic  
835 fluorescence of Ce6 [129]. The results of fluorescent imaging displayed that both free Ce6 and  
836 RC@TFC nanoparticles were extensively distributed throughout the mouse after 1 h of injection (Fig.  
837 14a). After 24 h, most of the free Ce6 was cleared from the mouse, while the RC@TFC nanoparticles  
838 accumulated in the tumor site and showed strong fluorescence, which might be ascribed to the EPR  
839 effect (Fig. 14b). Similarly, loading ICG photosensitizer through electrostatic adsorption on the gold  
840 nanocage@MnO<sub>2</sub>-hyaluronic acid nanoparticles to realize the fluorescence emission for fluorescence  
841 imaging-guided PDT was reported by He et al. [230] In addition, quantum dots (QDs) are utilized for

fluorescence imaging, but they are limited by the low water solubility and tendency to photooxidation [280]. Core-shell structured photodynamic nanoparticles can efficiently encapsulate the QDs to minimize these limitations. Hence these core-shell structured nanoparticles possessed excellent ability for the fluorescence imaging in PDT-based cancer treatment [108, 190, 219].

Benefiting from the exceptional photophysical properties, UCNPs also present good fluorescence imaging ability [281]. After coating them with suitable materials and photosensitizers, these core-shell structured nanoparticles can be applied in the fluorescence imaging-guided PDT. Wang et al. prepared UCNPs@SiO<sub>2</sub>(MB)@mSiO<sub>2</sub>(RhB)- $\beta$ -cyclodextrin nanoparticles for simultaneous fluorescence imaging, PDT and drug delivery [282]. Upon 980 nm NIR laser excitation, the UCNPs core emitted 540 nm green light for fluorescence imaging and 660 nm red light to activate photosensitizers for <sup>1</sup>O<sub>2</sub> generation. In the study of Tang et al., a mSiO<sub>2</sub> shell was decorated on UCNPs and then the ZnPc photosensitizer was incorporated into the mSiO<sub>2</sub> shell to construct a fluorescence imaging-guided PDT theranostic nanoplatfoms [56]. In this nanoplatfoms, the green emission excited by the 808 nm NIR laser was used for real time imaging, while the red emission excited by the 980 nm NIR laser was used to produce ROS for PDT (Fig. 14c). Furthermore, compared with the traditional fluorescence imaging in the first NIR window (NIR-I, 700-900 nm), recently developed fluorescence imaging in the second NIR window (NIR-II, 1000-1700 nm) possess deeper tissue penetration ability, better spatial resolution and higher signal-background-ratio [283-285]. For example, Wang et al. fabricated UCNPs@mSiO<sub>2</sub>(Ce6, atovaquone)@MnO<sub>2</sub> nanoparticles for NIR-II fluorescence imaging-guided PDT [286]. Upon 808 nm NIR laser excitation, the nanoparticles emitted intense NIR light: 1060 nm (Nd: <sup>4</sup>F<sub>3/2</sub> → <sup>4</sup>I<sub>11/2</sub>), 1350 nm (Nd: <sup>4</sup>F<sub>3/2</sub> → <sup>4</sup>I<sub>13/2</sub>) and 1520 nm (Er<sup>3+</sup>: <sup>4</sup>I<sub>11/2</sub> → <sup>4</sup>I<sub>15/2</sub>), which promoted the NIR-II fluorescence imaging.





**Fig. 14.** (a) *In vivo* (1 vs 24 h post injection) and (b) *ex vivo* (24 h post injection) fluorescent imaging of MDA-MB-231 tumor bearing mice treated with free Ce6 and RC@TFC. Reproduced with permission. [129] Copyright 2019, American Chemical Society. (c) Mechanism of the UCNPs@mSiO<sub>2</sub>/ZnPe nanoparticles for fluorescence imaging-guided PDT. Reproduced with permission. [56] Copyright 2019, American Chemical Society.

## 5.2. Photothermal imaging

Photothermal imaging is a sensitive imaging strategy based on the difference of temperature, which is often operated in conjunction with PTT [287, 288]. The core-shell structured nanoparticles that applied in combined therapy of PDT/PTT has the potential for photothermal imaging. For example, Wang et al. incorporated oxygen vacancy-enriched core-shell structured crystalline@amorphous black TiO<sub>2</sub> into a chitosan matrix for synchronous PDT/PTT and photothermal imaging [166]. As monitored by the photothermal images (Fig. 15a), the temperature of the tumor treated with the BT-CTS thermogels rapidly increased and exceeded 50 °C after being irradiated by a 808 nm laser (0.32 W cm<sup>-2</sup>) for 15 min. In the study of Ou et al., zinc porphyrin@PDA nanoparticles were synthesized for photothermal imaging-guided PDT/PTT [289]. Photothermal images demonstrated that the temperature of the tumor injected with zinc porphyrin@PDA nanoparticles quickly rose from 35.0 °C to 52.0 °C after 5 min of 660 nm laser (0.75 W cm<sup>-2</sup>) irradiation, while the temperature of the tumor injected with PBS only increased about 1 °C. Similarly,

882 Huang et al. constructed  $\text{Cu}_{2-x}\text{S}@\text{MnS}$  nanoparticles for photothermal imaging-guided PDT/PTT  
883 [61]. The intense optical absorption of the  $\text{Cu}_{2-x}\text{S}@\text{MnS}$  nanoparticles in NIR region resulted in the  
884 excellent photothermal conversion and photothermal imaging property.

### 885 5.3. Photoacoustic imaging

886 Photoacoustic imaging utilizes the (laser) light pulses to irradiate the sample to generate  
887 ultrasound signals for the images creation. It possesses the advantages of both optical and ultrasonic  
888 imaging, such as high spatial resolution, high optical contrast and deep penetration [290-292].  
889 Recently, core-shell structured nanoparticles with superior absorption properties in the visible or near-  
890 infrared light region have been favored in photoacoustic imaging during PDT-based cancer treatment.  
891 For example, Tan et al. monitored the tumor accumulation behavior of ICG-Ag@PANI nanoparticles  
892 during the PDT/PTT by photoacoustic imaging due to their strong optical absorbance [68]. Compared  
893 with the ICG, PANI and Ag@PANI-treated tumors, the ICG-Ag@PANI-treated tumor exhibited the  
894 strongest photoacoustic signals, which was about 7.9- and 2.5-fold that of ICG and Ag@PANI-treated  
895 tumors, respectively. Besides, Wang et al. reported that the Au@Rh-ICG nanoparticle coated with  
896 tumor cell membrane (CM) could serve as a contrast agent for photoacoustic imaging during PDT  
897 due to its strong NIR absorption [78]. As shown in Fig. 15b, after the intravenous injection of  
898 Au@Rh-ICG-CM nanoparticles, the photoacoustic signals in the tumor region gradually increased  
899 and reached the strongest after 12 h, which was beneficial to trace the tissue distribution of the  
900 nanoparticles and guide the treatment process.

### 901 5.4. Magnetic resonance imaging

902 Magnetic resonance (MR) imaging is a facile and noninvasive imaging technique that offers  
903 evident soft tissue contrast and anatomical details [293, 294]. The relaxation process in nuclear  
904 magnetic resonance can be divided into longitudinal relaxation time ( $T_1$ ) and transverse relaxation

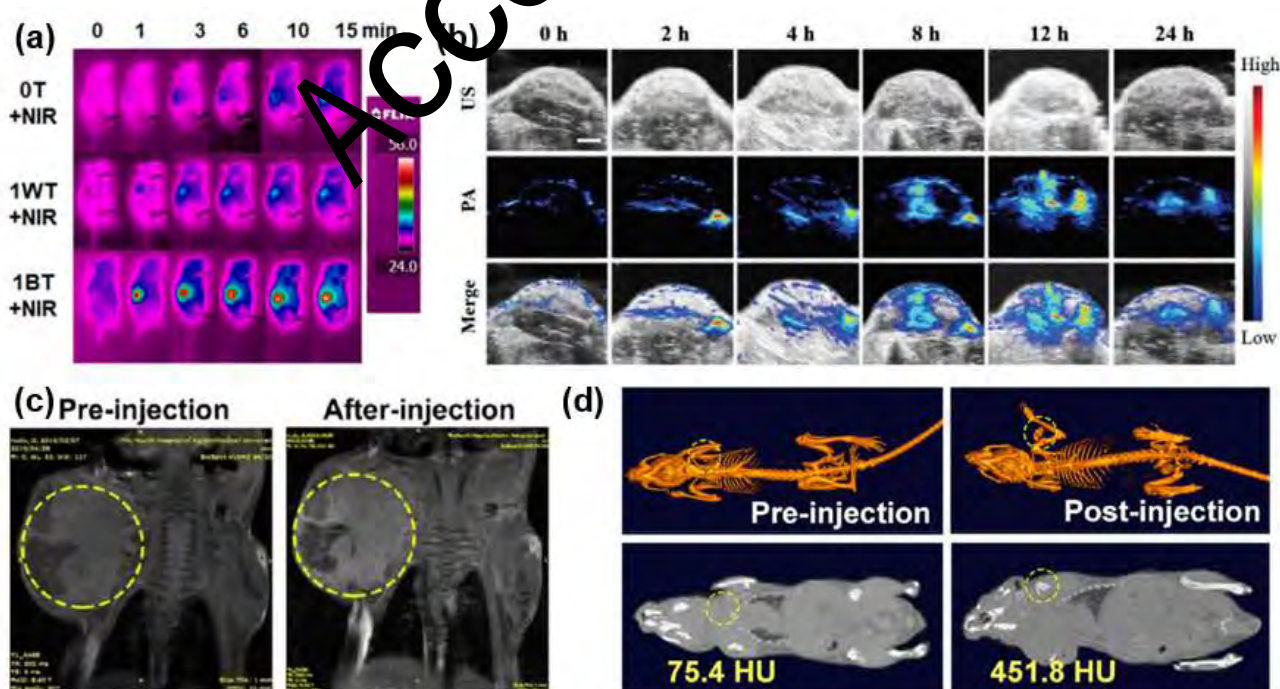
time ( $T_2$ ), both of which can be employed for MR imaging [295]. Commonly, lanthanide ions such as  $Gd^{3+}$  and  $Yb^{3+}$  incorporated in the core-shell structured nanoparticles could enhance the contrast in  $T_1$  MR imaging [188, 296]. For example, Cai et al. fabricated the UZNPs-PAA-DOX nanoparticles for MR imaging-guided PDT/chemotherapy. As exhibited in the MR images of Fig. 15c, there was no obvious difference between the normal tissue and cancerous tissue before the injection of UZNPs-PAA-DOX nanoparticles, while the cancerous tissue presented brighter image than that of the normal tissue after the injection of UZNPs-PAA-DOX nanoparticles [84]. With the increase of  $Yb^{3+}$  ions concentration, the MR signal intensity of UZNPs-PAA-DOX increased gradually. The longitudinal relaxivity of UZNPs-PAA-DOX was estimated to be  $10.36 \text{ mM}^{-1} \text{ s}^{-1}$ , indicating its great potential in  $T_1$  MR imaging. Moreover, owing to the unique MR contrast enhancement effect,  $Fe_3O_4$ -based core-shell structured nanoparticles exhibited excellent performance in  $T_2$  MR imaging [58, 105, 279].

Recently, Mn- and Fe-containing nanomaterials have drawn much attention as tumor microenvironment-enhanced MR contrast agents [207, 297]. In general, the content of GSH,  $H_2O_2$  and  $H^+$  in tumor microenvironment of solid tumor is high. The Mn- and Fe-containing nanomaterials can react with the GSH,  $H_2O_2$  and  $H^+$  in tumor microenvironment to release  $Mn^{2+}$  and  $Fe^{3+}$  ions for enhancing  $T_1$  and  $T_2$  MR signals, respectively. For example, Xu et al. decorated a mesoporous  $MnO_2$  shell on a UCNPs core for tumor microenvironment-enhanced PDT/chemotherapy and multimodal imaging [298]. In tumor microenvironment, the mesoporous  $MnO_2$  shell decomposed rapidly to release  $Mn^{2+}$  ions, which coupled with trimodal imaging of UCNPs to show a self-enhanced imaging. The longitudinal relaxivity of this nanoparticle in PBS was increased from 1.63 ( $pH$  7.4, GSH  $0 \times 10^{-3} \text{ M}$ ,  $H_2O_2$   $0 \times 10^{-6} \text{ M}$ ) to  $9.37 \text{ mM}^{-1} \text{ s}^{-1}$  ( $pH$  6.5, GSH  $10 \times 10^{-3} \text{ M}$ ,  $H_2O_2$   $50 \times 10^{-6} \text{ M}$ ). In the study of Ma et al., the  $Mn^{2+}$  ions released from  $SiO_2$ -MB@ $MnO_2$  nanoparticles due to the decomposition of  $MnO_2$  in acidic tumor microenvironment, which significantly improved the performance of  $T_1$  MR

928 imaging [60].

## 929 5.5. Computed tomography imaging

930 Computed tomography (CT) imaging is an X-ray imaging technique that holds the advantages  
931 of fast acquisition time, high resolution and easy three-dimensional modeling [273]. UCNPs-based  
932 core-shell structured nanoparticles have attracted much interest as CT contrast agents in PDT-based  
933 cancer treatment [59, 91, 149]. In the study of Wang et al., the UCNPs@mSiO<sub>2</sub>-CuS-ZnPc  
934 nanoparticles were fabricated for CT imaging-guided PDT [208]. The tumor site without injection of  
935 UCNPs@mSiO<sub>2</sub>-CuS-ZnPc possessed a CT value of 28.1 Hounsfield Units (HU), which was much  
936 lower than the sample-injected tumor site (313.5 HU). Xu et al. investigated the *in vitro* and *in vivo*  
937 CT contrast imaging properties of the PEG/LDNPs@CMSNs nanoparticles [207]. As the  
938 concentration of PEG/LDNPs@CMSNs nanoparticles increased, the CT signal intensity increased  
939 rapidly. As shown in Fig. 15d, the CT value of the tumor site with injection of PEG/LDNPs@CMSNs  
940 was 451.8 HU, which was significantly higher than that of the control group (75.4 HU), indicating  
941 that the PEG/LDNPs@CMSNs was a promising CT imaging contrast agent.



942

**Fig. 15.** (a) Infrared thermal images of B16F10 tumor-bearing mice treated with BT-CTS thermogels under the NIR irradiation. Reproduced with permission. [166] Copyright 2019, American Chemical Society. (b) Photoacoustic images of Au@Rh-ICG-CM nanoparticles at the tumor site. Reproduced with permission. [78] Copyright 2020, Wiley-VCH. (c) MR images of mice before and after administration UZNPs-PAA-DOX nanoparticles. Reproduced with permission. [84] Copyright 2020, American Chemical Society. (d) CT images of tumor-bearing mice by pre- and postinjection of PEG/LDNPs@CMSNs nanoparticles. Reproduced with permission. [207] Copyright 2020, American Chemical Society.

## 6. Conclusions and perspectives

In summary, core-shell structured nanoparticles are promising multifunctional nanoplateforms for PDT-based cancer treatment and related imaging. These nanoparticles are divided into three categories: inorganic, organic and hybrid on the basis of material compositions of the core and shell. During PDT of cancer, the core-shell structured nanoparticles serve as photosensitizer delivery vehicles, energy transducers, photosensitizers and hypoxic tumor microenvironment modulators to improve the therapeutic efficacy. The combination of PDT with chemotherapy, PTT, immunotherapy and other therapies resolves some challenging issues for monotherapy, involving the metastasis of tumors and the development of resistance. **Moreover, these nanoparticles possess excellent imaging performance in PDT-based cancer treatment.**

Despite considerable progress has been made, the core-shell structured nanoparticles are still far from the clinical application of PDT-based cancer treatment and related imaging. From the perspective of materials science, the synthetic steps of most core-shell structured nanoparticles are complex, which easily causes material differences between different batches, and will bring difficulties to the expansion of production and commercialization. Consequently, it is necessary to develop facile synthetic strategies for safely and quickly preparing core-shell structured nanoparticles.



966 Besides, the component of core-shell structured nanoparticles should be optimized for further  
967 improving the targeting ability, therapeutic efficacy and stability.

968 From the perspective of biology, the knowledge of cancers still exists deficiencies because of  
969 the limitations of modern technology and the complexity of biosystem. The metastasis of tumors and  
970 the development of resistance have always been troubles to be solved urgently for PDT and other  
971 therapies. Although many core-shell structured nanoparticles are effective in PDT-based cancer  
972 treatment, their systemic cytotoxicity and long-term human toxicity need more comprehensive and  
973 in-depth investigation. Meanwhile, the dosage of core-shell structured nanoparticles and light source  
974 parameters should be controlled to achieve precise treatment and alleviate side effects. In addition,  
975 bacterial infections are one of the inducing factors of cancer and has become increasingly serious  
976 with the rise of antibiotic resistance. PDT is a promising strategy to control the bacterial infections,  
977 and it is of great significance to develop novel and efficient core-shell structured nanoparticles for  
978 antibacterial PDT.

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